# Effect of High Glucose Concentrations on Expression of ELAM-1, VCAM-1 and ICAM-1 in HUVEC with and without Cytokine Activation

# TS. ALTANNAVCH<sup>1</sup>, K. ROUBALOVÁ<sup>2</sup>, P. KUČERA<sup>1</sup>, M. ANDĚL<sup>1</sup>

<sup>1</sup>Second Department of Internal Medicine, Third Faculty of Medicine, Charles University, <sup>2</sup>National Institute of Public Health, Prague, Czech Republic

Received June 6, 2002 Accepted March 10, 2003

### Summary

Diabetes mellitus is associated with an increased prevalence of endothelial dysfunction and development of atherosclerotic vascular diseases. We demonstrate here that hyperglycemia results in the expression of adhesion molecules on endothelial cells *in vitro*. Incubation of human umbilical vein endothelial cells (HUVEC) in a culture medium with 11.0 mM, 16.5 mM and 22.0 mM glucose concentrations induced the expression of intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1) and endothelial-leukocyte adhesion molecule-1 (ELAM-1). This effect was detectable after 24 h incubation of HUVEC with a high glucose concentration. The effect of high glucose concentration on TNF- $\alpha$  induced expression of ELAM-1, VCAM-1 and ICAM-1 was negligible, if at all. These results show that even a short-term exposure of endothelial cells (ECs) to high glucose concentration leads to their activation associated with increased expression of adhesion molecules such as ELAM-1, VCAM-1 and ICAM-1.

### Key words

Adhesion molecules • Atherosclerosis • Endothelial cells • Diabetes mellitus • High glucose

# Introduction

Prevalence of the atherosclerotic vascular disease is markedly increased among individuals with diabetes mellitus (Kannel and McGee 1979, Jang *et al.* 1998). Recent evidence has suggested that endothelial dysfunction, a proposed risk factor for atherosclerosis, plays a key role in the pathogenesis of diabetic atherosclerotic cardiovascular disease (Haffner *et al.* 1998).

Leukocyte adhesion to arterial endothelial cells is thought to be an important step in the development of atherosclerosis (Carter and Grant 1997). Adhesion molecules such as intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1) and the endothelial-leukocyte adhesion molecule-1 (ELAM-1) play an essential role in this step (Ross 1999). Although the exact mechanisms of diabetes-induced endothelial dysfunction are unknown, pathological elevation of glucose levels has been shown to induce various effects at the cell level either directly (Tesfamariam *et al.* 1990) or through the formation of intermediate products such as advanced glycation end-product (AGE) adducts (Esposito *et al.* 1989). It has been reported that exposure of the vascular endothelium to elevated glucose concentration induces expression of

# PHYSIOLOGICAL RESEARCH

ICAM-1 *in vitro* (Baumgartner-Parzer *et al.* 1995, Takami *et al.* 1998). Furthermore, Otsuki *et al.* (1997) and Matsumoto *et al.* (2000) reported that serum concentrations of soluble adhesion molecules (sICAM-1, sVCAM-1, sELAM-1) are elevated in patients with type 2 diabetes. An increased expression of endothelial cell adhesion molecules in response to glucose has also been correlated with an alteration of nitric oxide (NO) synthesis (Sobrevia and Mann 1997) and endotheliuminduced vasodilatation (Williams *et al.* 1998).

In this study, we have investigated the effects of elevated glucose concentrations on the expression of adhesion molecules (ELAM-1, VCAM-1 and ICAM-1) on basal and cytokine (TNF- $\alpha$ ) stimulated human umbilical vein endothelial cells (HUVEC).

### Methods

#### Cell cultures

Human umbilical cords were obtained from normal placentas, excised after birth (kindly provided by the Department of Obstetrics and Gynecology of the University Hospital Královské Vinohrady, Praha) and placed in a sterile container filled with a transfer buffer. HUVEC were isolated and cultured by a modified method according to Jaffe et al. (1973) and Marin et al. (2001). Briefly, primary endothelial cells (ECs) were harvested from umbilical cord veins treated with 0.2 % collagenase (Gibco, UK) and incubated for 25 min at 37 °C and 5 % CO<sub>2</sub>. After incubation, the collagenase solution containing HUVEC was flushed from the cord by perfusion with 20 ml of Medium 199 (M 199, BioWhittaker, USA). The cells were centrifuged for 5 min at 1500 rpm, the medium was discarded, and resuspended in 5 ml of fresh culture medium. The cell suspension cultured on gelatin-coated tissue culture flasks (Greiner GmbH) in M 199 containing 20 % fetal bovine serum (FCS, Gibco, BRL, Paisley, UK), 20 ng/ml fibroblast growth factor, 10 ng/ml epidermal growth factor (both from Gibco, BRL, Paisley, UK), penicillin 100 U/ml + streptomycin 0.1 mg/l (Sevapharma, Czech Republic), gentamycin 25 µg/ml (Gibco, UK) and amphotericin 2.5 mg/ml (Gibco, UK). The cultures were maintained at 37 °C and 5 % CO<sub>2</sub> and passaged once or twice a week by treating confluent monolayer with 0.125 % trypsin, washing the cell suspension with phosphate buffered saline (PBS) and resuspending the cells in doubled volume of fresh medium. Isolated ECs were identified by indirect immunofluorescence using monoclonal antibody anti-CD31, Human Endothelial Cell

(Dako, Denmark). For all experiments, endothelial cell cultures were used at passages 1-4.

# Incubation of HUVEC with various concentrations of glucose

Three to five days old confluent cultures of HUVEC were cultivated in culture medium containing 11.0 mM, 16.5 mM, or 22.0 mM glucose concentrations, respectively. The cells cultivated in a culture medium with a physiological glucose concentration (5.5 mM) were used as controls. After 24 h incubation with glucose, a portion of ECs cultures were treated with 10 ng/ml TNF- $\alpha$  (Sigma, Aldrich, USA). At 4 h (ELAM-1), 8 h (VCAM-1) and 12 h (ICAM-1) after TNF- $\alpha$  induction, samples from both TNF- $\alpha$  treated and untreated cultures were taken for measuring of adhesion molecule expression by flow-cytometry.

### Measurement of adhesion molecules expression by flowcytometry

The ECs cultures were washed with PBS, detached using ice-cold PBS and 0.005 % EDTA, thoroughly resuspended and incubated with fluorescent label-marked monoclonal antibodies anti-ICAM-1, anti-VCAM-1 (both from B.D. Pharmingen, San Diego) and anti-ELAM-1 (Bender, Vienna, Austria) for 15 min at 37 °C. A minimum of 5 x  $10^3$  cells were analyzed by fluorescence-activated flow-cytometry (FACS) on ORTHO-Diagnostic systems FACS-Cytoronabsolute (Johnson & Johnson).

#### Statistical analysis

The results were evaluated using methods of variation analysis (ANOVA test). The data are given as mean values  $\pm$  S.D. P $\leq$ 0.05 value was considered statistically significant.

# Results

The effect of high glucose concentrations on spontaneous and TNF- $\alpha$  induced expression of ELAM-1, VCAM-1 and ICAM-1 in cultured HUVEC is shown in Table 1. Cultivation of HUVEC for 24 h in a high glucose concentration stimulated an expression of all adhesion molecules studied (ELAM-1, VCAM-1 and ICAM-1). From the glucose concentrations tested, 16.5 mM glucose had the most pronounced effect on adhesion molecules expression. At this concentration, the expression of ELAM-1 was 6-fold increased above the baseline level found in the control culture (p=0.006).

VCAM-1 was increased 7-fold (p=0.003) and ICAM-1 two-fold (p=0.05), respectively. However, even a smaller elevation of glucose concentration (11.0 mM) stimulated the expression of ELAM-1 and VCAM-1 significantly. At very high glucose concentration (22.0 mM), the rate of adhesion molecule induction was not as pronounced as at 11.0 mM and 16.5 mM concentrations. This may be partially due to toxicity of this glucose concentration for HUVEC.

As was expected, TNF- $\alpha$  per se stimulated significantly the expression of ELAM-1, VCAM-1 and ICAM-1. The effect of high glucose on TNF- $\alpha$  induced expression of these adhesion molecules was negligible, if any (Figs 1-3).

**Table 1.** The expression of ELAM-1, VCAM-1 and ICAM-1 in cultured HUVEC following the exposure to 5.5 mM (control), 11.0 mM, 16.5 mM and 22.0 mM glucose with and without the induction by TNF- $\alpha$ .

		% of positive cells						
Adhesion molecules		5.5 mM glucose (control)	11.0 mM glucose	P value vs control	16.5 mM glucose	P value vs control	22.0 mM glucose	P value vs control
ELAM	TNF-α (–) TNF-α (+)	6.5±3.8 72±1.9	30.8±7.7 74±2.5	p=0.003	42.1±9.5 81±3.5	p=0.006	22.5±7.8 71±4.6	p=0.007
VCAM-1	TNF- $\alpha$ (-) TNF- $\alpha$ (+)	4.0±3.3 46±8.2	21.4±8.5 62±5.2	p=0.006	28±10 79±2.1	p=0.003	16.1±6.1 53±9.8	p<0.01
ICAM-1	TNF-α (-) TNF-α (+)	12.4±5.1 87±9.3	18.8±7.3 83±1.5	p=NS	25.2±10.2 88±1.5	p<0.05	$16.8\pm 5.8$ $83\pm 6.2$	p=NS

Values are mean  $\pm$  S.D. of four separate experiments; TNF- $\alpha$  (–) expression of adhesion molecules without induction by TNF- $\alpha$ , TNF- $\alpha$  (+) expression of adhesion molecules after induction by TNF- $\alpha$ .





**Fig 1.** Effect of high glucose concentrations on expression of ELAM-1 in HUVEC with (full bars) and without (open bars) induction by TNF- $\alpha$ . Endothelial cell monolayer were treated for 24 h with various glucose concentrations in Medium 199 supplemented with 20 % FCS and the cell surface expression of ELAM-1 was measured by flow-cytometry.

Fig 2. Effect of high glucose concentrations on expression of VCAM-1 in HUVEC with (full bars) and without (open bars) induction by TNF- $\alpha$ . After 24 h incubation with various glucose concentrations expression of VCAM-1 was measured by flow-cytometry.



Fig 3. Effect of high glucose concentrations on expression of ICAM-1 in HUVEC with (full bars) and without (open bars) induction by TNF- $\alpha$ . After 24 h incubation with various glucose concentrations expression of VCAM-1 was measured by flow-cytometry.

### Discussion

We have demonstrated in the present study that short-term exposure of HUVEC to high glucose concentrations results in the enhanced expression of surface ELAM-1, VCAM-1 and ICAM-1. This stimulative effect of high glucose concentration on the expression of adhesion molecules by endothelial cells has been previously reported in various studies performed both in vivo and in vitro. However, the results concerning the rate of induction of various types of adhesion molecules have not been consistent depending on the experimental system used. Stimulation of ICAM-1 expression by high glucose concentration was observed most often (Baumgartner-Parzer et al. 1995, Taki et al. 1996, Takami et al. 1998, Kado et al. 2001). Recently, the activation of VCAM-1 was demonstrated by Esposito et al. (2001) in aortic endothelial cells adapted by longterm cultivation at high glucose concentrations. Puente-Navazo et al. (2000) who exposed HUVEC for short periods of time to high glucose concentrations, observed activation of ICAM-1 and P-selectin, but not ELAM-1 and VCAM-1.

One of the main findings of this study was that short-term exposure of ECs to high glucose resulted in the induction of all three types of adhesion molecules on HUVEC *in vitro*. These results correspond to the findings of Marfella *et al.* (2000) and Matsumoto *et al.* (2002) who documented that hyperglycemia increases circulating ICAM-1 and VCAM-1 in type 2 diabetic patients. Moreover, increased plasma concentrations of these adhesion molecules were associated with insulin concentrations and insulin resistance (Bluher *et al.* 2002).

It has been documented that high glucose can influence various physiological processes in many cell types (Danne *et al.* 1993, Haneda *et al.* 1997). However, from the experiments *in vivo* it is not clear whether the effect of high glucose concentration is direct or indirect. For example, the indirect effect of oxidative stress and free radical formation is responsible for the induction of adhesion molecule expression.

In our experiments, we have demonstrated that the induction of adhesion molecules can reflect direct effects of glucose on ECs in culture. Three different elevated concentrations of glucose were tested in order to eliminate potential toxic effect of high glucose concentration on HUVEC. At 11.0 mM and 16.5 mM glucose neither cell viability nor proliferation was influenced. At 22.0 mM concentration, cell viability was not influenced, but cell doubling was inhibited (results not shown) There are several mechanisms by which high glucose can participate in endothelial dysfunction (Nagel et al. 1994, Ceriello 1997, Cominacini et al. 1997). Some studies described that high glucose content can activate a signaling pathway mediated by protein kinase C (PKC) (Inoguchi et al. 1994, Williams et al. 1997, Koya and King 1998). The activation of PKC results in the increased transcription of various genes including those for adhesion molecules. Upregulation of mRNA for ICAM-1 in the presence of high glucose concentration was documented by Kado et al. (2001). This hypothesis is further supported by the findings these authors and Booth et al. (2002) that PKC inhibitors (staurosporine, bisindolylmaleimide-1) attenuated stimulatory effect of high glucose concentrations on ICAM-1 expression (Kado et al. 2001, Booth et al. 2002). Another mechanism involving AGE adducts was reported by Esposito et al. (2001) who demonstrated that AGE are important mediators of endothelial dysfunction induced by a long-term exposure of ECs to high glucose.

The induction of adhesion molecules on the surface of ECs is one of the first steps in high glucosemediated endothelial dysfunction in diabetic patients. Together with other mechanisms, e.g. inhibition of NO synthesis (Takahashi *et al.* 1996) resulting in vasoconstriction and endothelial impairment due to the free radical formation (Kashiwagi *et al.* 1996), it contributes to the development of inflammation in vascular wall and finally to vascular pathological changes.

### Acknowledgements

This study was supported by the Research Program of the Third Faculty of Medicine, Charles University Prague, for the project "*The prevention, diagnosis and treatment* of initial studies of diabetes mellitus, endocrine, metabolic and toxic disorders" (FB MSM 111200001).

# References

- BAUMGARTNER-PARZER SM, WAGNER L, PETTERMANN M, GESSL A, WALDHAUSL W: Modulation by high glucose of adhesion molecules expression in cultured endothelial cells. *Diabetologia* **38**: 1367-1370, 1995.
- BLUHER M, UNGER R, RASSOUL F, RICHTER V, PASCHKE R: Relation between glycaemic control, hyperinsulinaemia and plasma concentrations of soluble adhesion molecules in patients with impaired glucose tolerance or Type II diabetes. *Diabetologia* **45**: 210-216, 2002.
- BOOTH G, STALKER TJ, LEFER AM, SCALIA R: Mechanisms of amelioration of glucose-induced endothelial dysfunction following inhibition of protein kinase C in vivo. *Diabetes* **51**: 1556-1564, 2002.
- CARTER AM, GRANT PJ: Vascular homeostasis, adhesion molecules, and macrovascular disease in non-insulindependent diabetes mellitus. *Diabet Med* 14: 423-432, 1997.
- CERIELLO A: Acute hyperglycemia and oxidative stress generation. Diabet Med 14 (Suppl 3): S45-S49, 1997.
- COMINACINI L, FRATTA PASINI A, GARBIN U, CAMPAGNOLA M, DAVOLI A, RIGONI A, ZENTI MG, PASTORINO AM, LO CASCIO V: E-selectin plasma concentration is influenced by glycemic control in NIDDM patients: possible role of oxidative stress. *Diabetologia* 40: 584-589, 1997.
- DANNE T, SPIRO MJ, SPIRO RG: Effect of high glucose on type IV collagen production by cultivated glomerular, epithelial, endothelial and mesangial cells. *Diabetes* **42**: 170-177, 1993.
- ESPOSITO C, GERLACH H, BRETT J, STERN D, VLASSARA H: Endothelial receptor-mediated binding of glucosemodified albumin is associated with increased monolayer permeability and modulation of cell surface coagulant properties. J Exp Med 170: 1387-1407, 1989.
- ESPOSITO C, FASOLI G, PLATI AR, BELLOTTI N, CONTE MM, CORNACCHIA F, FOSCHI A, MAZZULLO T, SEMERARO L, DAL CANTON A: Long-term exposure to high glucose up-regulates VCAM-1-induced endothelial cell adhesiveness to PBMC. *Kidney Int* **50**: 1842-1849, 2001.
- HAFFNER SM, LEHTO S, RONNEMAA T, PYORALA K, LAAKSO M: Mortality from coronary heart disease in subjects with type 2 diabetes and in nondiabetic subjects with and without prior myocardial infarction. *N Engl J Med* **339**: 229-234, 1998.
- HANEDA M, ARAKI S, TOGAWA M, SUGIMOTO T, ISONO M, KIKKAWA R: Activation of mitogen-activated protein cells cultured under high glucose conditions. *Kidney Int* **51** (Suppl 60): S66-S69, 1997.
- INOGUCHI T, XIA P, KUNISAKI M, HIGASHI S, FEENER EP, KING GL: Insulin's effect on protein kinase C and diacylglycerol induced by diabetes and glucose in vascular tissues. *Am J Physiol* **267**: E369-E379, 1994.
- JAFFE EA, NACHMAN RL, BECKER CG, MINICK CR: Culture of human endothelial cells derived from umbilical veins. Identification by morphologic and immunologic criteria. *J Clin Invest* **52**: 2745-2756, 1973.
- JANG Y, LINCOFF AM, PLOW EF, TOPOL EJ: Cell adhesion molecules in coronary artery disease. J Am Coll Cardiol 24: 1591-1601, 1998.
- KADO S, WAKATSUKI T, YAMAMOTO M, NAGATA N: Expression of ICAM-1 induced by high glucose concentrations in human aortic endothelial cells. *Life Sci* 68: 727-737, 2001.
- KANNEL WB, MCGEE DL: Diabetes and cardiovascular disease. The Framingham study. JAMA 241: 2035-2038, 1979.
- KASHIWAGI A, ASAHINA T, NISHIO Y, IKEBUCHI M, TANAKA Y, KIKKAWA R, SHIGETA Y: Glycation, oxidative stress, and scavenger activity: glucose metabolism and radical scavenger dysfunction in endothelial cells. *Diabetes* **45** (Suppl 3): S84-S86, 1996.
- KOYA D, KING G: Protein kinase C activation and the development of diabetic complications. *Diabetes* **47**: 859-866, 1998.

- MARFELLA R, ESPOSITO K, GIUNTA R, COPPOLA G, DE ANGELIS L, FARZATI B, PAOLISSO G, GIUGLIANO D: Circulating adhesion molecules in humans. Role of hyperglycemia and hyperinsulinemia. *Circulation* **101**: 2247-2251, 2000.
- MARIN V, KAPLANSKI G, GRES S, FARNARIER C, BONGRAND P: Endothelial cell culture: protocol to obtain and cultivate human umbilical endothelial cells. *J Immunol Methods* **254**: 183-190, 2001.
- MATSUMOTO K, MIYAKE S, YANO M, UEKI Y, TOMINAGA Y: High serum concentrations of soluble E-selectin in patients with impaired glucose tolerance hyperinsulinemia. *Atherosclerosis* **152**: 415-420, 2000.
- MATSUMOTO K, SERA Y, NAKAMURA H, UEKI Y, MIYAKE S: Serum concentrations of soluble adhesion molecules are related to degree of hyperglycemia and insulin resistance in patients with type 2 diabetes mellitus. *Diabetes Res Clin Pract* **55**: 131-138, 2002.
- NAGEL T, RESNICK N, ATKINSON WJ, DEWEY CF Jr, GIMBRONE MA Jr: Shear stress selectively upregulates ICAM-1 expression in cultured human vascular endothelial cells. *J Clin Invest* **94**: 885-891, 1994.
- OTSUKI M, HASHIMOTO K, MORIMOTO Y, KISHIMOTO T, KASAYAMA S: Circulating vascular cell adhesion molecule-1 (VCAM-1) in atherosclerotic NIDDM patients. *Diabetes* **46**: 2096-2101, 1997.
- PUENTE NAVAZO MD, CHETTAB K, DUHAULT J, KOENIG-BERARD E, McGREGOR JL: Glucose and insulin modulate the capacity of endothelial cells (HUVEC) to express P-selectin and bind a monocytic cell line (U937). *Thromb Haemost* **86**: 680-685, 2001.
- ROSS R: Atherosclerosis an inflammatory disease. N Engl J Med 340: 115-126, 1999.
- SOBREVIA L, MANN GE: Dysfunction of the endothelial nitric oxide signalling pathway in diabetes and hyperglycemia. *Exp Physiol* 82: 423-452, 1997.
- TAKI H, KASHIWAGI A, TANAKA Y, HORIIKE K: Expression of ICAM-1 via osmotic effect in human umbilical vein endothelial cells exposed to high glucose medium. *Life Sci* **58**: 1713-1721, 1996.
- TAKAHASHI M, IKEDA U, MASUYAMA J, FUNAYAMA H, KANO S, SHIMADA K: Nitric oxide attenuates adhesion molecule expression in human endothelial cells. *Cytokine* **8**: 817-821, 1996.
- TAKAMI S, YAMASHITA S, KIHARA S, KAMEDA-TAKEMURA K, MATSUZAWA Y: High concentration of glucose induces the expression of intercellular adhesion molecule-1 in human umbilical vein endothelial cells. *Atherosclerosis* **38**: 35-41, 1998.
- TESFAMARIAM B, BROWN ML, DEYKIN D, COHEN RA: Elevated glucose promotes generation of endotheliumderived vasoconstrictor prostanoids in rabbit aorta. *J Clin Invest* **85**: 929-932, 1990.
- WILLIAMS B, GALLACHER B, PATEL H, ORME C: Glucose-induced protein kinase C activation regulates vascular permeability factor mRNA expression and peptide production by human vascular smooth muscle cells in vitro. *Diabetes* 46: 1497-1503. 1997.
- WILLIAMS SB, GOLDFINE AB, TIMIMI FK, TING HH, RODDY MA, SIMONSON DC, CREAGER MA: Acute hyperglycemia attenuates endothelium-dependent vasodilation in humans in vivo. *Circulation* **97**: 1695-1701, 1998.

# **Reprint requests**

Prof. Michal Anděl, Second Department of Internal Medicine, Third Faculty of Medicine, Charles University, Šrobárova 50, 100 34 Prague 10, Czech Republic, e-mail: michal.andel@lf3.cuni.cz