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MINIREVIEW

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## ATP-Binding Cassette (ABC) Transporters in Human Metabolism and Diseases

J. ŠTEFKOVÁ, R. POLEDNE, J. A. HUBÁČEK

*Institute for Clinical and Experimental Medicine, Laboratory for Atherosclerosis Research, Center for Experimental Cardiovascular Research, Prague, Czech Republic*

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### Summary

The ATP-binding cassette (ABC) superfamily of active transporters involves a large number of functionally diverse transmembrane proteins. They transport a variety of substrates including amino acids, lipids, inorganic ions, peptides, saccharides, metals, drugs, and proteins. The ABC transporters not only move a variety of substrates into and out of the cell, but also are also involved in intracellular compartmental transport. Energy derived from the hydrolysis of ATP is used to transport the substrate across the membrane against a concentration gradient. The typical ABC transporter consists of two transmembrane domains and two nucleotide-binding domains. Defects in 14 of these transporters cause 13 genetic diseases (cystic fibrosis, Stargardt disease, adrenoleukodystrophy, Tangier disease, etc.). Mutations in three genes affect lipid levels expressively. Mutations in ABCA1 cause severe HDL deficiency syndromes called Tangier disease and familial high-density lipoprotein deficiency, which are characterized by a severe deficiency or absence of high-density lipoprotein in the plasma. Two other ABCG transporters, ABCG5 and ABCG8, mutations of which cause sitosterolemia, have been identified. The affected individuals absorb and retain plant sterols, as well as shellfish sterols.

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### Key words

ABC transporter • Lipid metabolism • Cholesterol • Mutation

### Introduction

The ATP-binding cassette (ABC) superfamily of active transporters is composed of about 50 functionally diverse prokaryotic and eukaryotic transmembrane proteins (Higgins 1992, Michaelis and Berkower 1995). These proteins are fundamental to membrane transport of a broad variety of substrates including amino acids, lipids, lipopolysaccharides, anorganic ions, peptides, saccharides, metals, drugs and proteins (Higgins 1992).

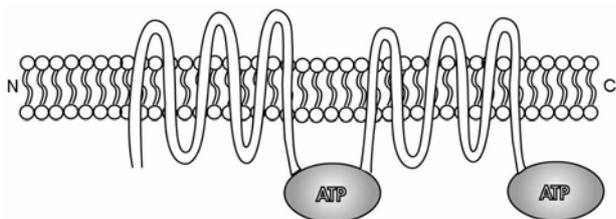
The ABC transporters not only move a variety of

substrates into and out of the cell, but also are involved in intracellular compartmental transport. These proteins utilize energy derived from the hydrolysis of ATP to transport the substrate across the membrane against a concentration gradient. It has been estimated that the hydrolysis of two ATP molecules results in the transport of one molecule of substrate. Substrate specificity for each transporter is determined by the amino acid sequence in the transmembrane domain (TMD). As a result, there is usually little similarity between the transmembrane domains of different ABC transporters.

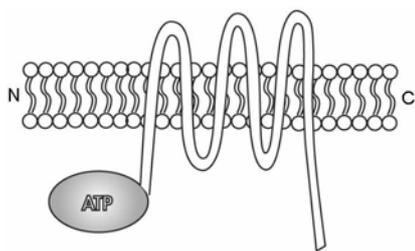
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Even if sequence similarity is noted, substrate similarity is not implied.

The typical ABC transporter consists of two transmembrane domains and two nucleotide-binding domains (NBDs) encoded by a single polypeptide (Fig. 1). ABC transporters may also represent a multi-component unit, in which a different gene encodes for each domain or half molecule (i.e. one TMD and one NBD, Fig. 2) (Dean and Allikmets 1995, Higgins 1995). The TMD contains 6-11 membrane-spanning  $\alpha$ -helices. In contrast, the hydrophilic NBDs of the ABC transporters are highly conserved. The amino acid identity in these regions varies from 30 to 50 % between transporters. NBDs consist of approximately 90-110 amino acids. Each nucleotide-binding domain contains two sequence motifs, the Walker A and Walker B motifs, which are common to the general category of nucleotide-binding proteins. The distinctive feature of all ABC transporters is the C motif that has the consensus sequence "LeuSerGlyGlyGln" (Hyde *et al.* 1990).



**Fig. 1.** Full ABC transporter consists of two transmembrane domains and two nucleotide binding domains encoded by a single polypeptide.



**Fig. 2.** Half ABC transporter consists of one transmembrane domain and one nucleotide binding domain.

Some ABC transporters are not directly involved in transporting substrates, but appear to be part of ion channels (such as the cystic fibrosis transmembrane regulator or the sulfonyleurea regulator). In these cases, it appears that the hydrolysis of ATP is linked to movement of the protein itself (for cystic fibrosis transmembrane regulator) or to other proteins (for

sulfonyleurea regulator) to regulate opening and closing of ion channels (Obmolova *et al.* 2000).

Many ABC transporters are major determinants of human disease. Among the first ABC transporters to be identified in man was the P-glycoprotein (Pgp) multigene family (Juliano and Ling 1976, Juranka *et al.* 1989, Endicott and Ling 1989) of which Pgp1 plays a critical role in drug resistance in human cancers.

At present, there are approximately 50 known ABC transporters in human. Phylogenetic analysis is used to divide ABC transporters into seven distinct subfamilies of proteins. The subfamilies are named ABCA-ABCG.

Defects in 14 of these transporters cause 13 genetic diseases, the most common of which are cystic fibrosis, Stargardt disease, adrenoleukodystrophy, and Tangier disease (Table 1).

## Human ABC gene subfamilies participating in lipid transport

### ABCA subfamily

This is a unique group that, in contrast to any other human ABC transporters, lacks a structural counterpart in yeast. The structural hallmark of the ABCA subfamily is the presence of a stretch of hydrophobic amino acids thought to span the membrane within the putative regulatory domain. Up to now, 12 ABCA-encoding genes have been identified, but only four ABCA transporters have been fully characterized. Full transporters are further divided into two subgroups based on phylogenetic analysis and intron structure (Luciani *et al.* 1994). The first group includes seven genes dispersed on six different chromosomes, ABCA1 (9q22-q31), ABCA2 (9q34), ABCA3 (16p13,3), ABCA4 (1p22), ABCA7 (19p13,3), ABCA11 (4p16), ABCA12 (2q35), whereas the second group contains five genes (ABCA5, ABCA6, ABCA8, ABCA9 and ABCA10) arranged in a cluster on chromosome 17q24 (Klugbauer and Hofman 1996, Allikmets 1997, Azarian and Travis 1997).

A pairwise comparison of the published full nucleotide sequences for ABCA cDNAs shows an overall identity of around 60 % among the different genes, irrespectively of the pairs analyzed. Cross-species conservation of individual genes is extremely high, exceeding 85 % identity (Luciani *et al.* 1994).

The gatekeeper of the reverse cholesterol transport pathway is an ATP-binding cassette called ABCA1, a 2261 amino acid integral membrane protein

(Rust *et al.* 1999, Oram and Vaughan 2000, Santamarina-Fojo *et al.* 2001). ABCA1 transports cellular cholesterol and phospholipids (mostly phosphatidylcholine) to cell surface-bound apolipoproteins (Oram and Yokoyama 1996, Oram 2000). This protein therefore represents the first and rate-controlling step in the reverse cholesterol transport pathway. Our knowledge of ABCA1 functioning is still insufficient. Although the exact structure of ABCA1 is unknown, electron microscopic analysis has suggested a structural model for a closely related ABC transporter called P-glycoprotein (Rosenberg *et al.* 1997). This model predicts that the two membrane-spanning domains form a large aqueous chamber in the plasma membrane that opens through pores on the cell surface and within the membrane lipid phase. Because the chamber does not open to the cytosol, P-glycoprotein is believed to act as a "floppase" by translocating lipophilic compounds from the inner aspect of the plasma membrane into the aqueous environment of the ABC transporter chamber. Both the cell-surface and inner membrane pores are estimated to be relatively large, allowing for simultaneous translocation of multiple molecules. This may explain why the ABCA1 pathway generates diffuse and irregular structures that protrude from the plasma membrane and interact with apolipoproteins (Lin and Oram 2000).

Because cholesterol is an integral and necessary membrane component, it is likely that ABCA1 targets specific pools of excess abundant cholesterol molecules for secretion. It is still not clear what mechanisms are involved in this process. However, some studies have shown that the interaction of apolipoproteins with cholesterol-loaded cells stimulates translocation of free cholesterol away from intracellular esterifying enzymes to sites accessible to apolipoproteins (Slatte *et al.* 1987, Oram *et al.* 1991, Rogler *et al.* 1995) that presumably contain ABCA1. The properties of these ABCA1 membrane domains are unknown, but they differ from cholesterol- and phospholipid-rich rafts that contain caveolae (Mendez *et al.* 2000). As one possible mechanism, the interaction of apolipoproteins with ABCA1 or a partner protein might stimulate translocation of intracellular cholesterol and phospholipids from the Golgi to plasma membrane ABCA1 by a signal-responsive vesicular transport pathway. Another possibility could be that ABCA1-containing vesicles travel to intracellular lipid deposits, ABCA1 pumps lipids into the vesicle lumen, and the vesicles transport the lipid cargo back into the plasma membrane (Takahashi and

Smith 1999).

ABCA2, ABCA3, and ABCA7 mRNA levels have been reported to be upregulated by the sustained cholesterol influx mediated by modified low-density lipoprotein (Kaminski *et al.* 2001). ABCA3 may play an important role in the formation of pulmonary surfactant, which is composed mainly of phospholipids and specific surfactant proteins (Johansson and Crustedt 1997).

ABCA4 is proposed to be transmembrane transporters for N-retinylidene phosphatidylethanolamin. ABCA4 protein transports vitamin A derivatives in the outer segments of photoreceptor cells and therefore performs a crucial step in the visual.

ABCA transporters also play an important role in genetic diseases. Mutations in ABCA1 cause a severe HDL deficiency syndrome called Tangier disease (Hoffman *et al.* 1965). Tangier disease is a rare autosomal recessive disorder characterized by a severe deficiency or absence of high-density lipoprotein in plasma and accumulation of cholesterol esters in macrophages and other reticuloendothelial cells of tissues including tonsils, thymus, lymph node, bone marrow, spleen, liver, gallbladder, and intestinal mucosa. Many patients also have lipid deposits in neuronal Schwann cells, smooth muscle cells, and fibroblasts. Lipid-free apolipoproteins are unable to remove cholesterol and phospholipids from fibroblasts isolated from Tangier disease patients, which is consistent with a defective ABCA1. Characteristic features of Tangier disease include large yellow-orange tonsils, neuropathies, splenomegaly, hepatomegaly, ocular abnormalities, hyper-cholesterolemia and cardiovascular disease (Assmann *et al.* 1995). Familial high-density lipoprotein deficiency had features of an autosomal dominant disorder characterized by clear manifestations of low HDL in the heterozygotes, with no obvious features of cholesterol ester deposition in cells. Genetic analysis has revealed that Tangier disease is due to mutations on both alleles of ABCA1, whereas familial high-density lipoprotein deficiency is due to a mutation on a single allele (Brooks-Wilson *et al.* 1999, Rust *et al.* 1999, Bodzioch *et al.* 1999).

It has been shown that ABCA1 polymorphisms also plays a role in the plasma lipid levels. Three novel polymorphisms were identified in the promoter region of the ABCA1 gene, and associations of the common polymorphisms with the plasma levels of lipids, angiographic indices of the severity, progression, and regression of coronary atherosclerosis were assumed

(Lutucuta *et al.* 2001). The -477C/T variant was strongly associated with the severity of coronary atherosclerosis and modestly associated with the plasma levels of HDL

cholesterol and ApoAI, whereas the other two polymorphisms were not associated with plasma lipid levels.

**Table 1.** List of human ABC transporters, their functions and diseases caused by *ABC* genes

Transporter name	Function	Disease
<i>ABCA1</i>	Cholesterol and phospholipids transport	Tangier disease, Familial hypoapoproteinemia
<i>ABCA2</i>	Drug resistance	unknown
<i>ABCA4</i>	Rod photoreceptor retinoid transport	Stargardt/fundus flavimaculatis, Retinitis pigmentosa, Cone-rod dystrophy, Age-related macular degeneration
<i>ABCB1</i>	Drug resistance	unknown
<i>ABCB2</i>	Peptide transport	Immune deficiency
<i>ABCB3</i>	Peptide transport	Immune deficiency
<i>ABCB4</i>	Bile-acid transport	Progressive familial intrahepatic cholestasis-3
<i>ABCB6</i>	Iron transport	unknown
<i>ABCB7</i>	Iron transport	X-linked sideroblastosis and anemia
<i>ABCB11</i>	Bile-acid transport	Progressive familial intrahepatic cholestasis-2
<i>ABCC1</i>	Drug resistance	unknown
<i>ABCC2</i>	Bile-acid transport	Dubin-Johnson Syndrome
<i>ABCC4</i>	Nucleoside transport	
<i>ABCC6</i>	unknown	Pseudoxanthoma elasticum
<i>ABCC7</i>	Chloride ion channel	Cystic fibrosis
<i>ABCC8</i>	Sulfonylurea receptor	unknown
<i>ABCD1</i>	Very long chain fatty acids transport	Adrenoleukodystrophy
<i>ABCE1</i>	Oligoadenylate-binding protein	unknown
<i>ABCG1</i>	Cholesterol transport	unknown
<i>ABCG2</i>	Drug resistance	unknown
<i>ABCG5</i>	Sterol transport	Sitosterolemia
<i>ABCG8</i>	Sterol transport	Sitosterolemia

#### *ABCG* subfamily (*white*)

The ABCG or White subfamily with its five fully characterized human members consists of half-size ABC proteins which probably dimerize to form active membrane transporters. Among the half-size molecules, ABCG proteins have a peculiar domain organization characterized by a nucleotide-binding domain at the N-terminus followed by six transmembrane-spanning domains (Klein *et al.* 1999). The founding member of this group, ABCG1, was described as the human homolog of the *Drosophila white* gene (Croop *et al.* 1997). Various transcripts of ABCG1 have been detected in different cells arising from alternative splicing events or the use of different transcription initiation sites (Lorkowski *et al.* 2001). Interestingly, the 12-amino acid linker region between the ATP-binding cassette and the

transmembrane region is also subject to alternative splicing generating two major protein forms of ABCG1. The mammalian ABCG1 is involved in the transport of cholesterol and phospholipids in macrophages. ABCG1 shows the highest phylogenetic relationship to the ABCA subfamily, particularly to the ABCA1 gene. There is a significant similarity in the differentiation and sterol-dependent regulation of the ABCA1 and ABCG1 in human monocytes and macrophages. Even though there is an experimental evidence for the involvement of ABCG1 in cholesterol and phospholipid transport, no loss of function mutation of the ABCG1 gene has been described that would help to clarify the physiological function of ABCG1 (Croop *et al.* 1997).

The second well-known member of the ABCG subfamily, ABCG2, has been identified by different

approaches and is known as placenta-specific ABC, a breast cancer resistance protein (Doyle *et al.* 1998), and a mitoxantrone resistance-associated protein. The protein has been shown to be amplified and overexpressed in human cancer cells and is capable of mediating drug resistance even in absence of the classic multidrug resistance proteins MDR1 and MRP1 (Robey *et al.* 2001). Most interestingly, new evidence of the function of ABCG2 as a direct drug efflux pump is provided by data localizing the bulk of the ABCG2 protein to the plasma membrane, with a minor fraction found within intracellular membranes.

Two other ABCG transporters, ABCG5 and ABCG8, encoding sterolin-1 and -2 respectively, mutations of which cause the human disease called sitosterolemia have been identified. Both genes are arranged in a head-to-head configuration, and only 140 bases separate their two respective start-transcription sites (Lu *et al.* 2002). Expression of both genes is confined to the liver and intestines. Sitosterolemia (also known as phytosterolemia) is a rare autosomal recessively inherited metabolic disorder (Salen *et al.* 1992). Sitosterolemia patients develop tendon and tuberous xanthomas, hemolytic episodes, arthralgias, and arthritis, and premature coronary and aortic atherosclerosis leading to cardiac fatalities (Lee *et al.* 2001). Affected individuals have very high levels of plasma plant sterols (sitosterol, campesterol, stigmasterol, avenosterol, and others) and 5 $\alpha$ -saturated stanols, particularly sitostanol, but their blood cholesterol levels may be normal or only moderately increased. Thus, in contrast to subjects, patients absorb and retain plant sterols (Salen *et al.* 1992) as well as shellfish sterols (Gregg *et al.* 1986). Clinical studies further show that affected individuals have an inability to excrete sterols, both plant sterols as well as cholesterol, into bile (Salen *et al.* 1989). Increased intestinal absorption, decreased hepatic excretion of sitosterol (the major plant sterol), and abnormally low cholesterol biosynthesis are reported to be features of sitosterolemia (Nguyen *et al.* 1990).

Five polymorphisms in the ABCG5 and ABCG8 genes have been identified (Hubáček *et al.* 2001) that influenced plasma lipid levels. The analyses indicated an association between a polymorphism (D19H) in exon 1 of ABCG8 and the plasma concentrations of campesterol. The finding of an association between plasma sitosterol concentrations and this nonconservative substitution suggests that the substitution of histidine for aspartic acid at amino acid 19 alters the function of ABCG8. Two

other polymorphisms examined were also associated with plasma sterol concentrations. A common nonconservative substitution (T400K) was associated with the plasma concentrations of sitosterol, although the association of this polymorphism with plasma campesterol concentration was marginal (Berge *et al.* 2002). The substitution of valine for alanine at amino acid 632 was associated with plasma cholesterol concentrations, but was not consistently associated with any of the other sterols. The other two polymorphisms (Q604E) in ABCG5 and (Y54C) in ABCG8 were not associated with plasma lipid levels.

The functions of the ABCG3 and ABCG4 transporters are unknown.

## Other ABC gene subfamilies

### *ABCB subfamily*

The ABCB subfamily is unique in that it contains both full and half transporters. Four full transporters and seven half transporters are currently identified as members of this subfamily. ABCB1 is the first human ABC transporter cloned and characterized through its ability to confer a multidrug-resistance phenotype to cancer cells (Juliano and Ling 1976). The functional sites of ABCB1 include the blood-brain barrier and the liver.

The ABCB2 and ABCB3 genes are half transporters that form a heterodimer to transport peptides into the endoplasmic reticulum, which are presented as antigens by the Class I HLA molecules.

The ABCB4 and ABCB11 proteins are both located in the liver and are involved in the secretion of bile acids (van Helvoort *et al.* 1996). Mutations in the human ABCB4 gene are the underlying cause of progressive familial intrahepatic cholestasis type 3 (Deleuze *et al.* 1996, de Vree *et al.* 1998). Patients have increased cholestatic serum markers and high  $\gamma$ -glutamyltranspeptidase levels. Mutations in the human ABCB11 gene are associated with progressive familial intrahepatic cholestasis type 2, a disease characterized by low biliary bile salt concentration, elevated serum bile salt concentrations, and normal  $\gamma$ -glutamyltranspeptidase levels (Strautnieks *et al.* 1998).

The remaining four half transporters, ABCB6, ABCB7, ABCB8 and ABCB10 localize on mitochondria, where they function in iron metabolism and transport of Fe/S protein precursors (Kispal *et al.* 1997). The closest homolog of the transporter associated with antigen

processing (TAPs) (ABCB2 and ABCB3), the ABCB9 half transporter, has been localized on lysosomes.

#### *ABCC subfamily*

The ABCC subfamily contains 12 full transporters with a diverse functional spectrum that includes ion transport, cell surface receptor, and toxin secretion activities. MRP-like proteins are organic anion transporters, i.e. they transport anionic drugs as well as neutral drugs conjugated to acidic ligands (glutathione, glucuronate, or sulfate) and play a role in resistance to nucleoside analogs (Borst *et al.* 2000).

The ABCC1 protein is a chloride-ion channel that has a role in exocrine secretion, and its mutations in ABCC1 cause cystic fibrosis (Quinton 1999). Cystic fibrosis is a genetic autosomal recessive disease. Cystic fibrosis causes the production of abnormally thick and sticky mucus in several different parts of the body, predominantly in the lungs and other parts of the respiratory system (Dean *et al.* 1990). It also affects the pancreas, leading to serious digestive problems.

Mutations in ABCC2 cause Dubin-Johnson syndrome. Dubin-Johnson syndrome is inherited as an autosomal recessive disorder. The transport of bilirubin from the liver into the biliary system is abnormal with bilirubin accumulating in the liver. Affected people have life-long low-grade jaundice, which may be aggravated by alcohol, pregnancy, infection, and other environmental factors (Wada *et al.* 1998).

Mutation in ABCC6 caused pseudoxanthoma elasticum. Pseudoxanthoma elasticum is an inherited systematic disorder affecting the connective tissue, and is characterized by progressive calcification of the elastic fibers of the eye, skin, and vasculature (Neldner 1988). Mutation in ABCC8 caused familial persistent hyperinsulinemic hypoglycemia of infancy. This is an autosomal recessive disorder in which subjects display unregulated insulin secretion (Thomas *et al.* 1995).

The ABCC4, ABCC5, ABCC11, and ABCC12 proteins are smaller than the other MRP1-like genes and lack an amino-terminal domain that is not essential for transport function.

## References

ALLIKMETS R: A photoreceptor cell-specific ATP-binding transporter gene (ABCR) is mutated in recessive Stargardt macular dystrophy. *Nat Genet* **17**: 122, 1997.

#### *ABCD subfamily*

The ABCD subfamily contains four genes in the human genome and two each in the *Drosophila* and yeast genomes. All of the genes encode half transporters that are located in the peroxisome (Shani *et al.* 1998), where they function as homo- and/or heterodimers in the regulation of very long chain fatty acid transport. Mutations in the ABCD1 gene have been identified in the majority of X-linked adrenoleukodystrophy (X-ALD). It is a neurodegenerative disorder characterized by progressive demyelination of the nervous white matter system and adrenal insufficiency (Mosser *et al.* 1993). Biochemically, the accumulation of very long chain fatty acids is the hallmark of all X-ALD forms.

#### *ABCE and ABCF subfamilies*

The ABCE and ABCF subfamilies contain proteins, which have ATP-binding domains that are clearly derived from others ABC transporters, but have no transmembrane domain and are not known to be involved in any membrane transport functions. The ABCE subfamily is comprised solely of the oligo-adenylate binding protein; the molecule recognizes oligo-adenylate that is produced in response to infection by certain viruses. Each ABCF gene contains a pair of nucleotide-binding folds. The best characterized member, the *S. cerevisiae* GCN20 gene mediates the activation of eIF-2  $\alpha$ -kinase (Marton *et al.* 1997) and its human homolog, ABCF1, is associated with the ribosome and appears to have a similar role (Tyzack *et al.* 2000).

## Conclusions

Study of ABC transporters is still in its infancy, but recent knowledge in this area has led to the elucidation of the cause of several significant inherited human diseases, and to new insights into possible treatments of cancer and drug pharmacokinetics.

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- ASSMANN G, VON ECKARDSTEIN A, BREWER HB: Familial high density lipoprotein deficiency: Tangier disease. In: *The Metabolic and Molecular Bases of Inherited Disease*. CR SCRIVER, AL BEAUDET, WS SLY, D VALLE (eds), New York 1995, pp 2053-2072.
- AZARIAN SM, TRAVIS GH: The photoreceptor rim protein is an ABC transporter encoded by the gene for recessive Stargardt's disease (ABCR). *FEBS Lett.* **409**: 247-252, 1997.
- BERGE KE, VON BERGMANN K, LUTJOHANN D, GUERRA R, GRUNDY SM, HOBBS HH: Heritability of plasma noncholesterol sterols and relationship to DNA sequence polymorphism in ABCG5 and ABCG8. *J Lipid Res* **43**: 486-494, 2002.
- BODZIOCH H, ORSO E, KLUCKEN J, LANGMANN T, BOTTCKER A, DIEDERICH W, DROBNIK W, BARLAGE S, BUCHLER C, PORSCH-OZCUMER M, KAMINSKI WE, HAMANN HW, OETTE K, ROTHE G, ASLANIDIS C, LACHNER KJ, SCHMITZ G: The gene encoding ATP-binding cassette transporter 1 is mutated in Tangier disease. *Nat Genet* **22**: 347-351, 1999.
- BORST P, EVERS R, KOOL M, WIJNHOLDS J: A family of drug transporters: the multidrug resistance-associated proteins. *J Natl Cancer Inst* **92**: 1295-1302, 2000.
- BROOKS-WILSON A, MARCIL M, CLEE SM, ZHANG LH, ROOMP K, VAN DAM M: Mutations in ABC1 in Tangier disease and familial high-density lipoprotein deficiency. *Nat Genet* **22**: 336-345, 1999.
- CROOP JM, TILLER GE, FLETCHER JA, LUX ML, RAAB E, GOLDENSON D, SON D, ARCINIEGAS S, WU RL: Isolation and characterization of a mammalian homolog of the *Drosophila* white gene. *Gene* **185**: 77-85, 1997.
- DE VREE JM, JACQUEMIN E, STURM E, CRESTEIL D, BOSMA PJ, ATEN J, DELEUZE JF, DESROCHERS M, BURDELSKI M, BERNARD O: Mutations in the MDR3 gene cause progressive familial intrahepatic cholestasis. *Proc Natl Acad Sci USA* **95**: 282-287, 1998.
- DEAN M, ALLIKMETS R: Evolution of ATP-binding cassette transporter genes. *Curr Opin Genet Dev* **5**: 779-785, 1995.
- DEAN M, WHITE MB, AMOS J, GERRARD B, STEWART C, KHAW KT, LEPPERT M: Multiple mutations in highly conserved residues are found in mildly affected cystic fibrosis patients. *Cell* **61**: 863-870, 1990.
- DELEUZE JF, JACQUEMIN E, DUBUISSON C, CRESTEIL D, DUMONT M, ERLINGER S, BERNARD O, HADCHOUEL M: Defect of multidrug-resistance 3 gene expression in a subtype of progressive familial intrahepatic cholestasis. *Hepatology* **23**: 904-908, 1996.
- DOYLE LA, YANG W, ABRUZZO V, KROGMANN T, GAO Y, RISHI AK, ROSS DD: A multidrug resistance transporter from human MCF-7 breast cancer cell. *Proc Natl Acad Sci USA* **95**: 15665-15670, 1998.
- ENDICOTT JA, LING V: The biochemistry of P-glycoprotein-mediated multidrug resistance. *Annu Rev Biochem* **58**: 137-171, 1989.
- GREGG RE, CONNOR WE, LIN DS, BREWER H Jr.: Abnormal metabolism of shellfish sterols in a patient with sitosterolemia and xanthomatosis. *J Clin Invest* **77**: 1864-1872, 1986.
- HIGGINS CF: ABC transporters: from microorganisms to man. *Annu Rev Cell Biol* **8**: 67-113, 1992.
- HIGGINS CF: The ABC of channel regulation. *Cell* **82**: 693-696, 1995.
- HOFFMAN HN, FREDRICKSON DS: Tangier disease (familial high density lipoprotein deficiency). Clinical and genetic features in two adults. *Am J Med* **39**: 582-593, 1965.
- HUBÁČEK JA, BERGE KE, COHEN JC, HOBBS HH: Mutations in ATP-cassette binding proteins G5 (ABCG5) and G8 (ABCG8) causing sitosterolemia. *Hum Mutat* **18**: 359-360, 2001.
- HYDE SC, EMSLEY P, HARTSHORN MJ, MIMMACK MM, GILEADI U, PEARCE SR, GALLAGHER MP, GILL DR, HUBBARD RE, HIGGINS CF: Structural model of ATP-binding proteins associated with cystic fibrosis, multidrug resistance and bacterial transport. *Nature* **346**: 362-365, 1990.
- JOHANSSON J, CRUSTEDT T: Molecular structures and interactions of pulmonary surfactant components. *Eur J Biochem* **244**: 675-693, 1997.
- JULIANO RL, LING V: A surface glycoprotein modulating drug permeability in Chinese hamster ovary cell mutants. *Biochim Biophys Acta* **455**: 152-162, 1976.
- JURANKA PF, ZASTAWNY RL, LING V: P-glycoprotein: multidrug-resistance and a superfamily of membrane-associated transport proteins. *FASEB J* **3**: 2583-2592, 1989.

- KAMINSKI WE, PIEHLER A, PULLMANN K, PORSCH-OZCURUMEZ M, DUONG C, MAA BARED G, BUCLER C, SCHMITZ G: Complete coding sequence, promoter region, and genomic structure of the human ABCA2 gene and evidence for sterol-dependent regulation in macrophages. *Biochem Biophys Res Commun* **281**: 249-258, 2001.
- KISPAL G, CSERE P, GUIARD B, LILL R: The ABC transporter Atm1p is required for mitochondrial iron homeostasis. *FEBS Lett* **418**: 346-350, 1997.
- KLEIN I, SARKADI B, VARADI A: An inventory of the human ABC proteins. *Biochim Biophys Acta* **1461**: 237-262, 1999.
- KLUGBAUER N, HOFMANN F: Primary structure of a novel ABC transporter with a chromosomal localization on the band encoding the multidrug resistance-associated protein. *FEBS Lett* **391**: 61-65, 1996.
- LEE MH, LU K, PATEL SB: Genetic basis of sitosterolemia. *Curr Opin Lipid* **12**: 141-149, 2001.
- LIN G, ORAM JF: Apolipoprotein binding to protruding membrane domains during removal of excess cellular cholesterol. *Atherosclerosis* **149**: 359-370, 2000.
- LORKOWSKI S, RUST S, ENGEL T, JUNG E, TEGELKAMP K, GALINSKI EA, ASSMANN G, CULLEN P: Genomic sequence and structure of the human ABCG1 (ABC8) gene. *Biochem Biophys Res Commun* **280**: 121-131, 2001.
- LU K, LEE MH, YU H, ZHOU Y, SANDELL SA, SALAN G, PATEL SB: Molecular cloning, genomic organization, genetic variations, and characterization of murine sterolin genes *Abcg5* and *Abcg8*. *J Lipid Res* **43**: 565-578, 2002.
- LUCIANI MF, DENIZOT F, SAVARY S, MATTEI MG, CHIMINI G: Cloning of two novel ABC transporters mapping on human chromosome 9. *Genomics* **21**: 150-159, 1994.
- LUTUCUTA S, BALLANTYNE CM, ELGHANNAM H, GOTTO AM, MARIAN AJ: Novel polymorphisms in promoter region of ATP binding cassette transporter gene and plasma lipids, severity, progression, and regression of coronary atherosclerosis and response to therapy. *Circ Res* **88**: 969-973, 2001.
- MARTON MJ, VAZQUEZ DE ALDANA CR, QIU H, CHAKRABURTTY K, HINNEBUSCH AG: Evidence that GCN1 and GCN20, translational regulators of GCN4, function on elongating ribosomes in activation of eIF2 $\alpha$  kinase GCN2. *Mol Cell Biol* **17**: 4474-4489, 1997.
- MENDEZ AJ, LIN G, WADE DP, LAWN RM, ORAM JF: Membrane lipid domains distinct from cholesterol/sphingomyelin-rich rafts are involved in the ABCA1-mediated lipid secretory pathway. *J Biol Chem* **276**: 3158-3166, 2000.
- MICHAELIS S, BERKOWER C: Sequence comparison of yeast ATP-binding cassette proteins. *Cold Spring Harbor Symp Quant Biol* **60**: 291-307, 1995.
- MOSSER J, DOUAR AM, SARDE CO, KIOSCHIS P, FEIL R, MOSER H, POUSTKA AM, MANDEL JL, AUBOURG P: Putative X-linked adrenoleukodystrophy gene shares unexpected homology with ABC transporters. *Nature* **361**: 726-730, 1993.
- NGUYEN LB, SALEN G, SHEFER S, TINT GS, SHORE V, NESS GC: Decreased cholesterol biosynthesis in sitosterolemia with xanthomatosis: diminished mononuclear leukocyte 3-hydroxy-3-methylglutaryl coenzyme A reductase activity and enzyme protein associated with increased low-density lipoprotein receptor function. *Metabolism* **39**: 436-443, 1990.
- OBMOLOVA G, BAN C, HSIEH P, YANG W: Crystal structures of mismatch repair protein MutS and its complex with a substrate DNA. *Nature* **407**: 703-710, 2000.
- ORAM JF: Tangier disease and ABCA1. *Biochim Biophys Acta* **1529**: 321-330, 2000.
- ORAM JF, VAUGHAN AM: ABCA1-mediated transport of cellular cholesterol and phospholipids to HDL apolipoproteins. *Curr Opin Lipidol* **11**: 253-260, 2000.
- ORAM JF, YOKOYAMA S: Apolipoprotein-mediated removal of cellular cholesterol and phospholipids. *J Lipid Res* **37**: 2473-2491, 1996.
- ORAM JF, MENDEZ AJ, SLOTTE JP, JOHNSON TF: High density lipoprotein apolipoproteins mediate removal of sterol from intracellular pools but not from plasma membranes of cholesterol-loaded fibroblasts. *Arterioscler Thromb* **11**: 403-414, 1991.

- QUINTON PM: Physiological basis of cystic fibrosis: a historical perspective. *Physiol Rev* **79** (Suppl): S3-S22, 1999.
- ROBEY RW, MEDINA-PEREZ WY, NISHIYAMA K, LAHUSEN T, MIYAKE K, LITMAN T, SENDEROWICZ AM, ROSS DD, BATES SE: Overexpression of the ATP-binding cassette half-transporter, ABCG2 (Mxr/BCrp/ABCP1), in flavopiridol-resistant human breast cancer cells. *Clin Cancer Res* **7**: 145-152, 2001.
- ROGLER G, TRUMBACH B, KLIMA B, LACKNER KJ, SCHMITZ G: HDL-mediated efflux of intracellular cholesterol is impaired in fibroblasts from Tangier disease patients. *Arterioscler Thromb Vasc Biol* **15**: 683-690, 1995.
- ROSENBERG MF, CALLAGHAN R, FORD RC, HIGGINS CF: Structure of the multidrug P-glycoprotein to 2.5 nm resolution determined by electron microscopy and image analysis. *J Biol Chem* **272**: 10685-10694, 1997.
- RUST S, ROSIER M, FUNKE H, REAL J, AMOURA Z, PIETTE JC, DELEUZE JF, BREWER HB, DUVERGER N, DENEFLÉ P, ASSMANN G: Tangier disease is caused by mutations in the gene encoding ATP-binding cassette transporter 1. *Nat Genet* **22**: 352-355, 1999.
- SALEN G, SHORE V, TINT GS, FORTE T, SHEFER S, HORAK I, HORAK E, DAYAL B, NGUYEN L, BATTAAK: Increased sitosterol absorption, decreased removal, and expanded body pools compensate for reduced cholesterol synthesis in sitosterolemia with xanthomatosis. *J Lipid Res* **30**: 1319-1330, 1989.
- SALEN G, SHEFER S, NGUYEN L, NESS GC, TINT GS, SHORE V: Sitosterolemia. *J Lipid Res* **33**: 945-955, 1992.
- SANTAMARINA-FAJO S, REMALEY A, NEUFELD E, BREWER HB: Regulation of intracellular trafficking of ABCA1. *J Lipid Res* **41**: 1339-1345, 2001.
- SHANI N, VALLE D: Peroxisomal ABC transporters. *Methods Enzymol* **292**: 753-776, 1998.
- SLOTTE JP, ORAM JF, BIERMAN EL: Binding of high density lipoproteins to cell receptors promotes translocation of cholesterol from intracellular membranes to the cell surface. *J Biol Chem* **262**: 12904-12907, 1987.
- STRAUTNIEKS S, BULL LN, KNISELY AS, KOCOSHIS SA, DAHL N, ARNELL H, SOKAL F, DAHAN K, CHILDS S, LING V: A gene encoding a liver-specific ABC transporter is mutated in progressive familial intrahepatic cholestasis. *Nat Genet* **20**: 233-238, 1998.
- TAKAHASHI Y, SMITH JD: Cholesterol efflux to apolipoprotein A-I involves endocytosis and resecretion in a calcium-dependent pathway. *Proc Natl Acad Sci USA* **96**: 11358-11363, 1999.
- THOMAS PM, COTE GJ, WOHLK N, NADDAD B, MATHEW PM, RABL W, AGUILAR-BRYAN I, GAGEL RF, BRYAN J: Mutations in the sulfonylurea receptor gene in familial persistent hyperinsulinemic hypoglycemia of infancy. *Science* **268**: 426-429, 1995.
- TYZACK JK, WANG X, BELSHAM GJ, PROUD CG: ABC50 interacts with eucaryotic initiation factor 2 and associates with the ribosome in an ATP-dependent manner. *J Biol Chem* **275**: 34131-34139, 2000.
- VAN HELVOORT A, SMITH AJ, SPRONG H, FRIETZSCHE I, SCHINKEL AH, BORST P, VAN MEER G: MDR1 P-glycoprotein is a lipid translocase of broad specificity, while MDR3 P-glycoprotein specifically translocates phosphatidylcholine. *Cell* **87**: 507-517, 1996.
- WADA M, TOH S, TANIGICHI K, NAKAMURA T, UCHIUMI T, KOHNO K, YOSHIDA I, KIMURA A, SAKISAKA S, ADACHI Y: Mutations in the canalicular multispecific organic anion transporter (cMOAT) gene, a novel ABC transporter, in patients with hyperbilirubinemia II/Dubin-Johnson syndrome. *Hum Mol Genet* **7**: 203-207, 1998.

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**Reprint requests**

Jana Štefková, CEM IKEM, Vídeňská 1958/9, 141 28 Praha 4-Krč, Czech Republic. E-mail: jana.stefkova@mk.cvut.cz