Oxidative Stress and Neuronal NOS Activity: Putative Determinants of Rapid Blood Pressure Increase After Renal Denervation in Anesthetized Rats

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Received September 18, 2012
Accepted January 3, 2013
On-line March 14, 2013

Summary
Long-term effects of renal denervation (DNX) commonly include a decrease in blood pressure (BP), observed in both normotensive animals and various models of hypertension. On the other hand, short term BP responses vary. We examined how post-DNX increase in BP observed in this study depends on baseline metabolic and functional status of animals, with a special interest for the role of oxidative stress. Anesthetized Wistar rats on standard (STD), low-sodium (LS) or high-sodium (HS) diet were used, untreated or pre-treated with tempol, a superoxide scavenger, or N(omega)-propyl-L-arginine (L-NPA), an inhibitor of neuronal NOS (nNOS). Early BP and renal hemodynamic responses were examined to right- and then left-side DNX performed using an own relatively non-invasive technique. Left kidney cortical, outer- and inner-medullary blood flows (CBF, OMBF, IMBF) were continuously recorded as laser-Doppler fluxes. Sequential denervations significantly increased BP to final 19 %, 12 %, and 6 % above control level in HS, LS, and STD groups, respectively. CBF, a measure of total renal perfusion, increased in LS and STD but not in HS rats. Tempol pretreatment prevented the post-denervation BP increase on each diet. Selective inhibition of nNOS prevented BP increase in STD and HS groups, a modest increase persisted in LS rats. We propose that enhanced afferent impulsation from intrarenal chemoreceptors related to oxidative stress in the kidney was the background for acute BP increase after DNX. The response was triggered by a release of brain sympatho-excitatory centers from inhibition by renal afferents, this was followed by widespread sympathetic cardiovascular stimulation.

Key words
Nitric oxide • Renal denervation • Free radicals

Introduction
Complex interrelation of renal nerve activity and cardiovascular functions determining arterial blood pressure (BP) has been studied in detail over the past decades and was the subject of a number of reviews (DiBona and Kopp 1997, Stella and Zanchetti 1991, Bie 2009, Pintérová et al. 2011). The long-term effect of renal denervation was usually a decrease in BP, as was documented in normal rats (Jacob et al. 2003, Kompanowska-Jezierska et al. 2008) and in various models of hypertension (Liard 1977, Wyss et al. 1992, Campese and Kogosov 1995). This was an expected phenomenon, given the stimulatory influence of renal sympathetic nerve activity (RSNA) on tubular reabsorption, renin release and renal vascular tone, and the crucial role of renal excretion in control of body fluid volume. By contrast, rapid cardiovascular and BP responses to changes in afferent and/or efferent renal sympathetic nerve activity were not uniform. This is not surprising since there is evidence that the effect of renal afferent impulse traffic on brain cardiovascular centers...
which control peripheral vascular resistance and BP may be either excitatory or inhibitory (reviewed in Stella and Zanchetti 1991). Selective electrical stimulation of renal afferents was reported to induce a decrease in BP in anesthetized rats (Lappe et al. 1985, Webb and Brody 1987, Herman and Kostreva 1996), which was ascribed to inhibitory influence of the afferent traffic on sympathetic centers controlling cardiovascular functions. On the other hand, renal denervation (DNX) did not usually induce short-term BP elevation (Kline et al. 1980, Winternitz et al. 1980). It was suggested that under the experimental conditions applied there was no tonic inhibitory influence on brain sympathetic centers (Stella and Zanchetti 1991). In this study we attempted to define the basal circumstances which might unmask the role of renal afferents in cardiovascular control.

As suggested above, the baseline level of RSNA, especially of the afferent traffic, is likely to affect cardiovascular response to DNX. Since this basal activity may depend, among other factors, on the metabolic status of the kidney which is strictly coupled to renal sodium handling, we examined BP responses in rats on either low or high sodium intake. An important feature of tissue metabolism under normal and pathologic conditions is the level of reactive oxygen and NO radicals (Wilcox 2005, Vanhoutte et al. 2009). To test if oxidative stress was, indeed, a crucial factor determining a possible pressure response to DNX, we repeated experiments after having scavenged the superoxides (O₂⁻) using tempol. Moreover, considering complex interaction of the autonomous nervous system activity and generation of ROS and nitric oxide (Xu et al. 2001, Wilcox 2005), we also examined how the response to DNX is affected by inactivation of neuronal NOS (nNOS), the isoform which, under conditions of oxidative stress may be an important source of ROS (Edlund et al. 2010). It is noteworthy that nNOS is also the isoform which prevails in the brain and influences the function of both central sympato-excitatory neurons and peripheral sympathetic nerve endings, e.g. in the vascular wall. To reduce the risk of transient stimulation of nerve fibers during denervation procedure, a crucial issue when short-time responses are studied, we applied here a relatively non-invasive technique which was proven to enable reliable renal hemodynamic measurements in the early post-denervation period (Kompanowska-Jezierska et al. 2001).

Material and Methods

The experimental procedures were in accordance with the Guide for the Care and Use of Laboratory Animals, NIH (Bethesda 1985) and were also approved by the extramural IV Local Ethical Committee, Warsaw. Male Wistar rats were fed a standard diet (STD, 0.25 % Na w/w) or, for 3 weeks, a low-sodium (LS, 0.15 % Na w/w) or high-sodium (HS, 4 % Na w/w) diet (SSNIFF GmbH, Soest, Germany). Our previous experience has shown that although in terms of Na concentration the difference between STD and LS diet is small, effects on renal sodium handling and body sodium status are remarkably different. The rats were anesthetized with intraperitoneal sodium thiopental (Sandoz GmbH, Kundl, Austria), 100 mg kg⁻¹, which provided stable anesthesia for about three hours. The animals were placed on a heated surgery table to maintain rectal temperature at about 37 °C. A tracheal cannula ensured free airways. The femoral vein was cannulated for fluid infusions and the carotid artery for BP measurement (Stoelting blood pressure meter and transducers, Wood Dale, Illinois, USA). During surgery, 3 % bovine serum albumin solution was infused i.v. at 3 ml h⁻¹ to preserve plasma volume.

Experimental procedures

The right kidney was accessed by flank incision. Stainless steel wire (0.1 mm in diameter) was used to form a loop which was placed around the fat and connective tissue located cephalad to the right renal artery-aortic junction. Dissection studies had shown that this tissue encompassed most of renal nerve fibers traveling from the coeliac ganglion to the kidney (Kompanowska-Jezierska et al. 2001). The loop was threaded through a segment of thick-walled PE tubing, and left loose, prepared for later right-side DNX. Subsequently, the left kidney was exposed from flank incision and immobilized in a plastic holder. Another wire loop was placed around the tissue encompassing the nerves, to prepare for left DNX. The effectiveness of this denervation technique was confirmed by demonstration of the depletion of renal tissue noradrenaline content to <5 % of control as well as of an acute rapid increase in CBF (confirmed also in the present study) and of distinct denervation natriuresis (Kompanowska-Jezierska et al. 2001). A laser-Doppler probe, type PF 407, was placed on left kidney surface to record superficial cortical laser-Doppler blood flow (CBF). Outer medullary blood flow...
(OMBF) and inner medullary blood flow (IMBF) were measured using needle probes (PF 402) inserted to the respective depths of 3 and 5 mm from kidney surface. The probes were connected with a three-channel Periflux 5010 flow-meter (Perimed, Jarfalla, Sweden). Other details of the experimental techniques were described previously (Kompanowska-Jezierska et al. 2001, Kompanowska-Jezierska et al. 2008).

After placement of wire loops for denervation, flow probes in the left kidney, and after stabilization of the parameters measured, control values were recorded during 30 min. Subsequently, right-side DNX was performed. At first, 1 % novocain solution was injected into the PE tubing segment containing the wire loop, to provide a gradual rather than abrupt interruption of conduction in the renal nerves and prevent transient stimulation of nerve fibers (Kompanowska-Jezierska et al. 2001). The loop was tightened and the tissue within was cauterized using a 5 MHz current generated for 5-10 s using a neurosurgical cautery device (Famed, Warsaw, Poland). Five minutes later a 30-min experimental and measurement period was made. Thereafter left DNX was performed, followed by another 30-min observation period. In two rats on STD diet and three rats on HS diet the observation period after left-side DNX was prolonged to a total of 60 min. The averaged data for the last five min of each post-denervation period were taken as a final result of DNX I (right-side) and DNX II (left-side) and shown in figures. While the standard denervation technique involves cutting the nerve fibers adjacent to the renal artery and damaging those in the artery wall using phenol, our approach interrupts neural conduction distant from the artery, whereby the usual vessel spasm and derangement of intrarenal circulation is avoided. This enables starting experimental measurements within a few minutes post-denervation.

It will be noticed that within the experimental design applied here the renal hemodynamic parameters are determined in the left kidney only; consequently, the responses to right-side denervation (DNX I) are those of the contralateral innervated kidney and the responses to DNX II are simply those of ipsilateral denervation. Three groups of rats were examined within the protocol described above, on each of the three diets. Group 1. Untreated rats (LS: n=6; STD: n=7; HS: n=7), group 2. Rats pretreated with N(omega)-propyl-L-arginine (L-NPA, Tocris, UK), a selective inhibitor of nNOS, infused i.v. at 1 mg kg⁻¹ h⁻¹ (LS: n=6; STD: n=6; HS: n=5).

In groups 2 and 3, after a control period, about 40 min were allowed from the start of L-NPA or tempol infusion, for stabilization of the variables measured. For sham denervation (STD, n=5), fat and connecting tissue in the area caudal to the kidney were cauterized.

In a part of experiments with LS and HS rats arterial blood samples were taken for determination of plasma renin activity by antibody-trapping technique (Poulsen and Jorgensen 1974).

Statistics

Significance of changes within one group over time was first evaluated by repeated measurement analysis of variance (ANOVA), followed by modified t statistics for dependent variables. Differences between groups were first analyzed by the classical one-way ANOVA, followed by Bonferroni’s tests for multiple comparisons.

Results

After right-side and left-side DNX, BP almost always showed a progressing increase. Figure 1 shows BP and renal cortical blood flow (CBF) changes after sequential denervation procedures in rats on standard, low and high sodium intake (LS, STD, HS). In LS and HS rats bilateral DNX increased BP 12 and 19 %, respectively; significant increases were seen already after unilateral (right-side) denervation. BP elevations were not associated with any changes in heart rate: in the group with greatest responses (HS) it was 365±5 before and 366±7 beats per min after second denervation (n=8, data not shown). Nor were there significant heart rate changes in STD or LS groups. In STD rats no significant change in BP was seen after right-side DNX and the final increase was less than 6 %. Both after right-side and left-side DNX, the pressure usually increased progressively during the first 15 min, which was followed by a plateau. Prolongation of observation by an additional 30 min (to a total of 60 min after left side DNX) showed that the plateau was maintained over the whole period.

In STD rats, left kidney CBF increased after the right side- and then further after left-side denervation. In the LS group, CBF also increased progressively (ANOVA) but the ultimate change did not reach significance level in a Bonferroni test. Remarkably, in HS pretreated with 4-hydroxy-Tempo (Tempol, Sigma, USA), a scavenger of O₂−, infused i.v. at 12 mg kg⁻¹ h⁻¹ (LS: n=6; STD: n=6; HS: n=5).
rats CBF did not change at all. Both BP and CBF remained stable in STD rats subjected to sham denervation. Figure 2 shows that in LS and STD rats, DNX did not significantly alter perfusion of the inner medulla; IMBF decreased slightly in untreated HS rats. No changes in OMBF were seen (data not shown).

**Fig. 1.** Effects of sequential right- and left-side renal denervation (Den I, II) on systemic blood pressure (BP) and left kidney cortical blood flow (CBF) in rats on low, standard and high sodium diet (LS, n=6; STD, n=7 and HS, n=7, respectively). Mean % ± SEM; sham denervation data for STD group are also shown. * significantly different from pre-denervation control at p<0.05; † significant progressive increase (repeated measurements ANOVA); the final increase did not reach significance level in Student t test (0.05<p<0.06).

**Fig. 2.** A comparison of final effects of sequential right and left renal denervation on blood pressure (BP) and left kidney cortical blood flow (CBF), and inner medullary blood flow (IMBF) in untreated, L-NPA- or tempol-treated rats (open, black or striped bars, respectively) maintained on low-sodium (LS), standard (STD) or high-sodium (HS) diet. Cumulated percent changes after two denervations are shown, ± SEM. * significant change from pre-denervation control at p<0.05; † change from pre-denervation control significantly different from that in untreated rats. ‡ change significantly different from that in STD rats (p<0.02).
Table 1. Effects of Tempol on blood pressure (BP) and perfusion parameters of the left kidney.

<table>
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<th>LOW SODIUM DIET</th>
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<th>HIGH SODIUM DIET</th>
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<td></td>
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<td>Tempol</td>
<td>Control</td>
<td>Tempol</td>
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<tr>
<td><strong>BP (mmHg)</strong></td>
<td>116±1</td>
<td>111±3</td>
<td>117±4</td>
<td>118±4</td>
<td>126±5</td>
<td>127±4</td>
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<tr>
<td><strong>CBF (PU)</strong></td>
<td>550±33</td>
<td>684±76*</td>
<td>617±71</td>
<td>700±76*</td>
<td>801±86</td>
<td>776±85</td>
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<tr>
<td><strong>OMBF (PU)</strong></td>
<td>143±26</td>
<td>181±27*</td>
<td>159±20</td>
<td>160±11</td>
<td>194±39</td>
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<td><strong>IMBF (PU)</strong></td>
<td>140±8</td>
<td>162±19</td>
<td>169±20</td>
<td>175±28</td>
<td>200±17</td>
<td>213±28</td>
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Mean values ± SEM (n=5-7). CBF, OMBF, IMBF – cortical, outer and inner medullary laser-Doppler fluxes, respectively. PU – perfusion units. * Significantly different from control at p<0.04.

Table 2. Effects of L-NPA on blood pressure (BP) and perfusion parameters of the left kidney.

<table>
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<td>NPA</td>
<td>Control</td>
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<tr>
<td><strong>BP (mmHg)</strong></td>
<td>120±3</td>
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<td>117±2</td>
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<td>129±9</td>
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<td><strong>CBF (PU)</strong></td>
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<td>412±39</td>
<td>580±41</td>
<td>621±44*</td>
<td>719±43</td>
<td>679±48*</td>
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<td><strong>OMBF (PU)</strong></td>
<td>132±27</td>
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<tr>
<td><strong>IMBF (PU)</strong></td>
<td>133±23</td>
<td>121±23</td>
<td>189±31</td>
<td>203±31*</td>
<td>117±8</td>
<td>137±17</td>
</tr>
</tbody>
</table>

Mean values ± SEM (n=5-7). CBF, OMBF, IMBF – cortical, outer and inner medullary laser-Doppler fluxes, respectively. PU – perfusion units. * Significantly different from control at p<0.02.

The basal plasma renin concentration (PR) on HS diet (10±2 mIU l⁻¹) was significantly lower than in LS and STD groups (range of means: 79-81 mIU l⁻¹). While in both latter groups the basal values were quite comparable, bilateral DNX significantly decreased plasma renin (~45 %) in LS rats whereas no substantial change (~6 %) was seen in STD rats.

Pre-denervation CBF tended to be higher with increasing salt intake (Tables 1, 2) Treatment with 4-hydroxy-Tempo (tempol), a scavenger of O₂⁻ did not alter BP on any diet (Table 1) but significantly increased CBF in LS and STD groups. OMBF increased in LS rats only. L-NPA, a selective inhibitor of nNOS, did not alter BP on any diet (Table 2). In HS rats L-NPA decreased CBF, which suggested that NO derived from nNOS was on high-salt diet an important factor sustaining renal perfusion. Apparently, this was not so in LS and STD groups where CBF (and IMBF) actually increased after L-NPA.

The final cumulated effect of sequential right and left DNX on BP, and left kidney CBF and IMBF in rats on different diets, untreated or under tempol or L-NPA treatment, is presented in Figure 2. The BP increase in untreated HS rats was significantly greater than in the STD group where it was least pronounced. In LS rats treated with L-NPA the post-denervation increase in BP was significantly smaller than in untreated controls. Remarkably, in STD and HS groups significant decreases occurred.

Bilateral DNX modestly increased CBF in untreated STD and LS (not significant) rats, much less so in rats pretreated with L-NPA. CBF never changed in HS rats.

In rats pretreated with tempol, the post-denervation increases of BP observed in untreated LS and STD rats were abolished. In HS rats, similarly as under L-NPA treatment, a significant decrease occurred. IMBF significantly increased only in STD rats under L-NPA or tempol treatment.

Discussion

In normotensive anesthetized Wistar rats acute sequential right-side and left-side renal denervation (DNX) induced a rapid progressing elevation of BP. Earlier studies usually revealed no alteration of BP within the first hours post-denervation (Kline et al. 1980, Winternitz et al. 1980). In contrast to these observations,
the usual long term effect of DNX was BP lowering (Stella and Zanchetti 1991, Jacob et al. 2003), a complex phenomenon obviously related to profound alterations of endocrine and body fluid status. There is no rationale to set aside or directly compare the early and the distant responses to DNX.

**Reflex nature of the hypertensive response to renal denervation**

The mechanism of the rapid blood pressure elevation after renal denervation is not clear. It is more likely to be neural rather than humoral. Indeed, at least in LS rats, DNX decreased the circulating plasma angiotensin II level, which would have vasodilator and hypotensive rather than hypertensive effects. It will be noticed that the advantage of denervation technique applied here is that no surgical severing of nerve fibers adjacent to the renal artery occurs. This reduces the risk of a BP increase secondary to the artery spasm which is typically observed after the classical DNX manoeuvre; moreover, early fluctuations of renal hemodynamics are avoided and measurements can be conducted immediately (Kompanowska-Jezierska et al. 2001). An obvious disadvantage limiting interpretation of the present results is that the denervation is “complete” i.e. both efferent and afferent.

The data on the reflex influence of renal afferent traffic on the activity of cardiovascular sympathetic centers are ambiguous. Some authors demonstrated excitatory effects, observed in normal animals (reviewed in Stella and Zanchetti 1991) and in some but not in other models of hypertension and renal failure (Wyss et al. 1992, Campese and Kogosov 1995). On the other hand, there is ample evidence on the sympatho-inhibitory influence of the renal afferent traffic. Unilateral afferent or complete DNX increases efferent sympathetic input to the contralateral kidney within the reno-renal reflex phenomenon, leading, on this side, to antidiuresis and antinatriuresis, occasionally to increased renin release and phenomenon, leading, on this side, to antidiuresis and

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using the same denervation technique (Kompanowska-Jezierska et al. 2001). Interestingly, no such increase was observed in HS rats, possibly because under baseline conditions the renal circulation was here under dominating vasodilator influence of NO, as indicated by a decrease in CBF after NOS blockade (Table 2).

Role of sodium intake, ROS and nNOS: the background for hypertensive response to renal denervation

The post-DNX elevation of BP was minimal in STD rats but pronounced in those on high or low sodium intake. It will be noticed that the common feature of both conditions is enhanced oxidative stress: at low sodium intake this is largely mediated by high plasma Ang II activity (Majid and Kopkan 2007). High sodium intake induces oxidative stress by mechanisms that are still being evaluated; there is also evidence that in rats on high-salt diet ROS generation is increased in kidney tissue (Kitiyakara et al. 2003, Wilcox 2005).

Local oxidative stress might increase stimulation of intrarenal chemoreceptors, enhance the inhibitory afferent traffic and lower the activity of sympathetic vasomotor centers. Under such baseline conditions, DNX (strictly: de-afferentation) i.e. release of the centers from the pronounced inhibition might be more effective than without enhanced oxidative stress. It must be admitted, however, that because of the diversity of intrarenal chemo- and mechanoreceptor types, individual afferent fibers might conduct both inhibitory and excitatory signals so that the final effect on sympathetic cardiovascular centers is difficult to predict, depending, for instance, on the actual baseline metabolic and functional status. The stress level would probably be lower in rats on STD diet, hence relatively modest BP increase in rats of this group.

Baseline conditions of our experiments, including anesthesia, extensive traumatic surgery and increased or decreased dietary sodium intake, were the probable factors promoting the development of oxidative stress. To check the hypothesis that high ROS activity was the necessary background for the pressor response to DNX, we repeated experiments in rats pretreated with tempol, a scavenger of superoxide radicals.

Tempol treatment did not affect BP per se, irrespective of sodium intake (Table 1). The increase in CBF suggests that under conditions of our experiments increased \( O_2^- \) activity induced tonic intrarenal vasoconstriction. However, this effect was not seen in HS rats even though their kidneys are expected to be in the state of pronounced oxidative stress (Majid and Kopkan 2007). As suggested earlier, most probably local NO activity was here the dominating factor responsible for vasodilatation and high perfusion.

Under tempol treatment the post-DNX elevation of BP was abolished in LS rats and reverted to a significant decrease in HS rats in which the original hypertensive response was most pronounced, which confirmed the crucial role of the oxidative stress. As discussed earlier, of critical importance for the hypertensive response might be high ROS activity in the kidney. However, since tempol was administered intravenously, the present studies do not provide the basis to define the relative contribution of the effects of superoxide scavenging in the kidney, in the brain sympathetic centers and in target vascular tissues.

There were good reasons to suspect that nitric oxide (NO) status might be an important co-determinant of the BP response to DNX. NO specifically inhibits the sympathetic cardiovascular centers (Sakuma et al. 1992, Hansen et al. 1994, Hakim et al. 1995, Zanzinger 1999, Pintérová et al. 2011) and antagonizes ROS in most tissues, including the brain, kidney, peripheral vessels and the heart; this is observed both in normotensive animals and in rat models of hypertension (Golín et al. 1987, Zicha et al. 2001, Wilcox 2005). Of the two constitutive NO synthases, nNOS and NO derived from this isoform are particularly likely to be involved in the reflex control of cardiovascular functions and blood pressure. First, nNOS is the main source of NO in the brain (Bredt et al. 1990). Second, compared to eNOS, it can easier undergo uncoupling under oxidative stress, and generate ROS rather than NO (Sun et al. 2008). Indeed, in the diabetic kidney nNOS was reported to be a major if not the main source of ROS (Edlund et al. 2010). Therefore we examined how previous inhibition of this isoform with L-NPA would affect the BP response to DNX.

We saw that L-NPA did not change baseline BP, in agreement with other studies in which this inhibitor was used in rats on normal (Kakoki et al. 2001, Vaněčková et al. 2002) or high sodium diet (Walkowska and John, unpublished observations). On the other hand, L-NPA treatment reverted the post-denervation increase in BP in HS and STD rats to a significant decrease; only a modest increase in BP was seen in the LS group. The mechanism of the blockade of the increase may be complex. The inhibitory influence of the renal afferent sympathetic traffic is transmitted via the nucleus tractus
solitarii to the neurons of the paraventricular nucleus (PVN). There is good evidence that release of NO by nNOS located in PVN mitigates the center’s firing activity, which might inhibit the vasomotor function of sympathetic centers, such as rostral ventrolateral medulla (Li et al. 2003), local nNOS-derived NO was also found to buffer the rise in arterial pressure in 2K1C hypertensive rats (Rossi et al. 2010). Therefore, a deficit of NO mediator induced in our rats by L-NPA would cut off this influence and thereby prevent BP elevation after de-afferentation as observed in untreated rats.

Another reason for the blockade by L-NPA of the blood pressure response to DNX relates to the observation that in oxidative stress uncoupled nNOS in the kidney generates large amounts of ROS (Edlund et al. 2010). Therefore, blockade of this NOS isoform would have consequences similar as those of tempol treatment.

Taken together, these results strongly suggest that acute BP elevation after DNX was triggered by a release of brain sympatho-excitatory centers from the inhibitory influence of afferent impulse traffic from the kidneys, followed by increased sympathetic stimulation of target vascular tissues and widespread vasoconstriction. Additional studies involving measurements of sympathetic nerve activity (e.g. muscle SNA) would be needed to verify the hypothesis. Thereafter, it would be crucial to check if MSNA response to denervation is inhibited by ablation of the nucleus tractus solitarii or ganglionic blockade. A high level of oxidative stress, especially in the kidney, seems a necessary condition for the pressor response: previous reduction of the oxidative stress with tempol prevents the increase in BP after DNX. The response is also prevented by elimination of nNOS-derived NO, a likely mediator of the inhibition of the sympathetic centers by renal afferent traffic and also an important source of ROS in the kidney. The study helps understand some new aspects of interaction of the sympathetic nervous system and the enzymes generating ROS and NO in control of cardiovascular functions and blood pressure.

The present results showing acute BP rise refer exclusively to the earliest post-DNX period and by no means undermine the utility of catheter-based renal sympathetic denervation successfully used for treatment of refractory hypertension (Symplicity HTN-1 Investigators 2011). No serious early adverse effects of radiofrequency nerve ablations were reported until quite recently: four out of a group of 12 patients subjected to the procedure had to be temporarily transferred to an emergency unit because of “severe hypertensive episodes” (Brinkmann et al. 2012). It can be speculated that this occurred in patients with pronounced oxidative stress or endothelial dysfunction, which rendered them particularly sensitive to de-afferentation.

Conflict of Interest
There is no conflict of interest.

Acknowledgements
We are indebted to Professor Peter Bie, in whose department (Institute of Molecular Medicine, University of Southern Denmark, Odense, Denmark) renin assays were performed. Supported by the Polish Ministry of Science and Higher Education (grant NN401108038).

Abbreviations
BP, blood pressure; CBF, cortical blood flow; DNX, renal denervation; HS, high-sodium diet; IMBF, inner medullary blood flow; L-NPA, N(omega)-propyl-L-arginine; LS, low-sodium diet; NO, nitric oxide; OMBF, outer medullary blood flow; ROS, reactive oxygen species; RSNA, renal sympathetic nerve activity; RVLM, rostral ventrolateral medulla; SOD, superoxide dismutase; Tempol, 4-hydroxy-Tempo.

References


