

Protein Interactions With Quaternized Chitosan/Heparin Multilayers

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Summary

Understanding the behavior of single proteins at the polyelectrolyte multilayer film/solution interface is of prime importance for the designing of bio-functionalized surface coatings. In the present paper, we study the adsorption of the model proteins, albumin and lysozyme, as well as basic fibroblast growth factor (FGF-2) on a polysaccharide multilayer film composed of quaternized chitosan and heparin. Several analytical methods were used to describe the formation of the polysaccharide film and its interactions with the proteins. Both albumin and lysozyme adsorbed on quaternized chitosan/heparin films, however this process strongly depended on the terminating polysaccharide. Protein adsorption was driven mainly by electrostatic interactions between protein and the terminal layer of the film. The effective binding of FGF-2 by the heparin-terminated film suggested that other interactions could also contribute to the adsorption process. We believe that this FGF-2-presenting polysaccharide film may serve as a biofunctional surface coating for biologically-related applications.

Key words

Heparin • Chitosan • Protein • Basic fibroblast growth factor • Layer-by-layer

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Introduction

Layer-by-layer (LbL) assembly is a simple method that enables the creation of versatile, robust,

nanometer scale coatings on a variety of surfaces. Generally, the LbL fabrication process is performed in non-denaturing aqueous conditions, thus allowing for the incorporation of fragile biomolecules. The polyelectrolyte multilayer films functionalized with proteins, such as growth factors, are of great interest as bioactive surface coatings for biologically-related applications (Monge *et al.* 2015, Gribova *et al.* 2012). Recently, we reported on cross-linked albumin/heparin multilayer films for the immobilization of the basic fibroblast growth factor (FGF-2) (Kumorek *et al.* 2015).

Two polysaccharides, chitosan exhibiting an antimicrobial property (Wiarachai *et al.* 2012, Follmann *et al.* 2012), and heparin known as an anticoagulant and growth factor binding agent (Faham *et al.* 1996, Wang *et al.* 2014), have been extensively exploited to develop biofunctional multilayer films (Almodovar *et al.* 2010, Monge *et al.* 2015). However, the formation of the chitosan-based coatings can lead to some difficulties due to the limited solubility of chitosan above pH=6 (Wiarachai *et al.* 2012). Furthermore, the LbL assembly of chitosan as a weak polycation with polyanions requires a very acidic pH to ensure an appropriate protonation of amino groups of chitosan (Almodovar *et al.* 2010). One way to improve the solubility of chitosan is the quaternization of amine groups. Such cationically modified chitosan exhibits good solubility over a broader pH range (Follmann *et al.* 2012). Furthermore, quaternized chitosan/heparin multilayers displayed an improved antibacterial property as compared to the native chitosan-based film (Follmann *et al.* 2012). Also, N,N,N-trimethyl chitosan with heparin in the form of polyelectrolyte complex nanoparticles demonstrated an efficient delivery system of growth factor (Volpatto *et al.* 2012).

Understanding the behavior of single proteins at the polyelectrolyte film/solution interface is of prime importance for designing bio-functionalized multilayer coatings. Depending on the application, one can select LbL films which enhance or reduce the adsorption of the specific biomolecules (Ariga *et al.* 2014). Protein interactions with LbL films depend on many factors, particularly on the charge of the protein and the terminating layer of film (Salloum and Schlenoff 2004). In the present paper, we study the adsorption of two model proteins of different isoelectric points, I_p , i.e. albumin ($I_p=4.9$ (Salloum and Schlenoff 2004)) and lysozyme ($I_p=11.4$ (Salloum and Schlenoff 2004)) as well as FGF-2 ($I_p=9.59$ (Dephillips and Lenhoff 2004)) on a polysaccharide multilayer film composed of chitosan quaternized with glycidyltrimethylammonium chloride and heparin. The LbL films were terminated with quaternized chitosan or heparin. The surface plasmon resonance and UV-Vis spectroscopy were used to quantitatively describe the formation of polysaccharide LbL films and their interaction with proteins. Simultaneously, the qualitative surface information of quaternized chitosan/heparin LbL films with or without adsorbed protein was obtained by atomic force microscopy.

Material and Methods

Materials

Chitosan (Ch, LMW), heparin sodium salt from porcine intestinal mucosa (Hep, 17-19 kDa), glycidyltrimethylammonium chloride (GTMAC, ≥90 %), bovine serum albumin (Alb, ≥96 %), lysozyme from chicken egg white (Lys, ≥90 %), albumin-fluorescein isothiocyanate conjugate (Alb^{FITC} , FITC:Alb molar ratio 10:1), fluorescein isothiocyanate isomer I (FITC, ≥90 %), 16-mercaptophexadecanoic acid (ACID-16, 99 %), phosphate buffer saline (PBS) were purchased from Sigma-Aldrich.

The synthesis of cationically modified chitosan (CMCh)

Chitosan was modified by GTMAC as described previously (Cho *et al.* 2006). Briefly, 2.5 g of Chitosan was dispersed in 100 ml of distilled water containing 21 ml of 0.5 % acetic acid. After one hour of stirring, 6.9 ml of GTMAC was added and the reaction was carried out at 55 °C for 18 h. The product was precipitated into acetone and isolated by centrifugation (6000 rpm for 5 min). This process was repeated twice.

Afterward, the product was dissolved in water, dialyzed (Spectra/Por, cut off 6-8 kDa) against distilled water for 2 days, and freeze-dried. The product was characterized with elemental analysis (CMCh, C% = 41.88; H% = 8.45; N% = 7.83). The reaction's occurrence was confirmed by ^1H NMR ($\text{D}_2\text{O}/\text{CF}_3\text{COOD}$, 300 MHz): $\delta=3.2$ ppm corresponds to the protons of the $-\text{N}(\text{CH}_3)_3$ group of GTMAC, and FTIR: 1480 cm^{-1} assigned to an asymmetric angular bending of C-H of $-\text{CH}_3$ groups of GTMAC; the disappearance of the band at 1590 cm^{-1} assigned to vibrations of N-H of the primary amino groups of Chitosan.

The degree of deacetylation (DDA) of Chitosan was 78.6 % as calculated from elemental analysis data (Kasaai *et al.* 2000).

The degree of substitution (DS) of CMCh was obtained from conductometric titration of the CMCh aqueous solution with the AgNO_3 solution according to (Cho *et al.* 2006) using a digital conductivity meter (Jenway, UK). The DS was calculated using the following equation:

$$DS = \frac{1.7 \times 10^{-5} V'_{\text{AgNO}_3}}{\left(\frac{W_{\text{CMCh}} - 1.7 \times 10^{-5} V^*_{\text{AgNO}_3} m_{\text{GTMAC}}}{m_G DDA + (1 - DDA)m_{\text{AG}}} \right) DDA} \times 100\% \quad (1)$$

where W_{CMCh} is the weight of CMCh in 10 ml of the analyzed sample; m_{GTMAC} is the molecular weight of GTMAC; m_G and m_{AG} are the molecular weights of glucosamine and acetylated glucosamine respectively; DDA is the degree of deacetylation of chitosan (i.e. 78.6 %); $V^*_{\text{AgNO}_3}$ is the total volume of the titrant added at the endpoint of titration; and 1.7×10^{-5} is the number of AgNO_3 moles in 1 ml of the titrant solution.

The labeling of compounds

FITC labeling of CMCh ($\text{CMCh}^{\text{FITC}}$). The CMCh was conjugated with FITC according to (Geisberger *et al.* 2013) with slight modifications. Briefly, the solutions of CMCh (32 mg) in distilled water (3 ml) and FITC (3 mg) in methanol (5 ml) were mixed and stirred for 36 h in a dark at room temperature. Then the reaction mixture was dialyzed (Spectra/Por, cut off 6-8 kDa) against distilled water for 3 days, and freeze-dried.

FITC labeling of lysozyme (Lys^{FITC}). The FITC-labeled lysozyme was prepared according to (Zhang *et al.* 2010). Briefly, 400 μl of a freshly prepared solution of FITC (1 mg/ml) was dropped into 2.4 mg/ml of lysozyme in a 0.1 M carbonate-bicarbonate buffer (pH 9). The reaction was carried out overnight in the dark at 4 °C.

The product was dialysed (Spectra/Por, cut off 6–8 kDa) against a PBS buffer (pH 7.4) and freeze-dried. The molar ratio of FITC to Lys was 1, as determined using the protocol provided by the Sigma-Aldrich.

Substrate preparation

Gold-coated SPR chip, quartz, and round-shaped glass (with a diameter of 12 mm) were firstly washed by sonication in methanol and then in water. All substrates were immersed in a H₂O₂/NH₄OH/H₂O mixture (1:1:5 vol/vol/vol) at 70 °C for 15 min, then rinsed with water, dried, and exposed to air plasma for 10 min. The gold surface of the SPR chips were then modified with self-assembled monolayers (SAM) using ACID-16. The SPR chip was kept in 1 mM ACID-16 in ethanol in the dark for 24 h, then rinsed with ethanol and water, and dried with nitrogen.

LbL film preparation

All cleaned substrates were immersed in 1 mg/ml CMCh in 10 mM PBS for 5 minutes, then rinsed with PBS, and immersed in 1 mg/ml Hep in PBS for 5 minutes. These steps were repeated to obtain the required number of layers.

Surface analysis

Surface plasmon resonance (SPR). The formation of CMCh/Hep LbL films and protein adsorption on SAM-modified chips (see Substrate preparation) was monitored *in situ* as a shift in the resonance wavelength using a custom-built SPR sensor (Institute of Photonics and Electronics, Academy of Sciences of the Czech Republic) at 25 °C. The solutions of CMCh, PBS, and Heparin were subsequently pumped into the SPR flow cell with a flow rate of 25 µl/min. The Hep- and CMCh-terminated LbL films were exposed to Alb (1 mg/ml in PBS), Lys (1 mg/ml in PBS), or FGF-2 (100 ng/ml in PBS) solutions for 30 or 90 min, and then rinsed with PBS. The surface density (in ng/cm²) of the particular proteins was calculated from a difference in resonance wavelength ($\Delta\lambda_{\text{res}}$) registered in PBS before and after exposition to the protein solution; $\Delta\lambda_{\text{res}}=1 \text{ nm}$ corresponds to the protein surface density of 15 ng/cm² (Emmenegger *et al.* 2009).

UV-VIS Spectroscopy. The UV-Vis absorption spectra of CMCh^{FITC}/Hep LbL films and CMCh/Hep LbL films with adsorbed Alb^{FITC} or Lys^{FITC} were recorded using a spectrophotometer Specord Plus (Analytik Jena). LbL films were deposited on quartz slides, then exposed

to 1 mg/ml protein solution in PBS for 2 h, then rinsed with PBS and dried in a stream of nitrogen before the measurement.

Atomic force microscopy (AFM). The surface topography of LbL films on glass cover slides was observed on Nanoscope IIIa, Digital Instruments in tapping mode with standard silicon cantilevers OTESPA7 (Veeco Instruments, a spring constant of 42 N/m) and a scan rate of 0.8–1 Hz in air.

Results

Characterization of cationically modified (quaternized) chitosan, CMCh

Chitosan was subjected to cationic modification with GTMAC. The occurrence of the reaction was confirmed by FTIR and ¹H NMR spectroscopies (see experimental section). The received product, CMCh, showed good water solubility regardless of the pH. The degree of substitution (DS) of Ch with GTMAC was determined using the conductometric titration of CMCh with AgNO₃ (Fig. 1). Initially, the conductivity of the solution gradually decreased with the increasing volume of AgNO₃ added as a result of the formation of an insoluble AgCl salt. The lowest solution conductivity, considered as the endpoint of the titration, was registered when all chloride ions originated from the dissociation of trimethylammonium chloride were consumed by Ag⁺ ions from the titrant. The further addition of the titrant caused an increase in the solution's conductivity. The total volume of the AgNO₃ solution added at the endpoint of titration (Fig. 1, V*_{AgNO₃}) was used to calculate DS from Equation 1. The substitution of Chitosan with GTMAC was 77.8 %.

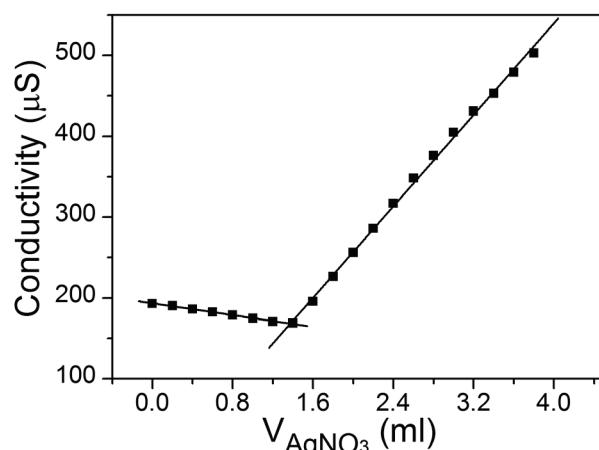


Fig. 1. Conductivity of CMCh as a function of volume of AgNO₃ added.

The construction of CMCh/Hep LbL films

The self-assembly process of CMCh and Hep was observed *in situ* using SPR. Figure 2 presents sensograms of the formation of a six-layer film terminated with Hep, (CMCh/Hep)₃ (line 1), and a seven-layer film terminated with CMCh, (CMCh/Hep)₃CMCh (line 2). The SPR signal increased after each CMCh and hep deposition, demonstrating that both polysaccharides adsorbed in the LbL process. During the rinsing step, the signal initially decreased and then reached a constant level due to desorption of the loosely bound topmost polyelectrolyte molecules. The adsorption of Hep reached saturation after approximately 10 minutes. Interestingly, when CMCh was brought into contact with the Hep-terminating layer, the signal first sharply increased and then continued to decrease. This phenomenon stopped and the SPR signal remained constant by replacing the CMCh solution with PBS. A difference in the resonance wavelength ($\Delta\lambda_{\text{res}}$) in PBS before and after each

adsorption step was plotted versus a number of the deposited layers (Fig. 2D). The $\Delta\lambda_{\text{res}}$ corresponds to the changes in the refractive index of the SPR chip surface and is proportional to the mass of adsorbed/desorbed compounds. The $\Delta\lambda_{\text{res}}$ increases with the increasing number of the layers in an exponential manner. This indicates that the amount of the adsorbed CMCh and Hep in each consecutive adsorption cycle is always higher than the amount of previously adsorbed polyelectrolytes. This observation was confronted with UV-Vis analysis. Figure 3A shows the dependence of absorbance at 510 nm of the FITC-labeled CMCh, CMCh^{FITC}, on a number of the deposited bilayers. The inset presents the UV-Vis spectra of a quartz plate coated up to five CMCh^{FITC}/Hep bilayers. The evolution of the absorbance with the number of the bilayers is well fitted by an exponential growth curve which is in line with the SPR findings.

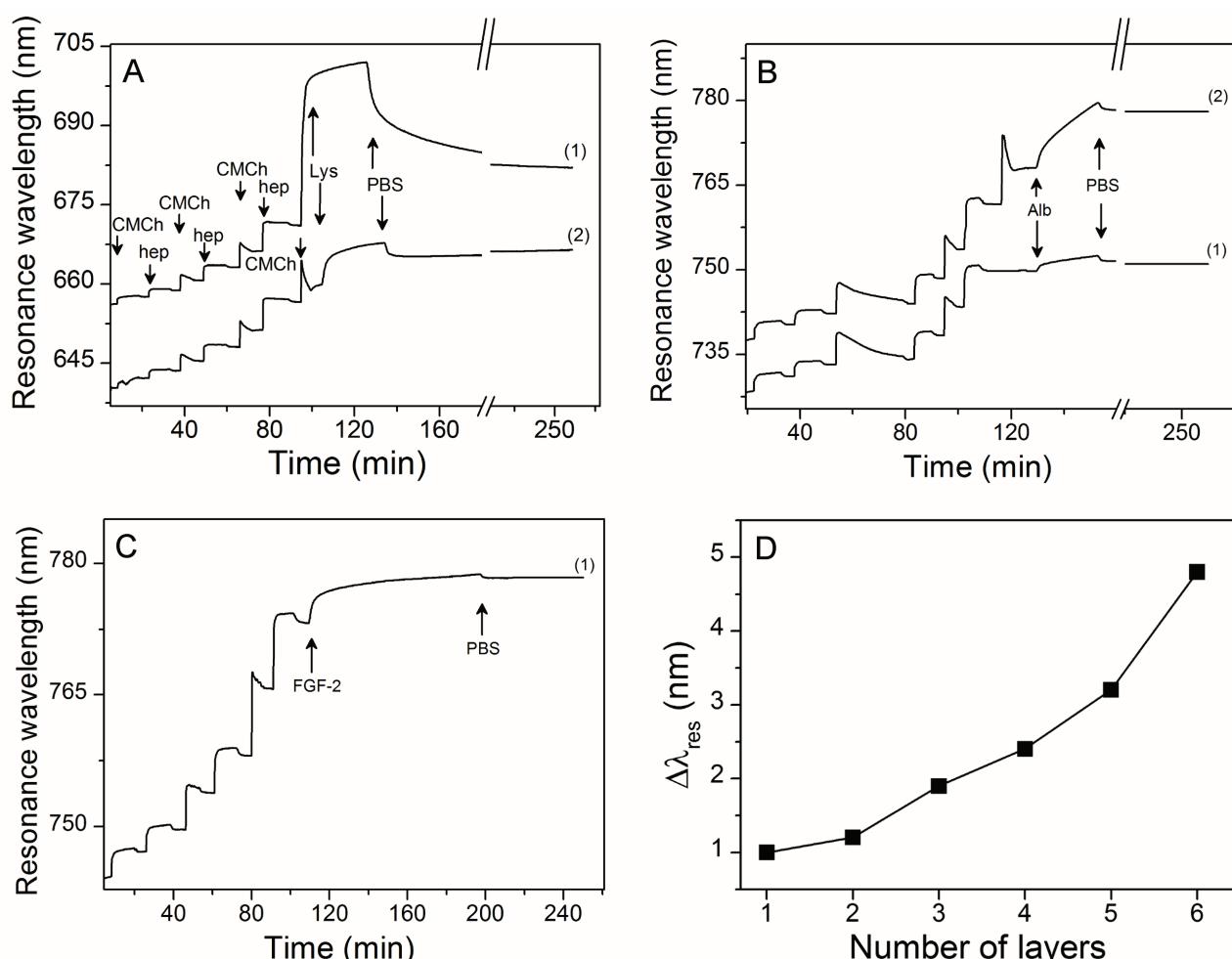


Fig. 2. *In situ* formation of (CMCh/Hep)₃ (1) and (CMCh/Hep)₃CMCh (2) LbL films observed by SPR. Arrows show the replacement of the solutions. The adsorption of Lys (1 mg/ml in PBS, **A**), Alb (1 mg/ml in PBS, **B**), and FGF-2 (100 ng/ml in PBS, **C**). Differences in the resonance wavelength shifts ($\Delta\lambda_{\text{res}}$) as a function of the number of deposited layers (**D**).

The LbL process was also followed by AFM. The topography of dry LbL films composed of 6-, 7- and 23-layers is presented in Figure 4. AFM images of the $(\text{CMCh}/\text{Hep})_3$ and $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ assemblies indicate the presence of characteristic granular structures, ‘islets’, that cover the whole surface of the substrate (Fig. 4D, E). The islets exhibit different sizes; smaller islets appear between the larger ones. Based on the cross-section graphs (Fig. 4A, B), the diameter and height of these islets reach up to 200 nm and 20 nm, respectively. The surface features depended on the terminated layer of LbL films. The particular islets in the Hep-ending LbL film displayed a rough surface (Fig. 4D), whereas in the CMCh-ending LbL film, the islets became more compact, separated from each other by forming distinct edges (Fig. 4E). The morphology of the LbL films significantly changed when more bilayers were deposited. The surface of the $(\text{CMCh}/\text{Hep})_{11}\text{CMCh}$ LbL film exhibited large objects of 800 nm in diameter and about 30 nm in height (Fig. 4C, F). Also, the root mean squared surface roughness, R_q , increased from 6.0 ± 0.1 nm to 9.7 ± 0.8 nm for $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ and $(\text{CMCh}/\text{Hep})_{11}\text{CMCh}$, respectively.

The protein adsorption on CMCh/Hep LbL films

The adsorption of Lys, Alb and FGF-2 on the LbL films was monitored in real time using SPR. The exposition of the Hep- and CMCh-terminated LbL film to Lys solution resulted in a signal shift of 30.8 nm and 7.8 nm, respectively (Fig. 2A). However, when the Lys solution was replaced by PBS, a continuous decay of λ_{res} over 2 h during rinsing with PBS was observed for $(\text{CMCh}/\text{Hep})_3$ (Fig. 2A, line 1). This indicates that about 60 % of the initially loaded Lys into the $(\text{CMCh}/\text{Hep})_3$ assembly was released within 2 h. On the contrary, λ_{res} slightly decreased and remained stable during rinsing with PBS for $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ (Fig. 2A, line 2). Here, only 18 % of the initially adsorbed Lys desorbed over 2 h. Albumin exhibited the opposite behavior when contacted with the LbL films. A signal shift corresponding to Alb adsorption was 4.3 times higher for the $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ as compared to the $(\text{CMCh}/\text{Hep})_3$ film (Fig. 2B). The replacement of the Alb solution to PBS resulted in a 53 % and 13 % release of the initially loaded Alb from the $(\text{CMCh}/\text{Hep})_3$ and $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ films, respectively.

The $(\text{CMCh}/\text{Hep})_3$ film was used to physically capture FGF-2. The SPR sensorgram (Fig. 2C) demonstrates a stepwise increase in λ_{res} during FGF-2

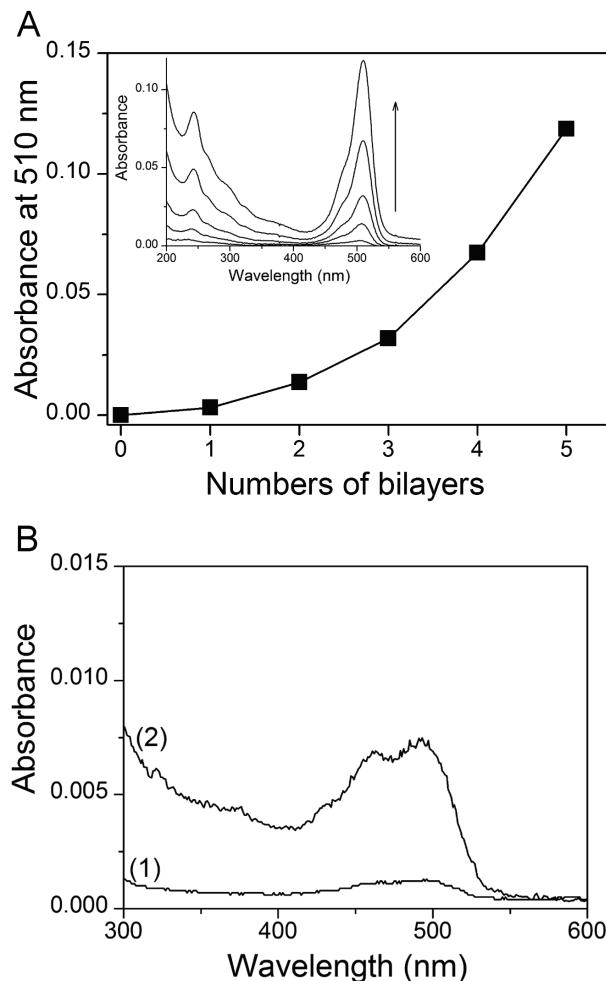


Fig. 3. The dependence of absorbance at 510 nm on the number of $\text{CMCh}^{\text{FITC}}/\text{Hep}$ bilayers (**A**). The inset shows the changes of the $\text{CMCh}^{\text{FITC}}$ absorption spectra during the building of the LbL film. The absorption spectra (**B**) of Lys^{FITC} adsorbed on the $(\text{CMCh}/\text{Hep})_3$ assembly (line 1) and Alb^{FITC} adsorbed on the $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ assembly (line 2).

adsorption. Then a slight decrease of the signal after rising with PBS corresponds to the 7 % release of the initially adsorbed FGF-2.

The surface coverage by the proteins (see Surface analysis) was as follows: the $(\text{CMCh}/\text{Hep})_3$ assembly bound 19.5 ng/cm^2 of Alb and 76.5 ng/cm^2 of FGF-2 whereas the $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ assembly bound 117 ng/cm^2 of Lys and 148.5 ng/cm^2 of Alb. The correct determination of the lysozyme surface density on the $(\text{CMCh}/\text{Hep})_3$ assembly was impossible due to continuous Lys desorption from the film (Fig. 2A, line 1).

Protein adsorption was also studied by UV-Vis spectroscopy. The FITC absorption band indicates the presence of Lys^{FITC} on the $(\text{CMCh}/\text{Hep})_3$ assembly (Fig. 3B, spectra 1) and Alb^{FITC} on the $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ assembly (Fig. 3B, spectra 2). There

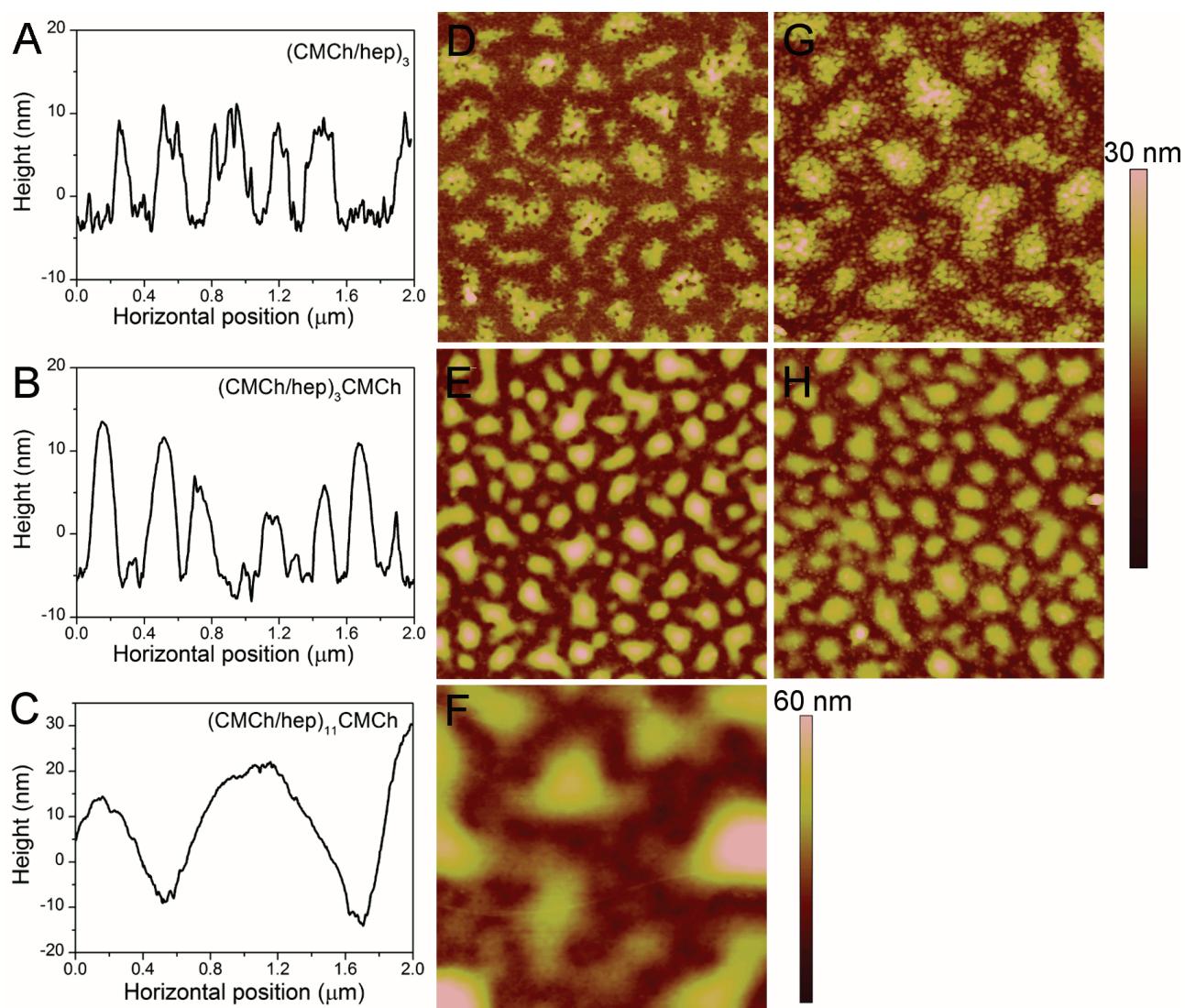


Fig. 4. AFM cross-sectional height and topography of the $(\text{CMCh}/\text{Hep})_3$ (**A**, **D**), $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ (**B**, **E**) and $(\text{CMCh}/\text{Hep})_{11}\text{CMCh}$ (**C**, **F**) assemblies. AFM images of $(\text{CMCh}/\text{Hep})_3$ and $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ assemblies after exposure to lysozyme (**G**) and albumin (**H**). Image size: $2 \times 2 \mu\text{m}$; the z scale: 30 nm or 60 nm shown on the right site of the image.

was no evidence of Lys^{FITC} on $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ and Alb^{FITC} on $(\text{CMCh}/\text{Hep})_3$ assemblies (data not shown).

Changes in the surface topography after the adsorption of Alb and Lys were studied by AFM. Many small dots with a diameter of approx. 10 nm appeared after protein adsorption and they covered both the islets and the space between the islets (Fig. 4G, H). Nevertheless, the surface roughness Rq did not change.

Discussion

In the present work, the quaternized chitosan- and heparin-terminated multilayer films were studied for their ability to adsorb proteins of differing isoelectric points. The buildup of the CMCh/Hep multilayer films was followed by SPR and UV-Vis spectroscopy (Fig. 2

and 3). The results demonstrated that the LbL film grew in an exponential-like fashion (Fig. 2D and 3A). Furthermore, the SPR sensorgram (Fig. 2A, B, C) showed a characteristic ‘peaked’ evolution of λ_{res} when the Hep-terminated film was brought into contact with the CMCh solution. This phenomenon has already been reported in the literature (Richert *et al.* 2004, Picart *et al.* 2001, Lavalle *et al.* 2004) and was explained by diffusion of the polyelectrolyte in and out of the film during the LbL buildup process. Confocal laser microscopy has proved that a fluorescently labeled chitosan migrated inside chitosan/hyaluronan multilayer films during their formation (Richert *et al.* 2004). This, in turn, results in exponential growth of the LbL film.

The AFM images of the LbL films showed the presence of the granular structures in the form of so-

called ‘islets’ which might result from the coacervation of both oppositely charged polyelectrolytes (Fig. 4D, E). These islets coalesced and formed larger sized structures as the number of deposited layers increased (Fig. 4F). Similar topographical features revealed LbL films composed of, e.g. poly(L-lysine) and hyaluronan (Picart *et al.* 2001), chitosan and hyaluronan (Cardoso *et al.* 2016), chitosan and poly(γ -glutamic acid) (Zhao *et al.* 2016). The islets’ formation was explained by the aggregation of the polyelectrolytes at a substrate surface during the deposition cycles. The islets became larger with the increasing number of bilayers which is in line with our observation.

The interaction of the CMCh/Hep films with lysozyme and albumin were studied by SPR, UV-Vis and AFM. The adsorption of those proteins strongly depended on the terminating polysaccharide. Lysozyme ($I_p=11.4$), bearing a net positive charge at pH 7.4, was ionically attracted by the negatively charged Hep-terminated film (Fig. 2A). On the contrary, the negatively charged Alb ($I_p=4.9$) adsorbed effectively on the positively CMCh-terminated film (Fig. 2B). If the topmost polysaccharide layer and protein were like-charged, the adsorption of protein on the film was minimized due to the electrostatic repulsion (Fig. 2A, B). It is evident that the adsorption of Lys and Alb was mainly driven by the electrostatic interactions with the terminal layer of the polysaccharide LbL film. A similar conclusion was drawn by (Hoven *et al.* 2007) who studied the behavior of several proteins on chitosan-based coatings. The negatively charged sulfonate-functionalized chitosan films suppressed the adsorption of the same charged proteins, i.e. albumin and fibrinogen, and electrostatically attracted the positively charged ribonuclease and lysozyme. Beykal *et al.* (2015) indicated that the electrostatic interactions played a key role in the adsorption of Alb to charged and uncharged gold surfaces. An increased mass adsorption was observed when the protein and the surface were oppositely charged, which is also confirmed by our results.

Based on the SPR data, protein adsorption also occurred on the alike-charged surface although to a lesser extent (Fig. 2A, B). It is worth noting that a globular protein like Lys and Alb, cannot be considered as a single charged point, since the surface of protein consists of positively and negatively charged domains that can interact with the LbL film (Cooper *et al.* 2005). Further, as observed in the AFM images (Fig. 4), both polysaccharides form complexes upon assembling on the

surface. Such polyelectrolyte complexes destroy the layer structure of the film and can undergo dynamic structural changes resulting from CMCh diffusion within the film. Thus, regardless of the last deposited layer, both polysaccharides can be accessible at the surface, and consequently, they can electrostatically interact with the protein molecules to some extent.

Both proteins demonstrated almost irreversible binding to the positively charged $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ film (Fig. 2A, B, line 2). By contrast, the $(\text{CMCh}/\text{Hep})_3$ film showed a reversible loading of Lys and Alb; the proteins were significantly released over 2 h of rinsing with PBS (Fig. 2A, B, line 1). This suggests that heparin exhibits a low binding affinity to both proteins. As reported, Lys binds to heparin with a dissociation constant (K_d) of 1.3×10^{-5} M, that is 100-fold higher than K_d determined for unsulfated glycosaminoglycans (GAGs), e.g. hyaluronic acid (Vandamme *et al.* 1994). This indicates that the sulfate groups of GAGs were not dominant in the interaction with the positively charged Lys.

A number of studies have shown the applicability of LbL films as reservoirs of fragile biomolecules and therapeutics (Kumorek *et al.* 2015, Monge *et al.* 2015). Based on the results obtained from the interactions of Lys with the LbL films, the $(\text{CMCh}/\text{Hep})_3$ assembly was selected to physically capture the positively charged biomolecule FGF-2. The exposure of the Hep-terminated film to the FGF-2 solution yielded 76.5 ng/cm^2 of the stably immobilized FGF-2; only 7 % of the initially loaded FGF-2 was released after a 2 h rinsing with PBS. In fact, FGF-2 possesses strong binding affinity to sulfated GAGs, such as heparin (Faham *et al.* 1996). Therefore, not only the electrostatic attractions, but also H-bond formation with electronegative atoms of Hep contributed to the FGF-2 adsorption process and stabilized the FGF-2-heparin complex (Schlessinger *et al.* 2000). This could explain the different behavior of both positively charged FGF-2 and Lys on the Hep-terminated LbL film.

The AFM imaging also proved the proteins adsorption on the LbL films. The small granules originating from the immobilized Lys and Alb were clearly visible on the whole surface of the $(\text{CMCh}/\text{Hep})_3$ and $(\text{CMCh}/\text{Hep})_3\text{CMCh}$ LbL films (Fig. 4G, H). Previously, Ladam *et al.* (2000) reported that albumin self-aggregated and formed a dense multimolecular layer on the positively charged polystyrene-sulfonate/polyallylamine LbL film. Similarly, Gergely *et*

al. (2004) found the formation of a thick layer of albumin exceeding the dimension of the native protein on the poly(L-lysine)/poly(glutamic acid) LbL film. However, from the AFM images, it is difficult to conclude if the proteins adsorbed in a monolayer or multilayer mode.

Conclusions

Herein, we studied the behavior of model proteins on the surface of the self-assembled quaternized chitosan (CMCh)/heparin multilayer film. Polysaccharide films forming *via* the layer-by-layer (LbL) method displayed an exponential growth at a neutral pH. The CMCh- or hep- terminated LbL films interacted with albumin and lysozyme regardless of the charge of the outermost layer. However, since protein adsorption was driven mainly by the electrostatic interaction, the amount of the adsorbed protein depended on the terminating polysaccharide. Both proteins were almost irreversibly

bound to the positively charged CMCh-terminated LbL film, (CMCh/Hep)₃CMCh. Whereas, the Hep-terminated film, (CMCh/hHep)₃, was more resistant to the albumin adsorption and demonstrated a relatively weak interaction with lysozyme. On the other hand, the (CMCh/Hep)₃ LbL film effectively bound FGF-2. Owing to that strong binding affinity to growth factors, the hep-terminated LbL film can be applied as a biofunctional surface coating bearing therapeutic proteins, such as FGF-2, for biomedical applications.

Conflict of Interest

There is no conflict of interest.

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