Early Changes of Brain Perfusion After Subarachnoid Hemorrhage – the Effect of Sodium Nitroprusside

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Summary
Causes of early hypoperfusion after subarachnoid hemorrhage (SAH) include intracranial hypertension as well as vasoconstriction. The aim of the study was to assess the effect of intracerebroventricular (ICV) administration of sodium nitroprusside (SNP) on early hypoperfusion after SAH. Male Wistar rats (220-240 g) were used, SAH group received 250 μl of fresh autologous arterial blood into the prechiasmatic cistern; sham-operated animals received 250 μl of isotonic solution. Therapeutic intervention: ICV administration of 10 μg SNP; 5 μl 5 % glucose (SNP vehicle) and untreated control. Brain perfusion and invasive blood pressure were monitored for 30 min during and after induction of SAH. Despite SNP caused increase of perfusion in sham-operated animals, no response was observed in half of SAH animals. The other half developed hypotension accompanied by brain hypoperfusion. There was no difference between brain perfusion in SNP-treated, glucose-treated and untreated SAH animals during the monitored period. We did not observe expected beneficial effect of ICV administration of SNP after SAH. Moreover, half of the SNP-treated animals developed serious hypotension which led to brain hypoperfusion. This is the important finding showing that this is not the option for early management in patient after SAH.

Key words
Subarachnoid hemorrhage • Sodium nitroprusside • Nitric oxide • Early brain injury • Rat

Introduction
Non-traumatic aneurysmal subarachnoid hemorrhage (SAH) causes both early and delayed changes of brain perfusion. The early brain injury (EBI) develops within first 72 h after aneurysm rupture (Cahill et al. 2006) and it is a major cause of mortality in such cases (Broderick et al. 1994). The arterial wall rupture causes extravasation of arterial blood into subarachnoid space under high pressure, leading to rapidly increased intracranial pressure (ICP) and reduced cerebral perfusion pressure (CPP) and cerebral blood flow (CBF) (Cahill et al. 2006, Sehba et al. 2012). Even animal models show that the early hypoperfusion is related not only to intracranial hypertension, but also to vasoconstriction associated with arterial blood present in subarachnoid space; reduction of CBF independent from increased ICP or decreased CPP was observed (Bederson et al. 1998, Schubert et al. 2009).

One of the mechanisms of early vasoconstriction is impairment of nitric oxide (NO)-mediated vasodilation. Decreased levels of NO metabolites were observed as early as 10 min after SAH induction (Sehba et al. 2000). The NO deficiency is probably caused by scavenging of NO (Sehba et al. 2012, Schwartz et al. 2000a,b). In physiologic conditions, the activity of NO is terminated by reaction with oxyhemoglobin which forms methemoglobin and nitrate (Helms and Kim-Shapiro 2013). Oxyhemoglobin in subarachnoid space is
considered to be one of the major NO scavengers after SAH and thus it contributes to the early vasoconstriction (Li et al. 2016, Sehba et al. 2012).

Due to this mechanism, it is reasonable to consider NO donors for treatment of early vasoconstriction. In previous studies the cerebral vessels did respond to external administration of NO (Sehba et al. 1999, Sehba et al. 2007).

One of the NO donors studied in conditions of delayed cerebral vasospasm is sodium nitroprusside (SNP). The desired effect of SNP in the treatment of vasospasm is reached by release of NO at the adventitial side of brain circulation from the SNP molecule. Such a replacement of NO in the vascular wall attenuates the vasospasm after intrathecal SNP administration (Thomas et al. 1999). Beneficial effect of SNP was observed in several experimental (Egemen et al. 1993, Vatter et al. 2007) and clinical studies (Pachl et al. 2005, Raabe et al. 2002).

As far as we are aware, there were no studies with administration of sodium nitroprusside conducted under conditions of early brain injury. We hypothesized that intracerebroventricular (ICV) administration of SNP could be used as an effective treatment of the early vasospasm immediately after SAH. Because SNP is a potent vasodilator, the ICV route could help to reduce systemic side effects seen after intravenous administration, preserve its vasodilating effect on brain circulation and increase availability of NO in brain tissue.

Materials and Methods

All procedures were performed in accordance with the Ethical Guidelines of the Third Faculty of Medicine, Charles University, Prague, Czech Republic. They were in accordance with the Guidelines of the Animal Protection Law of the Czech Republic, which correspond with European Guidelines on Laboratory Animal Care. Special care was taken to minimize animal suffering.

Adult male Wistar rats (AnLab, Czech Republic; 220-240 g) were used. The animals were housed in cages by four under a 12-h light/dark cycle, with food and water ad libitum.

Surgical preparation of animals and induction of non-traumatic SAH

All procedures were conducted in deep general anesthesia of the soft tissues of the head (trimecaine 1 % 0.3 ml).

Femoral artery was cannulated to obtain blood sample for SAH induction as well as to allow invasive monitoring of arterial blood pressure. Mean arterial blood pressure (MAP) was calculated as 2/3 of diastolic pressure + 1/3 of systolic pressure. Rectal temperature was monitored and maintained at 37 °C during the whole procedure.

The skull of the animals was exposed from soft tissues. Left lateral ventricle was cannulated for ICV administration access (Brain Infusion Kit 2, Alzet, USA). The catheter was inserted -2 mm and 2 mm left from bregma and 3 mm deep from the skull surface into the cerebral ventricle (Paxinos and Watson 2013).

The skull above right hemisphere was mechanically thinned to allow the measurement of brain perfusion. Prechiasmatic cistern injection model of subarachnoid hemorrhage was used (Prunell et al. 2002).

Briefly, a small borehole was drilled 7 mm rostral from the bregma. A needle with 26G cannula was inserted into prechiasmatic cistern at the 30° angle anteriorly. In the SAH groups, 250 μl of fresh non-heparinized autologous arterial blood (aBLD) was injected during 15 s immediately after withdrawal from femoral artery; sham animals received 250 μl of isotonic solution (ISOSOL) of body temperature.

Experimental groups

Sixty four animals were divided into six groups: 1) SAH-induced animals (SAH; n=12); 2) SAH + ICV administration of sodium nitroprusside (SAH-SNP; n=18); 3) SAH + ICV glucose 5 % (SAH-glc; n=10); 4) sham-operated animals (sham, n=8); 5) sham + ICV SNP (sham-SNP; n=8); and 6) sham + ICV glucose 5 % (sham-glc; n=8).

The SAH-SNP animals received 10 μg of sodium nitroprusside (Sigma-Aldrich) in 5 μl of 5 % glucose solution 3 min after SAH induction. If secondary decrease of perfusion occurred after previous increase (decrease >15 % of perfusion after reaching peak level), second dose of SNP solution was administered. The solution of SNP was meticulously protected from light during preparation as well as during the whole experiment.

The SAH-glc group received 5 μl of 5 % glucose solution according to the same time pattern, the SAH group remained untreated. The corresponding sham-operated groups received the treatment in the same pattern.
Evaluation of changes in the perfusion of cerebral cortex after SAH

Changes in the perfusion of cerebral cortex were assessed using Laser speckle-contrast analysis (LASCA). For detection and evaluation of changes, PeriCam PSI HR with PimSoft software (Perimed, Sweden) was used. The region of interest (ROI) for perfusion measurement was placed above the right hemisphere and it was limited by sagittal suture and lateral ridge, caudally by lambdoid suture and exceeded coronal suture for 3 mm in rostral direction. The tissue perfusion was measured in perfusion units (PU) standardized by manufacturer. Because the PUs cannot be converted to ml/min/100 g, changes in perfusion were described in percentage of PU deviation from baseline record. The baseline levels were obtained from measurement performed in each animal during 1 min before induction of SAH. Framing speed was set as 1 image/s and the mean perfusion in the designated ROI was used for evaluation.

Statistical analysis

GraphPad Prism 6 (GraphPad Software, Inc., USA) was used for statistical evaluation. Changes in perfusion between groups were compared using two-way ANOVA for repeated measures with Bonferroni’s post-test. Following changes in the perfusion of animals with SAH were analyzed using t-test: maximum values of perfusion which followed an initial drop after application of blood into prechiasmatic cistern; minimum values of perfusion decreased secondary after reaching maximum; as well as changes in mean arterial blood pressure and perfusion in groups treated with SNP.

Level of statistical significance was set to \( p<0.05 \). All data are presented as mean ± SEM.

Results

Changes of brain perfusion

Intracisternal administration of both aBLD and ISOSOL was associated with significant decrease of brain perfusion.

The perfusion in the SAH group decreased to 23.3±4.5 % (mean ± SE) of baseline values during the injection of aBLD. Three of 12 studied animals developed profound hypoperfusion leading to death within 5 min after the injection. In the remaining 9 animals, the perfusion recovered slowly after initial drop. Compared to control, the SAH group showed prolonged return towards baseline values and significantly reduced perfusion within first 5 min after SAH induction (Fig. 1).

Fig. 1. Perfusion changes in SAH and sham-operated group. The return of perfusion towards baseline levels was prolonged in SAH group. The perfusion was significantly reduced during and first 7 min after SAH induction in SAH animals and at the time of intracisternal injection and 6-8 min and 10-12 min in sham-operated animals (+, \( # p<0.05 \), ++ \( p<0.01 \), +++ \( p<0.001 \), compared to baseline values). Compared to sham-operated animals, the perfusion was significantly lower first 5 min after SAH induction (* \( p<0.05 \), ** \( p<0.01 \), *** \( p<0.001 \)).

The perfusion changes in SAH animals showed biphasic time-course in 8 of 9 animals. After the injection, the perfusion recovered slowly reaching its peak in 14 min 22 s ± 2 min 10 s and culminated at the level of 113±5.5 % of baseline values. This was followed by another significant decrease (p<0.001 and p<0.01, compared to peak and baseline values, respectively) in 24 min 52 s ± 1 min 36 s at the level 86.4±3 % of baseline after which the perfusion increased gradually. Similar pattern was observed in SAH-glc group, but only
in half SAH-SNP animals.

In the sham group, the initial drop in perfusion was followed by short period of hyperperfusion after which the perfusion returned toward baseline level. The perfusion was significantly reduced during the time of intracisternal injection and 6-8 min and 10-12 min in sham-operated animals, compared to baseline values (Fig. 1).

The effect of intracerebroventricular administration of sodium nitroprusside

In the study group assigned for SNP treatment after SAH, one of 18 animals died before administration of SNP and 2 died after SNP administration. The response to SNP showed two different patterns. Eight animals showed no change of MAP and the time-course of brain perfusion did not differ significantly from SAH and SAH-glc groups. The other 7 animals developed serious systemic hypotension (drop in MAP up to 50% of pre-dosing values) which was accompanied with decrease of brain perfusion (Fig. 2). In this subgroup, the brain perfusion was significantly lower 1 min after the SNP administration, compared to non-hypotensive animals (Fig. 3).

Both normotensive and hypotensive animals showed increase of perfusion above the pre-dosing levels 3 min after the first SNP injection. Comparison with SAH-glc and SAH groups nevertheless shows that this increase of brain perfusion occurs in this time period independently from the SNP treatment.

**Fig. 2.** Perfusion and MAP changes in SAH-SNP group after the first SNP dose. In 7 of 15 animals, SNP administration 3 min after SAH induction caused decrease of MAP, which corresponded with significant decrease of brain perfusion. Significant increase of brain perfusion above pre-dosing levels was observed 3 min after the injection; nevertheless this increase followed trends observed even in SAH and SAH-glc groups (*, + p<0.05, **, ++ p<0.01, ***, +++ p<0.001, compared to pre-dosing levels). In contrast to other figures, baseline is set as pre-dosing level.

**Fig. 3.** Perfusion changes after SNP administration. Administration of SNP 3 min after SAH induction was followed by arterial hypotension and brain hypoperfusion in 7 of 15 animals. Brain perfusion was significantly lower 1 min after SNP injection in those animals which developed hypotension, compared to normotensive animals (* p<0.05).
Eight SNP-treated animals developed secondary decrease of perfusion after previous reaching of peak values. This decrease corresponded with biphasic time-course of changes in perfusion seen in SAH group. These animals received another ICV dose of 10 μg of SNP after secondary decrease between 11 and 25 min. Sequentially a mild but significant decrease of MAP developed with insignificant decrease of perfusion. The perfusion increased 2 min after the second SNP injection and the raise above the baseline was significant 3 min 20 s after the injection (compared to pre-dosing values). This increase of perfusion however did not reach the values observed in sham-SNP animals.

Despite mild increase of the perfusion after the second dose of SNP, we did not observe any statistically significant increase of brain perfusion in SNP-treated animals, compared to SAH and SAH-glc group during the monitored period (Fig. 4). Also no difference was observed between animals which received 1 or 2 doses (data not shown).

The effect of intracerebroventricular administration of 5 % glucose

One of 10 SAH-glc group animals died before and 2 died after the glucose injection. In 7 animals, the first dose of glucose did not cause any change of either mean arterial pressure or brain perfusion, compared to SAH group.

All 7 animals received second dose of glucose due to decrease of the perfusion after reaching the peak. The MAP remained unaffected after the injection. The injection of glucose caused transient increase of brain perfusion which was significant at 1.5 min (compared to pre-dosing values). In contrast to the SNP treated animals, the increase was observed within first 2 min after the injection. The perfusion in SAH-glc was significantly (p<0.05) higher between 40 s and 1 min, compared to SAH-SNP group.

Sham-operated animals

Animals in the sham-operated groups received ISOSOL instead of aBLD into the prechiasmatic cistern. Three minutes later, ICV injection of 5 μl of either SNP or G5 % solution was administered; one group received no treatment. Intracerebroventricular injection of SNP led to significant increase in brain perfusion above pre-dosing levels in 7 of 8 animals; this effect persisted till the end of the monitored period. In 1 animal, the increase of brain perfusion was achieved after repeated dose of SNP. The ICV injection of SNP was followed by mild decrease of MAP, nevertheless the brain perfusion increased in the same time. ICV injection of 5 % glucose solution did not lead to any change of the perfusion (Fig. 5).

Discussion

Our study is the first to test the effect of intracerebroventricular administration of SNP in conditions of early hypoperfusion after subarachnoid hemorrhage.

Markedly reduced cortical perfusion is one of the initial consequences of SAH. The primary decrease of perfusion after intracisternal injection of both arterial...
Nitric oxide donors treatment was studied in several experimental (Egemen et al. 1993, Vatter et al. 2007) and clinical (Pachl et al. 2005, Raabe et al. 2002) trials in conditions of delayed cerebral vasospasm.

Nitric oxide donors for prevention or treatment of EBI were used only in experimental settings: administration of an NO donor S-nitrosoglutathion (GSNO) was successfully used to increase CBF and preserved vascular wall integrity briefly after SAH induction (Sehba et al. 1999, Sehba et al. 2007). Another more common and affordable NO donor is sodium nitroprusside. The dose used in the present study was calculated according to the doses used in previous laboratory and clinical experiments and was adjusted on rats’ body weight and volume of cerebrospinal fluid. The dose 10 μg of SNP for bolus administration was proven as most effective in preliminary experiments.

In the present study, no favorable effect was observed after administration of SNP 3 min after SAH simulation. Half of the SNP-treated animals developed brain hypoperfusion instead. This can be related to either arterial hypotension or intracranial hypertension. Disturbance of autoregulation of brain perfusion is a frequent feature of intracranial pathologies. The brain perfusion depends on CPP and MAP in conditions of impaired autoregulation (Lang et al. 2003) and thus arterial hypotension after SNP injection can lower the brain perfusion substantially. Intracranial hypertension can also contribute to further reduction of perfusion. Different methods of SAH induction can produce
different ICP profiles (Schwartz et al. 2000a,b); according to our previous yet unpublished findings, intracranial pressure 3 min after SAH induction is approx. 15-20 mm Hg in this model; similar results were obtained in another studies (Prunell et al. 2003). Injection of additional fluid volume (e.g. SNP) can increase ICP significantly in conditions of reduced brain compliance (Marmarou et al. 1975). Nevertheless this is doubted by fact that we observed no further brain perfusion decrease immediately after ICV injection of glucose solution.

Biphasic time course of the perfusion changes was observed in SAH animals. Reason for this pattern is uncertain. It can be related to the method of measurement of perfusion which includes both microcirculation and large pial vessels. Effect of SAH on vasoconstriction can differ between macro- and microcirculation (Sehba et al. 2012). The decrease of perfusion after previous peak can also reflect the changes in NO concentration in brain circulation (Sehba et al. 2000).

Mild increase of perfusion was observed after the second dose of SNP, which was administered after the second drop of perfusion. This increase did not reach the levels observed in sham-SNP group and there was no significant difference in perfusion between SAH-SNP group and SAH or SAH-glc groups. In contrast, in sham-SNP group, the ICV administration of SNP led to prompt increase of brain perfusion which persisted till the end of the monitored period. This discrepancy in the effect of SNP between SAH and sham-operated group can have more reasons. Lack of effect of ICV administration of SNP can be caused by impaired distribution of the drug in subarachnoid space. The arteries of the circle of Willis are coated with the coagulum which can prevent from diffusion of the vasodilator and thus the concentration of NO in the vessel wall does not reach adequate level (Pluta et al. 2009).

Another possible explanation is that the dose leading to perfusion increase in sham-SNP animals can be insufficient in situation of NO depletion and scavenging which occurs after SAH. This can be supported by the finding of mild increase of brain perfusion after the second dose of SNP, which did not reach the levels of perfusion observed in sham-SNP animals. Variable response on SNP administration, mainly after the first dose of SNP, is consistent with marked interindividual variability which was observed also in clinical studies (Agrawal et al. 2009, Pachl et al. 2005).

So far, SNP was successfully used in experiments with delayed vasospasm – dilation of spastic vessels after SNP administration was observed in experiments on isolated arteries (Vatter et al. 2007) as well as in animal (Egemen et al. 1993) and clinical studies (Pachl et al. 2005, Raabe et al. 2002). The observed effect of intrathecal SNP administration in conditions of delayed cerebral vasospasm can be explained by the degradation of blood clot and improvement of SNP penetration into the target structures. The velocity of clot resolution was estimated as 10.8 % of clot volume per day (Naff et al. 2001) and significant proportion of the hematoma is degraded at the time of occurrence of delayed cerebral vasospasm. Hemoglobin degradation products attribute to delayed vasospasm (Clark and Sharp 2006). Therefore, difference in mechanisms of vasoconstriction in the early and delayed phase after SAH can explain different finding in our model after SNP administration.

Hypotension after ICV injection of SNP was observed in significant proportion of study animals. This side effect was reported also in clinical studies (Agrawal et al. 2009). It can be attributed to direct effect on brain structures responsible for arterial blood pressure regulation – e.g. organum vasculosum laminae terminalis (Chang et al. 2001, Lin et al. 1999). Previous study demonstrated decrease of MAP after ICV injection of SNP (Lin et al. 1999), nevertheless the drop of MAP was less striking after dose equal to the dose used in our study.

The present study has some limitations, as well. Concentration of oxyhemoglobin in arterial blood can potentially affect the extent of the vasospasm, however neither hematocrit, nor SpO2 were measured in our study. The brain perfusion might also be influenced by anesthesia. Increase of CBF was described after ketamine; nevertheless the effect of ketamine on CBF was prevented by co-medication with midazolam (Strebel et al. 1995).

Conclusions

Intracerebroventricular administration of sodium nitroprusside did not produce expected effects in the conditions of early brain hypoperfusion in prechiasmatic cistern injection model of subarachnoid hemorrhage. It was also complicated with serious side effects – systemic hypotension that was connected with further decrease of cerebral perfusion pressure. The important outcome of the present study is that intracerebroventricular
administration of sodium nitroprusside is not the option for early management in patient after subarachnoid hemorrhage.

**Conflict of Interest**
There is no conflict of interest.

**References**


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