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20 Summary

21 Using continuous-wave near-infrared spectroscopy (NIRS), this study compared three
22 different methods, namely the slope method (SM), the amplitude method (AM), and the
23 area under the curve (AUC) method to determine the variations of intramuscular
24 oxygenation level as a function of workload. Ten right-handed subjects (22 ± 4 years)
25 performed one isometric contraction at each of three different workloads (30%, 50% and
26 90% of maximal voluntary strength) during a period of twenty seconds. Changes in
27 oxyhemoglobin ($\Delta[\text{HbO}_2]$) and deoxyhemoglobin ($\Delta[\text{HHb}]$) concentrations in the
28 superficial flexor of fingers were recorded using continuous-wave NIRS. The results
29 showed a strong consistency between the three methods, with standardized Cronbach
30 alphas of 0.87 for $\Delta[\text{HHb}]$ and 0.95 for $\Delta[\text{HbO}_2]$. No significant differences between the
31 three methods were observed concerning $\Delta[\text{HHb}]$ as a function of workload. However,
32 only the SM showed sufficient sensitivity to detect a significant decrease in $\Delta[\text{HbO}_2]$
33 between 30% and 50% of workload ($p < 0.01$). Among these three methods, the SM
34 appeared to be the only method that was well adapted and sensitive enough to determine
35 slight changes in $\Delta[\text{HbO}_2]$. Theoretical and methodological implications of these results
36 are discussed.

37

38 Keywords: NIRS, muscle hemodynamics, linear slope, amplitude, area under the curve.

39

40 **Introduction**

41 Since the end of the 1980s, continuous-wave near-infrared spectroscopy (NIRS) has been
42 increasingly used as a noninvasive technique for investigating local muscle oxygenation
43 changes during exercise or rest (Ferrari et al., 1997; Ferrari et al., 2011). The validity of
44 NIRS has been examined by different studies (Sako et al., 2001; Van Beekvelt et al.,
45 2001), and the results obtained using this technique are in agreement with those obtained
46 by other imaging techniques such as Phosphorus Magnetic Resonance Spectroscopy (P-
47 MRS) (Sako et al., 2001) and blood gas analysis (Van Beekvelt et al., 2001). Several
48 studies have also examined the reproducibility of this optical imaging technique (Van
49 Beekvelt et al., 2002; Celie et al., 2012; Lacroix et al., 2012). In this way, agreement was
50 observed between measurements of oxygen consumption of the superficial flexor of
51 fingers performed on three different days (Van Beekvelt et al., 2002). Based on intra-class
52 correlation coefficients (ICC), a high reproducibility of data derived from NIRS was
53 shown ($ICC = 0.85 - 1$, Lacroix et al., 2012). Furthermore, Celie et al. (2012) also
54 confirmed the reproducibility of this optical technique, particularly when the intensity of
55 the muscular contraction was increased. Beside its reproducibility, there has been shown a
56 strong relationship between NIRS signals and electromyography data during static and
57 sinusoidal isometric exercises of the biceps brachii muscle (Felici et al., 2009). However,
58 few studies have specifically examined the sensitivity of NIRS to measure muscle oxygen
59 consumption as a function of workload.

60 At the muscle level, near infrared light is absorbed by hemoglobin and myoglobin (Ferrari
61 et al., 2011). Because these two chromophores have identical spectral characteristics, it is
62 impossible to distinguish their respective light absorption (Binzoni et al., 1999; Van
63 Beekvelt et al., 2001; Bhambhani, 2004). As the investigation of muscular oxidative

64 metabolism is independent of the oxygen source, hemoglobin is then the term used to name
65 the two chromophores (Lacroix et al., 2012). In general, the main recorded parameters
66 using NIRS to study muscular oxidative metabolism are the following: (1) changes in
67 oxyhemoglobin concentrations ($\Delta[\text{HbO}_2]$) and deoxyhemoglobin concentrations ($\Delta[\text{HHb}]$);
68 (2) changes in total hemoglobin concentrations ($\Delta[\text{HbT}]$); and (3) muscle oxygen
69 saturation (SmO_2) (Ferrari et al., 2011).

70 To determine the level of change in hemodynamic parameters, previous studies have used
71 different methods such as the slope method (SM) (Quaresima et al., 2001), the amplitude
72 method (AM) (Celie et al., 2012) and the area under the curve (AUC) method (Manfredini
73 et al., 2009). The SM consists of calculating, in the entire recorded signals window, a
74 linear regression to obtain the slope coefficient, which indicates the magnitude and
75 direction of the hemodynamic parameters (e.g., $\Delta[\text{HbO}_2]$, $\Delta[\text{HHb}]$). The AM consists of
76 subtracting the level obtained during a resting state (typically the last 10 seconds of the rest
77 period) from an activation period (typically the last 10 seconds of the contraction) after
78 reaching a plateau for each trial. To determine the amplitude of changes of the
79 hemodynamic parameters, the AUC method integrates the surface obtained under the curve
80 of the hemodynamic parameter changes during the entire recorded signal window.

81 As stated above, few studies have used NIRS to investigate muscle oxygen consumption
82 during active contraction, and those studies have used different quantification methods
83 (Quaresima et al., 2001; Celie et al., 2012; Manfredini et al., 2009). Among these studies,
84 only Celie et al. (2012) used different workloads in their protocol. Their results indicate
85 that the muscle's hemodynamic response (increase in $\Delta[\text{HHb}]$ and decrease in $\Delta[\text{HbO}_2]$)
86 appears somewhat proportional to workload. However, these authors did not specifically
87 address this issue in their study. To our knowledge, no study has yet examined the
88 concurrent validity of these three methods (SM, AM, AUC) or examined whether they

89 show the same sensitivity to changes in workload. Accordingly, the aim of the present
90 study was to examine, in the same protocol involving the same participants, the sensitivity
91 of these three methods to determine slight changes in hemodynamic parameters during an
92 incremental isometric handgrip exercise.

93

94 **Material and methods**

95 Participants

96 Ten right-handed, healthy Caucasian adults (5 men and 5 women) participated in our
97 study. Their mean \pm SD age and Body Mass Index (BMI) were 22 ± 4 years and $21.11 \pm$
98 2.4 kg/m^2 , respectively. Because of potential effects of subcutaneous fat on NIRS signals
99 (McCully & Hamaoka, 2000), we recruited non-obese subjects whose BMI was no more
100 than 25 kg/m^2 (range $17\text{-}24 \text{ kg/m}^2$). Indeed, subcutaneous fat greatly influences the NIRS
101 signal intensity which must pass through the muscle (Hamaoka et al., 2011). All the
102 participants gave their written informed consent to participate in the study, which complied
103 with the declaration of Helsinki for human experimentation.

104

105 Experimental design

106 Handgrip strength was measured using a digital strain-gauge dynamometer (Takei TK 200,
107 Takei Scientific Instruments, Tokyo, Japan) with an accuracy of $\pm 2\text{kg}$. To standardize the
108 muscle location for all participants, the distance (D) between the medial epicondylus
109 humerus and the processus coronoideus ulnae was measured. The transmitter optode was
110 then positioned at a distance equal to $1/3 \text{ D}$ from the medial epicondylus. The receiving
111 optode was positioned laterally, four cm from the transmitter optode, allowing
112 measurement of muscle oxygenation of the flexor digitorum superficialis. The participants
113 were seated in front of a table with their non-dominant upper limb along the body and their

114 right hand in supination so that their forearm formed an angle of approximately 130° with
115 their arm. The width of the handle was adjusted to the size of the hand to ensure that the
116 middle phalanx rested on the inner handle. The participants were allowed to perform one
117 test trial. Then, after a complete 30-second period of rest, the participants performed one
118 isometric contraction at each of three different workloads: 30%, 50% and 90% of maximal
119 voluntary strength (MVS). Isometric contraction was chosen because this form of
120 contraction is more prevalent in the studies (Quaresima et al., 2001; Muthalib et al., 2010;
121 Celie et al., 2012) and minimizes noise due to movements. The duration of each isometric
122 contraction was 20 seconds with a rest period equal to 60 seconds between each
123 contraction.

124 A continuous-wave NIRS (Oxymon Mk III, Artinis Medical Systems^{BV}, Zetten, The
125 Netherlands) was used to measure the changes in concentration of oxygenated hemoglobin
126 ($\Delta[\text{HbO}_2]$), deoxygenated hemoglobin ($\Delta[\text{HHb}]$) and total hemoglobin ($\Delta[\text{HbT}]$), with a
127 sampling rate set at 10 Hz (Quaresima et al., 2001; Muthalib et al., 2010). This device
128 measures only hemodynamic relative changes and may be relatively sensitive to movement
129 artifacts (Wolf et al., 2007). This constraint was limited by using isometric contractions.
130 The measurement of changes in concentration involves the determination of optical
131 densities of two wavelengths (857 nm and 764 nm) in the near-infrared range once they
132 have passed through the muscle. The optical density of these two wavelengths, which are
133 absorbed by oxygenated hemoglobin (HbO_2) and deoxygenated hemoglobin (HHb),
134 respectively, are converted to concentrations of oxygenated hemoglobin [HbO_2] and
135 deoxygenated hemoglobin [HHb] by the modified Beer-Lambert law (Villringer &
136 Chance, 1997 - see Eq. [1]):

$$137 \quad A = \varepsilon \times c \times d \times \text{DPF} + G \quad [1]$$

138 Where A is the absorption of light expressed as optical density, ϵ is a specific extinction
139 coefficient, c is the chromophore concentration, d is the traveled distance, DPF is the
140 differential path length factor and G is the loss of signal due to light scattering.

141

142 Processing of NIRS data

143 NIRS data were analyzed using a commercially available software (Oxysoft, Artinis
144 Medical Systems^{BV}, Zetten, The Netherlands). A low-pass filter with a cutoff frequency of
145 0.7 Hz was used to remove the heart rate signal. Then, the changes in hemodynamic
146 parameters at each workload (30%, 50%, and 90% of MVS) were calculated using the SM,
147 AM and AUC methods. Concerning the SM, a linear regression technique was applied to
148 the 20-second time-series data to obtain the slope coefficient. For the AM, we subtracted
149 the level obtained at the resting state (mean of the last 10 seconds of the rest period) from
150 the one of the activation period (mean of the last 10 seconds of the isometric contraction).
151 Finally, for the AUC, we determined the area under the curve of each hemodynamic
152 parameter for the 20-second isometric contraction.

153

154 Statistical Analysis

155 Statistical analyses were performed using STATISTICA software version 7. The
156 assumption of data normality and homogeneity of distributions was assessed using
157 Kolmogorov-Smirnov and Levene tests, respectively. A standardized Cronbach alpha was
158 computed to measure the consistency between the three methods. To examine the
159 sensitivity of each method as a function of workload, separate ANOVAs with 3 repeated
160 measures (30% vs 50% vs 90%) on $\Delta[\text{HbO}_2]$ and $\Delta[\text{HHb}]$ were first conducted. Second, to
161 compare the three different methods, all $\Delta[\text{HbO}_2]$ and $\Delta[\text{HHb}]$ raw data were transformed
162 into z -scores (using means and standard deviations). Separate 3 (workload: 30% vs 50% vs

163 90%) \times 3 (method: SM vs AM vs AUC) MANOVAs with repeated measures were
164 performed on the z-scores of $\Delta[\text{HbO}_2]$ and $\Delta[\text{HHb}]$ data. For significant results, post-hoc
165 mean comparisons were performed using Bonferroni corrections for multiple comparisons.
166 The level of significance was set at $p < 0.05$ and partial estimated effect sizes (η^2_p) were
167 reported for significant results.

168 **Results**

169 Figure 1 shows an illustration of the typical hemodynamic changes observed in one
170 participant; $\Delta[\text{HHb}]$ increased and $\Delta[\text{HbO}_2]$ decreased as a function of workload. Globally,
171 the standardized Cronbach alphas were 0.87 for $\Delta[\text{HHb}]$ and 0.95 for $\Delta[\text{HbO}_2]$, indicating
172 a strong consistency between the three methods. The bivariate correlation coefficients
173 between the three methods at each workload were all significant (between .66 and .98)
174 with the exception of the AUC method and the SM at 30% of workload for both $\Delta[\text{HbO}_2]$
175 and $\Delta[\text{HHb}]$ (all $r = .5$, ns) and the AUC method and the AM for $\Delta[\text{HHb}]$ at 30% of
176 workload ($r = .57$, ns).

177 *Insert figure1 about here*

178 The SM

179 The analyses showed a main effect of workload on $\Delta[\text{HbO}_2]$ and on $\Delta[\text{HHb}]$: $F(2,18) =$
180 15.8 ; $p < 0.01$; $\eta^2_p = 0.63$ and $F(2,18) = 20.12$; $p < 0.01$; $\eta^2_p = 0.69$, respectively. Post-hoc
181 analyses showed that there was a significant difference between each workload for both
182 $\Delta[\text{HbO}_2]$ and $\Delta[\text{HHb}]$ slope coefficients (see Figure 2).

183 *Insert Figure 2 about here*

184 The AM

185 The analyses showed a main effect of workload on $\Delta[\text{HbO}_2]$ and on $\Delta[\text{HHb}]$: $F(2,18) =$
186 14.51 ; $p < 0.01$; $\eta^2_p = 0.61$ and $F(2,18) = 30.04$; $p < 0.01$; $\eta^2_p = 0.76$, respectively. For
187 $\Delta[\text{HbO}_2]$ data, post-hoc analyses showed that there was a significant difference between

188 30% and 90% and between 50% and 90%, but not between 30% and 50% ($p = 0.1$; see
189 Figure 3). For $\Delta[\text{HHb}]$ data, post-hoc analyses showed that there was a significant
190 difference between each workload.

191 *Insert Figure 3 about here*

192 The AUC

193 The analyses showed a main effect of workload on $\Delta[\text{HbO}_2]$ and on $\Delta[\text{HHb}]$: $F(2,18) =$
194 10.73 ; $p < 0.01$; $\eta^2_p = 0.54$ and $F(2,18) = 32.48$; $p < 0.01$; $\eta^2_p = 0.78$, respectively. For
195 $\Delta[\text{HbO}_2]$ data, post-hoc analyses showed that there was only a significant difference
196 between 30% and 90% (see Figure 4). For $\Delta[\text{HHb}]$ data, post-hoc analyses showed that
197 there was a significant difference between each workload.

198 *Insert Figure 4 about here*

199 Comparison of the three methods

200 The MANOVA performed on $\Delta[\text{HbO}_2]$ z-scores showed a significant effect of workload
201 and a main effect of the method: Wilk's lambda = 0.22; $F(2,9) = 15.85$; $p = 0.001$ and
202 Wilk's Lambda = 0.32; $F(2,9) = 9.53$; $p = 0.005$, respectively. A significant workload \times
203 method interaction was also revealed: Wilk's Lambda = 0.12; $F(4,7) = 11.73$; $p = 0.003$.
204 Post-hoc analyses showed that only the SM was sufficiently sensitive to indicate a
205 significant difference between 30% and 50% workloads ($p < 0.01$).

206 For $\Delta[\text{HHb}]$ z-scores, a significant main effect of workload was revealed: Wilk's lambda =
207 0.17 ; $F(2,9) = 21.43$; $p = 0.0003$. The main effect of the method was not significant ($p =$
208 0.22).

209 Discussion

210 In the present study, NIRS was used to examine relative changes in muscle oxygenation
211 parameters as a function of three workloads during an isometric exercise. These muscle
212 oxygenation changes were calculated using three different methods that are often reported

213 in the literature to investigate their respective sensitivity for determining the magnitude of
214 hemodynamic parameters as a function of workload.

215 In regard to the effect of workload, the results showed a consistent decrease in $\Delta[\text{HbO}_2]$
216 and an increase in $\Delta[\text{HHb}]$ induced by the isometric contractions as a function of workload
217 (see Figure 1). This result is important because no study has specifically examined the
218 sensitivity of the hemodynamic parameters as a function of workload.

219 This evolution of the hemodynamic parameters during muscular contraction can be
220 explained by an important and local consumption of oxygen. For example, it was shown
221 that at 10% of maximum voluntary contraction, the oxygen consumption of the superficial
222 flexor and brachioradial is five times higher than the quantity of oxygen needed at rest
223 (Van Beekvelt et al., 2001). Recent studies also confirmed the increase in oxygen
224 consumption during a voluntary isometric contraction (Ryan et al., 2012; Ryan et al.,
225 2013). These changes in oxygenation concentration would represent oxygen needed by the
226 muscle mitochondria during the contraction. Van Beekvelt et al. (2002) showed that local
227 muscle oxygen consumption at rest, as well as during exercise, can be reliably measured by
228 NIRS. Our results confirm this reliability of the continuous-wave NIRS technique.

229 The comparison of the sensitivity to workload between the three different methods used in
230 the present study showed that only the SM provided a significant decrease of $\Delta[\text{HbO}_2]$
231 concentration between 30% and 50% of MVS. The AM and the AUC method only
232 detected significant variations between 30% and 90% and between 50% and 90% of MVS.
233 However, the three methods appeared to be equivalent in detecting variations in $[\text{HHb}]$ as a
234 function of the three manipulated workloads. During an isometric contraction with forces
235 varying between 12, 18 and 24% of the maximum voluntary isometric contraction, Sako et
236 al. (2001) calculated the changes in $[\text{HbO}_2]$ using the least-square method and showed
237 important and significant decreases. The diminution in $[\text{HbO}_2]$ was strongly correlated ($r =$

238 0.97) with the decrease in phosphocreatine (Pcr), measured with P-MRS (Sako et al.,
239 2001). This result shows that a slight increase of workload contraction can induce an
240 important variation of hemodynamic parameters. Using the SM, the variation of workload
241 contraction between 30% and 50% induces important changes in hemodynamic
242 parameters. Therefore, it is important to use a method that is sufficiently sensitive to detect
243 slight changes, particularly in the case of a small sample of participants. In our study, only
244 the SM method was sufficiently sensitive to detect the changes between 30 and 50% of
245 MVS. Globally, using the SM, Δ [HHb] increased by 23%, 20% and 43% between 30%,
246 50% and 90% of MVS, respectively, and Δ [HbO₂] decreased by 22%, 21% and 42%
247 between 30%, 50% and 90% of MVS, respectively.

248 In the present study, the SM was shown to be the most sensitive method for detecting
249 significant muscular [HbO₂] decreases at low levels of workload (between 30 and 50% of
250 MVS) and for a small sample of participants. The sensitivity of this method could be
251 explained by the characteristics of the [HbO₂] signal, which increases linearly during the
252 entire activation period (see Figure 1) and thus appears particularly appropriate for this
253 type of method. However, the basis of the AM, which computes the mean difference
254 between rest and contraction periods during 10 second windows, is that only a portion of
255 the entire signal, thought to be representative of the complete hemodynamics, is captured.
256 This method seems particularly appropriate when the signal reaches and maintains a
257 plateau, which was clearly not the case for the hemodynamic data in the present study. As
258 such, this could indicate that this method is not well adapted for detecting significant
259 [HbO₂] decreases as a function of slight changes in low levels of MVS. The AUC method,
260 similar to the SM, uses the entire activation period for computing hemodynamic changes
261 but was shown in our study to be the less sensitive method for detecting [HbO₂] changes.
262 This result deserves future studies to more specifically examine this lack of sensitivity.

263 Taken together, the results of this study favor the use of the SM, which is more sensitive
264 and well adapted to this type of signal because NIRS signals have a linear evolution in
265 most investigations of muscle oxygenation (Ferrari et al., 1997). Moreover, the SM is easy
266 to use because it requires only a linear regression on the NIRS signals.

267 Some potential limitations of the present study should be addressed. First, quantifying
268 intramuscular oxygenation changes with NIRS requires the occlusion technique to control
269 the circulation (Van Beekvelt et al., 2001). This technique estimates the microcirculation
270 status of muscle (Gerovasili et al., 2010). In our study, we did not use this method.
271 However, this method is used to precisely calculate $m\dot{V}O_2$ (oxygen consumption of
272 muscle). In our study, this index was not the index of interest because the parameters that
273 we have examined were $\Delta[HbO_2]$ and $\Delta[HHb]$. Second, we did not counterbalance the
274 three workloads during the experiment. Although this could be a clear limitation to
275 ascertain that muscle oxygenation changes were actually due to workload increases and not
276 due to time on task or fatigue, this non-counterbalancing had no effect on the estimation of
277 the sensitivity of the three methods, which was the principal aim of the study. Finally,
278 although sufficiently powered to detect significant differences, our study's sample size was
279 quite small and a replication of these results using a larger population with a direct
280 measure of fat thickness would strengthen the conclusion.

281 In conclusion, the important difference between the three methods (SM, AM, AUC) used
282 in this investigation is their sensitivity in the quantification of variations in muscle $[HbO_2]$
283 according to workload. The SM appears to be a well-adapted, user-friendly method to
284 determine slight changes in hemodynamic parameters.

285

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289

290 **Conflict of Interest**

291 The authors have no conflict of interest.

292 **References**

293 BHAMBHANI YN: Muscle oxygenation trends during dynamic exercise measured by
294 near-infrared spectroscopy. *Can J Appl Physiol* **29**: 504-523, 2004.

295 BINZONI T, COLIER W, HILTBRAND E, HOOFD L, CERRETELLI P: Muscle O(2)
296 consumption by NIRS: a theoretical model. *J Appl Physiol* **87**: 683-688, 1999.

297 CELIE B, BOONE J, VAN COSTER R, BOURGOIS J: Reliability of near infrared
298 spectroscopy (NIRS) for measuring forearm oxygenation during incremental handgrip
299 exercise. *Eur J Appl Physiol* **112**: 2364-2374, 2012.

300 FELICI F, QUARESIMA V, FATTORINI L, SBRICCOLI P, FILLIGOI GC, FERRARI
301 M: Biceps brachii myoelectric and oxygenation changes during static and sinusoidal
302 isometric exercises. *J Electromyogr Kinesiol* **19**:e1- e11, 2009.

303 FERRARI M, BINZONI T, QUARESIMA V: Oxidative metabolism in muscle. *Philos*
304 *Trans R Soc Lond B Biol Sci* **352**: 677-683, 1997.

305 FERRARI M, MUTHALIB M, QUARESIMA V: The use of near-infrared spectroscopy in
306 understanding skeletal muscle physiology: recent developments. *Philos Trans A Math Phys*
307 *Eng Sci* **369**: 4577-4590, 2011.

308 GEROVASILI V, DIMOPOULOS S, TZANIS G, ANASTASIOU-NANA M, Nanas S:
309 Utilizing the vascular occlusion technique with NIRS technology. *Int J Ind Ergon* **40**: 218-
310 222, 2010.

311 HAMAOKA T, McCULLY KK, NIWAYAMA M, CHANCE B: The use of muscle near-
312 infrared spectroscopy in sport, health and medical sciences: recent developments. *Philos*
313 *Trans A Math Phys Eng Sci* **369**: 4591-4604, 2011.

314 LACROIX S, GAYDA M, GREMEAUX V, JUNEAU M, TARDIF JC, NIGAM A:
315 Reproducibility of near-infrared spectroscopy parameters measured during brachial artery
316 occlusion and reactive hyperemia in healthy men. *J Biomed Opt* **17**: 077010, 2012.

317 MANFREDINI F, MALAGONI AM, FELISATTI M, MANDINI S, MASCOLI F,
318 MANFREDINI R, BASAGLIA N, ZAMBONI P: A dynamic objective evaluation of
319 peripheral arterial disease by near-infrared spectroscopy. *Eur J Vasc Endovasc Surg* **38**:
320 441-448, 2009.

321 MCCULLY KK, HAMAOKA T: Near-infrared spectroscopy: what can it tell us about
322 oxygen saturation in skeletal muscle? *Exerc Sport Sci Rev* **28**: 123-127, 2000.

323 MUTHALIB M, JUBEAU M, MILLET GY, MAFFIULETTI NA, FERRARI M,
324 NOSAKA K: Biceps brachii muscle oxygenation in electrical muscle stimulation. *Clin*
325 *Physiol Funct Imaging* **30**: 360-368, 2010.

326 QUARESIMA V, COLIER WN, VAN DER SLUIJS M, FERRARI M: Nonuniform
327 quadriceps O₂ consumption revealed by near infrared multipoint measurements. *Biochem*
328 *Biophys Res Commun* **285**: 1034-1039, 2001.

329 RYAN TE, BRIZENDINE JT, MCCULLY KK: A comparison of exercise type and
330 intensity on the noninvasive assessment of skeletal muscle mitochondrial function using
331 near-infrared spectroscopy. *J Appl Physiol* **114**: 230-237, 2013.

332 RYAN TE, ERICKSON ML, BRIZENDINE JT, YOUNG HJ, MCCULLY KK:
333 Noninvasive evaluation of skeletal muscle mitochondrial capacity with near-infrared
334 spectroscopy: correcting for blood volume changes. *J Appl Physiol* **113**: 175-183, 2012.

335 SAKO T, HAMAOKA T, HIGUCHI H, KUROSAWA Y, KATSUMURA T: Validity of
336 NIR spectroscopy for quantitatively measuring muscle oxidative metabolic rate in
337 exercise. *J Appl Physiol* **90**: 338-344, 2001.

338 VAN BEEKVELT MC, COLIER WN, WEVERS RA, VAN ENGELEN BG: Performance
339 of near-infrared spectroscopy in measuring local O₂ consumption and blood flow in
340 skeletal muscle. *J Appl Physiol* **90**: 511-519, 2001.

341 VAN BEEKVELT MC, VAN ENGELEN BG, WEVERS RA, COLIER WN: In vivo
342 quantitative near-infrared spectroscopy in skeletal muscle during incremental isometric
343 handgrip exercise. *Clin Physiol Funct Imaging* **22**: 210-217, 2002.

344 VILLRINGER A, CHANCE B: Non-invasive optical spectroscopy and imaging of human
345 brain function. *Trends Neurosci* **20**: 435-442, 1997.

346 WOLF M, FERRARI M, Quaresima V: Progress of near-infrared spectroscopy and
347 topography for brain and muscle clinical applications. *J Biomed Opt* **12**: 062104, 2007.

348

Figure captions

349 Figure1: Illustration of the typical variations of hemodynamic parameters as a function of
350 workload for one participant.

351 Figure 2: Mean results using the SM as a function of workload. Bars represent standard
352 deviation. * $p < .05$ for HHb and † $p < .05$ for HbO₂.

353 Figure 3: Mean results using the AM as a function of workload. Bars represent standard
354 deviation. * $p < .05$ for HHb and † $p < .05$ for HbO₂.

355 Figure 4: Mean results using the AUC as a function of workload. Bars represent standard
356 deviation. * $p < .05$ for HHb and † $p < .05$ for HbO₂.

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358 Figure 1

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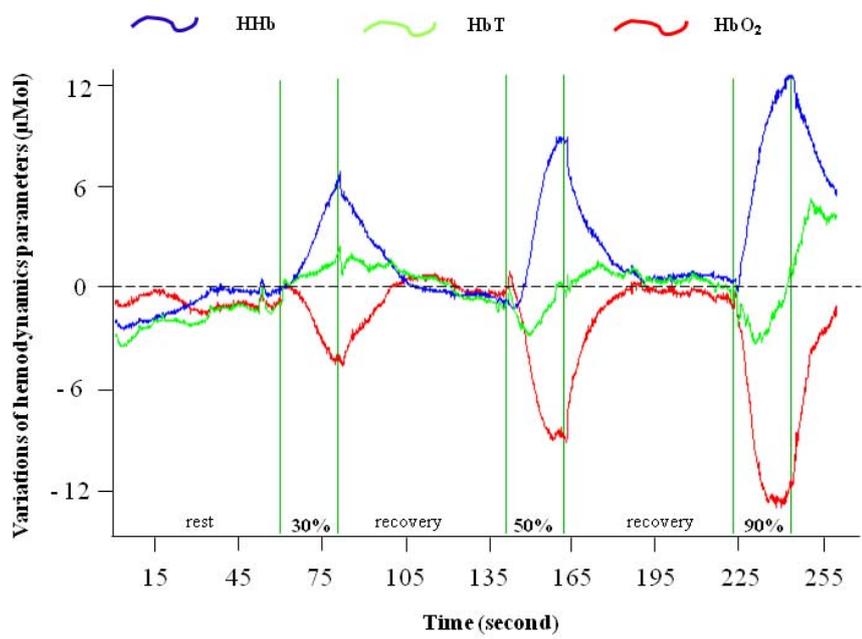
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370 Figure 2

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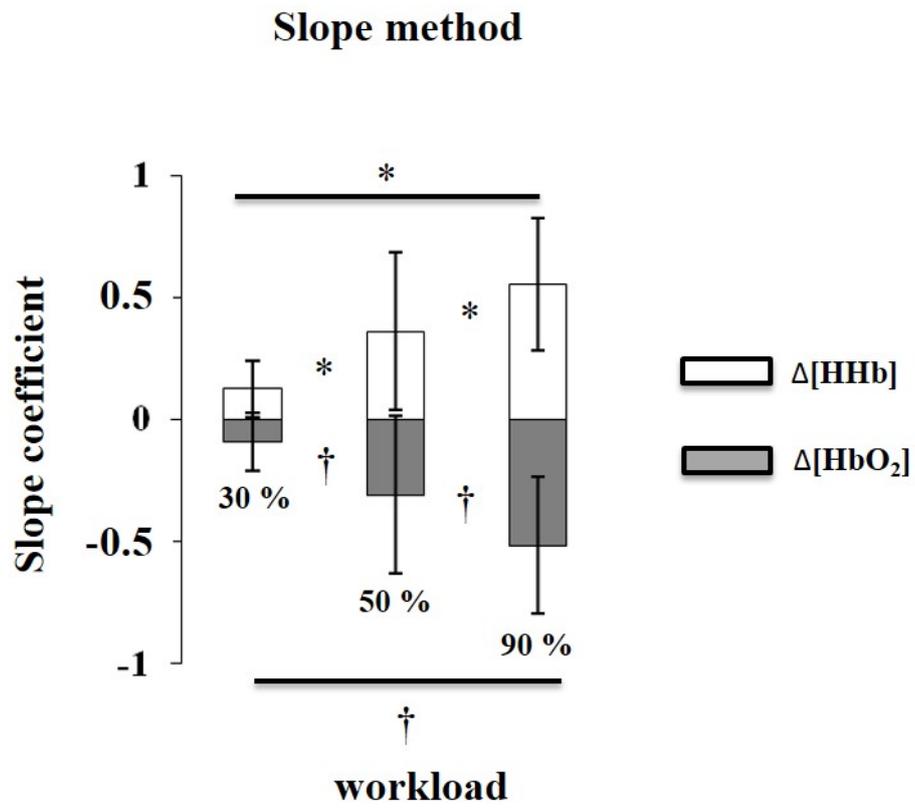
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384 Figure 3

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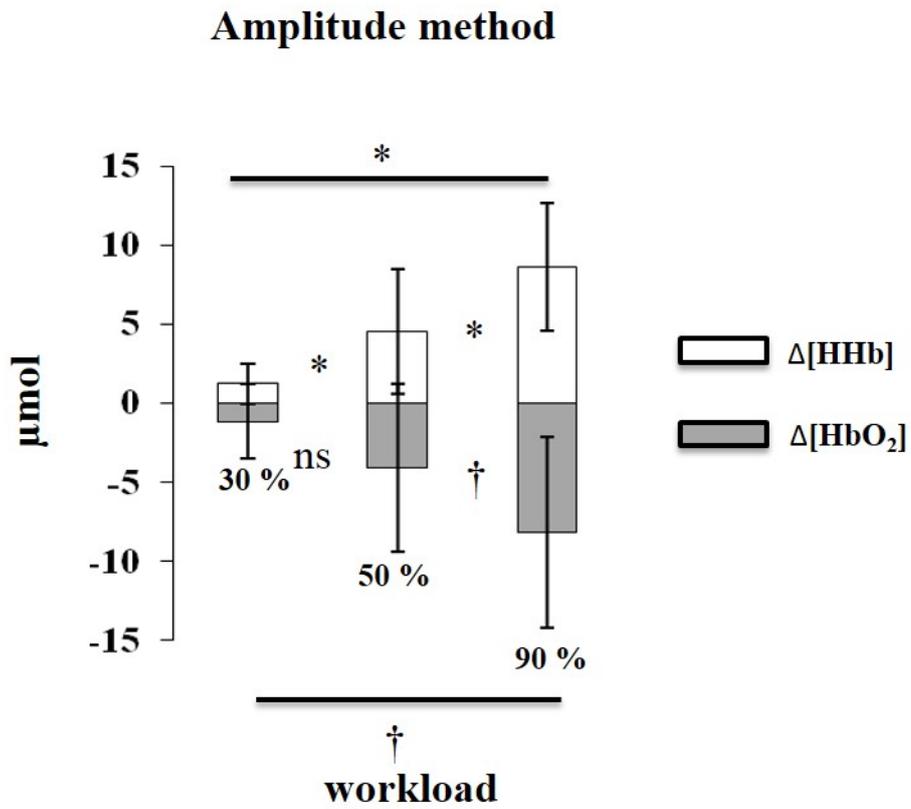
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397 Figure 4

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