

Plasma Ghrelin Levels in Patients with End-Stage Renal Disease

Z. JARKOVSKÁ, M. ROSICKÁ, M. KRŠEK, S. SULKOVÁ¹, M. HALUZÍK, V. JUSTOVÁ, Z. LACINOVÁ, J. MAREK

Third Department of Medicine and ¹Department of Medicine at Strahov, First Faculty of Medicine, Charles University, Prague, Czech Republic

Received October 10, 2003

Accepted September 15, 2004

On-line available December 9, 2004

Summary

Ghrelin is an acylated peptide stimulating secretion of the growth hormone (GH). It was originally isolated from the rat stomach as an endogenous ligand for the growth hormone secretagogue receptor. Although being predominantly produced by endocrine cells of the gastric fundus, its secretion has been found in various tissues including the kidney. To study the influence of renal failure on plasma ghrelin levels we examined 16 patients with end-stage renal disease (ESRD) receiving hemodialysis (8 men and 8 women) and 19 controls (10 men and 9 women). Both groups were comparable in age and BMI. In all subjects we assessed plasma levels of ghrelin, leptin, soluble leptin receptor, insulin, IGF-I, IGFBP-1, IGFBP-3 and IGFBP-6. Ghrelin levels were significantly higher in the group of dialyzed patients (4.49 ± 0.74 vs. 1.79 ± 0.15 ng/ml; $p < 0.001$). These patients had significantly higher levels of GH, IGFBP-1, IGFBP-6, leptin and percentage of body fat ($p < 0.05$). In the group of patients with ESRD plasma ghrelin levels positively correlated with IGFBP-1 ($p < 0.01$). In the control group, ghrelin positively correlated with GH concentrations ($p < 0.01$) and negatively correlated with the levels of insulin and creatinine ($p < 0.05$). In conclusion, patients with ESRD have higher ghrelin concentrations, which might be caused by a decreased excretion/metabolism of ghrelin in the kidney during renal failure.

Key words

Ghrelin • Leptin • Soluble leptin receptor • End-stage renal disease

Introduction

Ghrelin, an endogenous ligand for the growth hormone secretagogue receptor (GHS-R), is a novel peptide hormone predominantly produced by the stomach (Kojima *et al.* 1999, Date *et al.* 2000, Rosická *et al.* 2002). It is a 28-amino acid peptide with a unique structure, where serine at position 3 is octanoylated (Bednarek *et al.* 2000). Des-acyl ghrelin, which lacks a

hydrophobic chain substitution, is another form of ghrelin devoid of its endocrine functions. It predominates in the systemic circulation (Hosoda *et al.* 2000).

Ghrelin displays a strong growth hormone (GH) releasing activity, which is even stronger than that of the growth hormone-releasing hormone (GHRH) (Kojima *et al.* 1999, Seoane *et al.* 2000). Via NPY/AGRP neurons in the hypothalamus ghrelin stimulates food intake and causes body weight gain due to a significant increase in

fat tissue (Wren *et al.* 2000, Nakazato *et al.* 2001). Plasma ghrelin levels are decreased in obesity, elevated in cachexia and show a diurnal rhythm (Tschöp *et al.* 2001, Shiiya *et al.* 2002, Rosická *et al.* 2003).

Leptin is a protein hormone produced predominantly by adipocytes, which suppresses appetite and decreases food intake by inhibition of the NPY/AGRP system in hypothalamus (Ahima *et al.* 1996). Serum leptin levels are markedly elevated in patients with end-stage renal disease (ESRD), which suggests a possible role for leptin in the development of anorexia and protein-energy malnutrition in these patients (Haluzík *et al.* 2000).

Meanwhile, ghrelin production has been identified in many tissues and organs including the hypothalamus, pituitary, pancreas, bowel, thyroid or kidney (Date *et al.* 2000, Mori *et al.* 2000, Kršek *et al.* 2002). However, not much is known about ghrelin clearance or metabolism, and what factors other than nutrition affect the circulating ghrelin concentrations. To address this issue we determined plasma ghrelin concentrations in patients with ESRD and their relations to other studied parameters, such as leptin, the soluble leptin receptor, GH, IGF-I, IGF-I binding proteins and insulin.

Methods

We examined 16 patients with ESRD receiving hemodialysis (8 men and 8 women) and 19 control healthy subjects (10 men and 9 women). Both groups were comparable in age (patients 66.25 ± 2.46 years, controls 64.16 ± 2.18 years) and BMI (patients 25.14 ± 1.0 kg/m², controls 25.21 ± 1.03 kg/m²). In all subjects we assessed levels of ghrelin, leptin, soluble leptin receptor, insulin, GH, IGF-I, IGFBP-1, IGFBP-3 and IGFBP-6. The present study was conducted under written informed consent and approved by the Ethical Committee of the First Faculty of Medicine of the Charles University.

In the control group blood samples were collected at 08:00 h after an overnight fast. In patients with ESRD receiving hemodialysis, blood was collected similarly at 08:00 h before hemodialysis after an overnight fasting. All subjects were weighed and measured on the same day. The hemodialysis procedures were conducted with low flux polysulphone dialyser (1.5 m²) and bicarbonate dialysis solution, Kt/V index (OCM module) was above 1.2 in all patients.

The human plasma ghrelin levels were

determined using a commercial RIA kit measuring total plasma ghrelin, the sum of octanoylated ghrelin and des-acyl ghrelin (Linco Research, USA).

The human serum leptin levels and soluble leptin receptor levels were detected using the commercial ELISA kits (Bio Vendor, Czech Republic). GH and IGF-I serum levels were determined using the commercial IRMA kits (Immunotech, Czech Republic), IGFBP-1, IGFBP-3 and IGFBP-6 were measured by commercial IRMA kits (DSL, USA). Serum insulin levels were determined using commercial RIA kits (CIS Bio International, France). Body fat was determined using Best's calliper.

Using SigmaStat statistical analysis software (Jandel Scientific, San Rafael, CA, USA) the statistical analysis of the differences between both groups was performed by Student's t-test for unpaired data distribution or by the Mann-Whitney non-parametric test. Interdependence between variables within the separate groups were evaluated using Pearson's or Spearman's correlation. All data are presented as means \pm S.E.M. $P < 0.05$ values were considered significant.

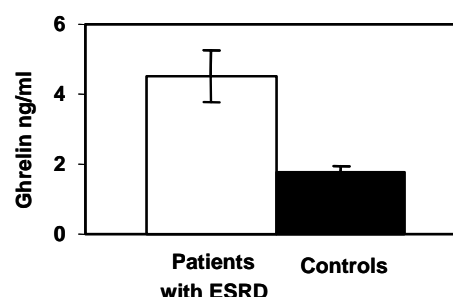


Fig. 1. Serum ghrelin levels in patients with end-stage renal disease (ESRD) compared with healthy subjects.

Results

Both studied groups were comparable in age and BMI, but significantly differed in serum creatinine level (patients 726 ± 51 μ mol/l, controls 94 ± 3 μ mol/l, $p < 0.0001$). Ghrelin plasma levels were significantly higher in the group of patients with ESRD (4.49 ± 0.74 vs. 1.79 ± 0.15 ng/ml, $p < 0.001$, Fig. 1). The serum levels of GH, IGFBP-1, IGFBP-6, leptin and percentage of body fat were also significantly higher in this group ($p < 0.05$). Comparison of laboratory parameters of the two groups is given in Table 1.

The relationships between laboratory variables in the group of patients with ESRD are given in Table 2.

In the group of patients with ESRD plasma ghrelin levels positively correlated with IGFBP-1 ($r = 0.53$, $p < 0.05$), in the control group plasma ghrelin positively correlated with GH ($r = 0.59$, $p < 0.02$) and leptin serum levels

($r = 0.48$, $p < 0.05$) and negatively correlated with serum levels of insulin ($r = -0.51$, $p < 0.05$) and creatinine ($r = -0.57$, $p < 0.02$).

Table 1. Comparison between laboratory parameters of patients with end-stage renal disease (ESRD) and healthy controls.

	Patients with ESRD	Controls
<i>Ghrelin (ng/ml)</i>	4.49±0.74	1.79±0.15**
<i>Leptin (ng/ml)</i>	26.53±4.64	15.58±2.77*
<i>Leptin receptor (ng/ml)</i>	25.52±3.69	21.88±2.36
<i>Insulin (μIU/ml)</i>	23.64±3.14	21.11±2.60
<i>Growth hormone (mIU/l)</i>	13.54±3.57	5.05±2.32*
<i>IGF-I (μg/l)</i>	139.94±15.21	155.33±16.93
<i>IGFBP-1 (μg/l)</i>	175.76±36.23	37.72±4.41*
<i>IGFBP-3 (mg/l)</i>	4.27±0.36	3.81±0.17
<i>IGFBP-6 (mg/l)</i>	2.26±0.56	0.39±0.03*
<i>Creatinine (μmol/l)</i>	725.94±51.16	93.79±3.24***

IGF-I – insulin-like growth factor-I; IGFBP-1 – insulin-like growth factor binding protein-1; IGFBP-3 – insulin-like growth factor binding protein-3; IGFBP-6 – insulin-like growth factor binding protein-6; * $p < 0.05$, ** $p < 0.001$, *** $p < 0.0001$

Table 2. Interdependence between Laboratory Variables in the Group of Patients with ESRD

	BMI	Ghrelin	GH	IGF-I	IGFBP-1	Leptin	Leptin-R	Creatinin
<i>BMI</i>		$r = -0.447$ $p = 0.083$	$r = 0.198$ $p = 0.463$	$r = 0.087$ $p = 0.749$	$r = -0.206$ $p = 0.443$	$r = 0.519$ $p = 0.039$	$r = -0.364$ $p = 0.165$	$r = -0.411$ $p = 0.114$
<i>Ghrelin</i>	$r = -0.447$ $p = 0.083$		$r = 0.222$ $p = 0.408$	$r = 0.209$ $p = 0.436$	$r = 0.525$ $p = 0.037$	$r = -0.192$ $p = 0.476$	$r = 0.318$ $p = 0.230$	$r = 0.399$ $p = 0.126$
<i>GH</i>	$r = 0.198$ $p = 0.463$	$r = 0.222$ $p = 0.403$		$r = 0.104$ $p = 0.701$	$r = 0.586$ $p = 0.017$	$r = 0.186$ $p = 0.491$	$r = 0.264$ $p = 0.324$	$r = -0.287$ $p = 0.282$
<i>IGF-I</i>	$r = 0.087$ $p = 0.749$	$r = 0.209$ $p = 0.436$	$r = 0.104$ $p = 0.701$		$r = 0.028$ $p = 0.919$	$r = 0.114$ $p = 0.674$	$r = 0.147$ $p = 0.588$	$r = 0.218$ $p = 0.418$
<i>IGFBP-1</i>	$r = -0.206$ $p = 0.443$	$r = 0.525$ $p = 0.037$	$r = 0.586$ $p = 0.017$	$r = 0.028$ $p = 0.919$		$r = -0.069$ $p = 0.799$	$r = 0.485$ $p = 0.57$	$r = -0.005$ $p = 0.987$
<i>Leptin</i>	$r = 0.519$ $p = 0.039$	$r = -0.192$ $p = 0.476$	$r = 0.186$ $p = 0.491$	$r = 0.114$ $p = 0.674$	$r = -0.069$ $p = 0.799$		$r = -0.455$ $p = 0.077$	$r = 0.050$ $p = 0.853$
<i>Leptin-R</i>	$r = -0.366$ $p = 0.165$	$r = 0.318$ $p = 0.230$	$r = 0.264$ $p = 0.324$	$r = 0.147$ $p = 0.588$	$r = 0.485$ $p = 0.057$	$r = -0.455$ $p = 0.077$		$r = -0.019$ $p = 0.946$
<i>Creatinine</i>	$r = -0.411$ $p = 0.114$	$r = 0.399$ $p = 0.126$	$r = -0.287$ $p = 0.282$	$r = 0.218$ $p = 0.418$	$r = -0.005$ $p = 0.987$	$r = 0.050$ $p = 0.853$	$r = -0.019$ $p = 0.946$	

BMI – body mass index; GH – growth hormone; IGF-I – insulin-like growth factor-I; IGFBP-1 – insulin-like growth factor binding protein-1; Leptin-R – soluble leptin receptor; r – correlation coefficient; p – p value

Discussion

Ghrelin is a novel peptide hormone which was

originally isolated as an endogenous ligand of the growth hormone secretagogue receptor (Kojima *et al.* 1999). Stomach is the main source of circulating ghrelin

(Toshinai *et al.* 2001), but not much is known about its clearance and metabolism.

Previous observations reported a negative correlation between ghrelin and BMI, body fat mass and plasma leptin levels (Tschöp *et al.* 2001). Low plasma ghrelin levels were found in obese individuals (English *et al.* 2002), whereas malnutrition was associated with high ghrelin levels (Otto *et al.* 2001).

Our group of patients with ESRD receiving hemodialysis was characterized by a significantly higher percentage of body fat. When compared with a group of controls with a comparable BMI, a suppression of plasma ghrelin levels could be expected, but we found higher ghrelin levels in this group. This could be caused by a decreased renal excretion.

A positive correlation between total plasma ghrelin and creatinine in renal failure has been previously reported. No correlation was found between acylated ghrelin and creatinine (Yoshimoto *et al.* 2002). In our study we measured total plasma ghrelin, which did not correlate with creatinine. The reason for this is not clear, we plan further investigations in patients with mild-to-severe renal failure.

Ghrelin has an orexigenic effect in humans, where its administration leads to hunger sensations (Cummings *et al.* 2002). Anorexia is often present in patients with renal failure so that low ghrelin levels could be expected in this condition. Our results, however, show significantly increased ghrelin levels. Higher ghrelin levels associated with anorexia were already found in malnutrition (Ariyasu *et al.* 2001) and anorexia nervosa (Becker *et al.* 1999). One possible explanation might be a resistance of hypothalamic centers to ghrelin in the presence of its high concentrations. Ghrelin is not the only compound affecting appetite; markedly elevated serum leptin levels in renal failure, which we also confirmed, at least contribute to anorexia. Because both leptin and ghrelin affect antagonistically appetite *via* the regulation of NPY/AGRP system in hypothalamus (Nakazato *et al.* 2001), a local interplay between these

two compounds could play a role.

Ghrelin is a strong stimulator of GH release (Kojima *et al.* 1999). High GH concentrations in our group of patients with ESRD, which is consistent with previous findings, could thus be at least in part caused by elevated ghrelin levels. In our study we found no correlation between plasma ghrelin levels and GH concentrations, similarly to previous observations (Cappiello *et al.* 2002).

The elevated levels of IGFBP-1 and normal levels of IGF-I in ESRD have been described in numerous studies (Divino *et al.* 1998, Nanba *et al.* 2001). The IGFBP-1 level is thought to be primarily determined by insulin level in portal blood (Suikkari *et al.* 1989). In our study we assessed serum insulin levels, which did not differ between both studied groups. This was also observed by Divino *et al.* (1998). Furthermore, IGFBP-1 concentration as well as its production in liver is markedly increased in malnutrition, a condition which is commonly seen in patients with ESRD (Sanaka 2003).

Patients with chronic renal failure had normal levels of IGF-I. Although there is a reduction of IGF-I gene expression and GH insensitivity in uremia (GH levels are high, but there is downregulation of hepatic GH receptors), the IGF-I level remains in the normal range. This phenomenon could be explained by increased concentrations of IGFBPs in uremia (Tonshoff *et al.* 1997).

Why are plasma ghrelin levels elevated in renal failure? One possibility is a reduction of ghrelin degradation/clearance in end-stage renal disease. However an overproduction of ghrelin in certain tissues might contribute to its higher plasma concentrations. In order to answer this question we plan further investigations.

Acknowledgements

This experiment was supported by the grant of IGA MHCR No. NB/7099-4.

References

- AHIMA RS, PRABAKARAN D, MANTZOROS C: Role of leptin in the neuroendocrinology of fasting. *Nature* **382**: 250-252, 1996.
- ARIYASU H, TAKAYA K, TAGAMI T, OGAWA Y, HOSODA K, AKAMIZU T, SUDA M, KOH T, NATSUI K, TOYOOKA S, SHIRAKAMI G, USUI T, SHIMATSU A, DOI K, HOSODA H, KOJIMA M, KANGAWA K, NAKAO K: Stomach is a major source of circulating ghrelin, and feeding state determine plasma ghrelin-like immunoreactivity levels in humans. *J Clin Endocrinol Metab* **86**: 4753-4758, 2001.

- BECKER AE, GRINSPOON SK, KLIBANSKI A, HERZOG DB: Eating disorders. *N Engl J Med* **340**: 1092-1098, 1999.
- BEDNAREK MA, FEIGHNER SD, PONG SS, MCKEE KK, HRENIUK DL, SILVA MV, WARREN VA, HOWARD AD, VAN DER PLOEG LH, HECK JV: Structure-function studies on the new growth hormone-releasing peptide, ghrelin: minimal sequence of ghrelin necessary for activation of growth hormone secretagogue receptor 1a. *J Med Chem* **43**: 4370-4376, 2000.
- CAPPIELLO V, RONCHI C, MORPURGO PS, EPAMINONDA P, AROSIO M, BECK-PECCOZ P, SPADA A: Circulating ghrelin levels in basal conditions and during glucose tolerance test in acromegalic patients. *Eur J Endocrinol* **147**: 189-194, 2002.
- CUMMINGS DE, WEIGLE DS, FRAYO RS, BREEN PA, MA MK, DELLINGER EP, PURNELL JQ: Plasma ghrelin levels after diet-induced weight loss or gastric bypass surgery. *N Engl J Med* **346**: 1623-1630, 2002.
- DATE Y, KOJIMA M, HOSODA H, SAWAGUCHI A, MONDAL MS, SUGANUMA T, MATSUKURA S, KANGAWA K, NAKAZATO M: Ghrelin, a novel growth hormone-releasing acylated peptide, is synthesized in a distinct endocrine cell type in the gastrointestinal tract of rats and humans. *Endocrinology* **141**: 4255-4261, 2000.
- DIVINO FILHO JC, HAZEL SJ, FURST P, BERGSTROM J, HALL K: Glutamate concentration in plasma, erythrocyte and muscle in relation to plasma levels of insulin-like growth factor (IGF)-I, IGF binding protein-1 and insulin in patients on haemodialysis. *J Endocrinol* **156**: 519-527, 1998.
- ENGLISH PJ, GHATEI MA, MALIK IA, BLOOM SR, WILDING JP: Food fails to suppress ghrelin levels in obese humans. *J Clin Endocrinol Metab* **87**: 2984-2987, 2002.
- HALUZÍK M, SULKOVÁ S, SVOBODOVÁ J, BEDÁROVÁ V, BODLÁKOVÁ B, MARKOVÁ M, TURKOVÁ G, JISKRA J, HAAS T: Serum leptin levels in diabetic patients on hemodialysis: the relationship to parameters of diabetes metabolic control. *Endocr Res* **26**: 303-317, 2000.
- HOSODA H, KOJIMA M, MATSUO H, KANGAWA K: Ghrelin and des-acyl ghrelin: two major forms of rat ghrelin peptide in gastrointestinal tissue. *Biochem Biophys Res Commun* **279**: 909-913, 2000.
- KOJIMA M, HOSODA H, DATE Y, NAKAZATO M, MATSUO H, KANGAWA K: Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature* **402**: 656-660, 1999.
- KRŠEK M, ROSICKÁ M, HALUZÍK M, SVOBODOVÁ J, KOTRLÍKOVÁ E, JUSTOVÁ V, LACINOVÁ Z, JARKOVSKÁ Z: Plasma ghrelin levels in patients with short bowel syndrome. *Endocr Res* **28**: 27-33, 2002.
- MORI K, YOSHIMOTO A, TAKAYA K, HOSODA K, ARIYASU H, YAHATA K, MUKOYAMA M, SUGAWARA A, HOSODA H, KOJIMA M, KANGAWA K, NAKAO K: Kidney produces a novel acylated peptide, ghrelin. *FEBS Lett* **486**: 213-216, 2000.
- NAKAZATO M, MURAKAMI N, DATE Y, KOJIMA M, MATSUO H, KANGAWA K, MATSUKURA S: A role for ghrelin in the central regulation of feeding. *Nature* **409**: 194-198, 2001.
- NANBA K, NAGAKE Y, MIYATAKE N, NAKAO K, AKAGI S, SUGIMOTO T, YAMASAKI H, OISCHI K, ICHIKAWA H, MAKINO H: Relationships of serum levels of insulinlike growth factors with indices of bone metabolism and nutritional conditions in hemodialysis patients. *Nephron* **89**: 145-152, 2001.
- OTTO B, CUNTZ U, FRUEHAUF E, WAWARTA R, FOLWACZNY C, RIEPL RL, HEIMAN ML, LEHNERT P, FICHTER M, TSCHÖP M: Weight gain decreases elevated plasma ghrelin concentrations of patients with anorexia nervosa. *Eur J Endocrinol* **145**, 669-673, 2001.
- ROSICKÁ M, KRŠEK M, JARKOVSKÁ Z, MAREK J, SCHREIBER V: Ghrelin – a new endogenous growth hormone secretagogue. *Physiol Res* **51**: 435-441, 2002.
- ROSICKÁ M, KRŠEK M, MATOULEK M, JARKOVSKÁ Z, MAREK J, JUSTOVÁ V, LACINOVÁ Z: Serum ghrelin levels in obese patients: the relationship to serum leptin levels and soluble leptin receptor levels. *Physiol Res* **52**: 61-66, 2003.
- SANAKA T: Nutritional effect of dialysis therapy. *Artif Organs* **27**: 224-226, 2003.
- SEOANE LM, TOVAR S, BALDELLI R, ARVAT E, GHIGO E, DIEGUEZ C: Ghrelin elicits a marked stimulatory effect on GH secretion in freely-moving rats. *Eur J Endocrinol* **143**: R7-R9, 2000.

- SHIYA T, NAKAZATO M, MIZUTA M, DATE Y, MONDAL MS, TANAKA M, NOZOE S, HOSODA H, KANGAWA K, MATSUKURA S: Plasma ghrelin levels in lean and obese humans and the effect of glucose on ghrelin secretion. *J Clin Endocrinol Metab* **87**: 240-244, 2002.
- SUIKKARI AM, KOIVISTO VA, KOISTINEN R, SEPPALA M, YKI-JARVINEN H: Dose-response characteristics for suppression of low molecular weight plasma insulin-like growth factor binding protein by insulin. *J Clin Endocrinol Metab* **68**: 135-140, 1989.
- TONSHOFF B, POWELL DR, ZHAO D, DURHAM SK, COLEMAN ME, DOMENE HM, BLUM WF, BAXTER RC, MOORE LC, KASKEL FJ: Decreased hepatic insulin-like growth factor (IGF)-I and increased IGF binding protein-1 and -2 gene expression in experimental uremia. *Endocrinology* **138**: 938-946, 1997.
- TOSHINAI I, MONDAL MS, NAKAZATO M, DATE Y, MURAKAMI N, KOJIMA M, KANGAWA K: Upregulation of ghrelin expression in the stomach upon fasting, insulin-induced hypoglycemia, and leptin administration. *Biochem Biophys Res Commun* **281**: 1220-1225, 2001.
- TSCHÖP M, WEYER C, TATARANNI PA, DEVANARAYAN V, RAVUSSIN E, HEIMAN ML: Circulating ghrelin levels are decreased in human obesity. *Diabetes* **50**: 707-709, 2001.
- WREN AM, SMALL CJ, WARD HL, MURPHY KG, DAKIN CL, TAHERI S, KENNEDY AR, ROBERTS GH, MORGAN DG, GHATEI MA, BLOOM SR: The novel hypothalamic peptide ghrelin stimulates food intake and growth hormone secretion. *Endocrinology* **141**: 4325-4328, 2000.
- YOSHIMOTO A, MORI K, SUGAWARA A, MUKOYAMA M, KENSEI Y, SUGANAMI T, TAKAYA K, HOSODA H, KOJIMA M, KANGAWA K, NAKAO K: Plasma ghrelin and desacyl ghrelin concentrations in renal failure. *J Am Soc Nephrol* **13**: 2748-2752, 2002.

Reprint requests

Z. Jarkovská, Third Department of Medicine, First Faculty of Medicine, Charles University, U nemocnice 1, 128 08 Praha 2, Czech Republic. E-mail: zuzana.jarkovska@email.cz