Reply to Artifacts in Electron Microscopic Research

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Dear Editor,

We carefully read the comments on our paper "Pineal Gland Calcification under Hypoxic Conditions" (Physiol Res 68 (Suppl 4): S405-S413, 2019) by Varga *et al.* (this issue); our response is below.

Calcification of the pineal gland has been known for a long time. Our data are in line with the literature and we demonstrated calcification by 3 different techniques: light microscopy, transmission electron microscopy (TEM) and energy-dispersive analysis (EDX) rather than by TEM only, which Varga *et al.* commented upon. The results of all three methods are consistent. The new findings of our study are related to the effects of prenatal hypoxic conditions on the process of pineal gland calcification. We found that prenatal hypoxia can accelerate the process of calcification, although the biological consequences are still unknown.

The application of contrast agents (uranyl acetate and lead citrate) in electron microscopy can cause the presence of artefacts (electron-dense particles) in the investigated samples. To avoid the misinterpretation of observed particles, EDX analysis was performed. We found particles consisting of Ca, P, S, and Na. EDX analysis revealed that the amount of Ca varied from 0.21 to 20.26 weight%, and the amount of U from 0.64 to 8.55 weight%. Some particles did not reveal uranium or lead

at all. Moreover, some authors observed that the cytoplasm of pinealocytes contains vacuoles filled with flocculent and fibrous material (Humbert *et al.* 1997, Swietoslavski 1999). From these results, we can conclude that the analyzed electron-dense particles are the result of calcium accumulation and not artefacts.

Biomineralisation (the formation of minerals in the tissue) is a complicated process; the results depend on many factors such as time, the presence of chemical elements and biomolecules, the pH of the surrounding microenvironment, the presence of water, etc. In this case, the structure (concentric lamination) can be strongly influenced by conditions during the hypoxia. The absence of concentric lamination can reflect these conditions.

Of course, the presence of vacuoles can be the result of compression. However, as we mentioned in our article, many authors observed the presence of vacuoles in the pineal gland under physiological conditions (Reiter *et al.* 1976, Humbert and Pevet, 1994, 1995, Schmid and Raykhtsaum 1995) and under hypoxia in other tissues (Zeng *et al.* 2019, Sun *et al.* 2019, Zhuravin *et al.* 2003). In addition, the preparation of all samples was the same for all groups and the control group showed almost no vacuoles. Therefore, we conclude that vacuolisation is not the result of compression during preparation but a reaction to the conditions.

We focused on calcium particles and described the fibrous materials inside the vacuoles in the top right corner of Fig. 4. We agree that the fibrous material outside of the vacuoles in the top right corner could be collagen fibres. Tomonari *et al.* (2012) investigated 383 rats from a histological viewpoint and found focal fibrosis on the periphery of the gland; sometimes it was observed in the gland. Therefore, we are convinced that Fig. 4 represents calcification in the parenchyma of the pineal gland.

The whole process of pineal gland calcification is not definitely understood and there are several hypotheses which explain underlying mechanisms. The authors Varga *et al.* mentioned only one hypothesis suggested by Vigh-Teichmann and Vigh (1992) many years ago. Since then, several other hypotheses have been formulated and are reviewed in a recent paper (Tan *et al.* 2018). The paper was either not read or pointedly ignored by Varga *et al.* Our explanation for the increased calcification is based on one of the hypotheses suggested by Tan *et al.* (2018). We did not deal with all possible hypotheses because this was not the aim of our study.

We found this new hypothesis challenging and our data may support it. Hypoxia is generally accompanied by the generation of reactive oxygen species causing oxidative stress, which promotes the production of different cytokines (Jomova et al. 2010, Pathipati et al. 2013). Cytokines are known to promote "reprogramming" of mesenchymal cells, the as demonstrated many times with different models (Mijiritsky et al. 2020, Krampeta 2011, Zimmermann et al. 2017). In this context, we discussed a possible mechanism for the calcification of the pineal gland in our study. On the basis of above mentioned arguments we are convinced that conclusions of our paper are valid and cannot be questioned.

References

- HUMBERT W, PEVET P: Calcium concretions in the pineal gland of aged rats: an ultrastructural and microanalytical study of their biogenesis. Cell Tissue Res 279: 565-573, 1994.
- HUMBERT W, PEVET P: The pineal gland of the aging rat: Calcium localization and variation in the number pinealocytes. J Pineal Res 18: 32-40, 1995.
- HUMBERT W, CUISINIER F, VOEGEL JC, PEVET P: A possible role of collagen fibrils in the process of calcification observed in the capsula of the pineal gland in aging rats. Cell Tissue Res 288: 435-439, 1997.
- JOMOVA K, VONDRACKOVA D, LAWSON M, VALKO M: Metals, oxidative stress and neurodegenerative disorders. Mol Cell Biochem 345: 91-104, 2010.
- KOPANI M, VRANIKOVA B, KOSNAC D, ZEMAN M, SISOVSKY V, POLAKOVICOVA S, BIRO C: Pineal gland calcification under hypoxic conditions. Physiol Res 68 (Suppl 4): S405-S413, 2019.
- KRAMPERA M: Mesenchymal stromal cell'licensing': a multistep process. Leukemia 25:1408-1414, 2011.
- MIJIRITSKY E, GARDIN C, FERRONI L, LACZA Z, ZAVAN B: Albumin-impregnated bone granules modulate the interactions between mesenchymal stem cells and monocytes under in vitro inflammatory conditions. Mater Sci Eng Mater Biol Appl 110: 110678, 2020.
- PATHIPATI P, MULLER S, JIANG XN, FERRIERO D: Phenotype and secretory responses to oxidative stress in microglia. Develop Neurosci 35: 241-254, 2013.
- REITER RJ, WELSH MG, VAUGHAN MK: Age-related changes in the intact and sympathetically denervated gebril pineal gland. Am J Anat 146: 427-431, 1976.
- SCHMID HA, RAYKHTSAUM G: Age related differences in the structure of human pineal calcium deposits: results of transmission electron microscopy and mineralographic microanalysis. J Pineal Res 18: 12-20, 1995.
- SUN SM, WU Y, FU HT, YANG M, GE XP, ZHU J, XUAN FJ, WU XG: Evaluating expression of autophagy-related genes in oriental river prawn Macrobrachium nipponense as potential biomarkers for hypoxia exposure. Ecotoxic Environ Safety 171: 484-492, 2019.
- SWIETOSLAWSKI J: The age-related quantitative ultrastructural changes in pinealocytes of gerbils. Neuro Endocrinol Lett 20: 391-396, 1999.
- TAN DX, XU B, ZHOU X, REITER RJ: Pineal calcification, melatonin production, aging, associated health consequences and rejuvenation of the pineal gland. Molecules 23: 1-31, 2018.
- TOMONARI Y, SATO J, WAKO Y, TSUCHITANI M: Age-related histological findings in the pineal gland of Crl:CD(SD) rats. J Toxicol Pathol 25: 287-291, 2012.
- VARGA I, GHALLAB A, DANISOVIC L: Artifacts in electron microscopic research. Physiol Res 69: 537-539, 2020.

- VIGH-TEICHMANN I, VIGH B: Immunocytochemistry and calcium cytochemistry of the mammalian pineal organ: a comparison with retina and submammalian pineal organs. Microsc Res Tech 21: 227-241, 1992.
- ZENG L, AI CX, ZHENG JL, ZHANG JS, LI WC: Cu pre-exposure alters antioxidant defense and energy metabolism in large yellow croaker Larimichthys crocea in response to severe hypoxia. Sci Total Environ 687: 702-711, 2019.
- ZHURAVIN IA, TUMANOVA NL, DUBROVSKAYA NM, FEDOSEEVA KN: Disturbances in formation of the new and old cortex at changes of conditions of embryonic development. J Evolut Biochem Physiol 39: 752-763, 2003.
- ZIMMERMANN JA, HETTIARATCHI MH, MCDEVITT TC: Enhanced immunosuppression of T cells by sustained presentation of bioactiveinterferon-gamma within three-dimensional mesenchymal stem cellconstructs. Stem Cells Transl Med. 6: 223-237, 2017.