

## REVIEW

# A Review of the Effects of Herbal Medicines on the Conductance of Ion Channels

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## Summary

Experimental evidence indicates that several phytochemicals can modulate ion channels currents, probably improving disorders linked to ion channels in cardiovascular, neurological, and gastrointestinal systems. This review critically evaluates the experimental evidence regarding the effects of bioactive phytochemicals from medicinal plants on ion channels properties. This review primarily focused on the beneficial effects of plant-derived bioactive agents on ion channels implicated in cardiovascular, neurological, and gastrointestinal disorders, specifically cardiac arrhythmias, epilepsy, anxiety, pain, and visceral smooth muscle dysfunction. Relevant literatures up to 2024 were gathered through comprehensive searches across multiple electronic databases. The results indicate that near 50 medicinal plants and their derivatives can modulate the conductivity of various Ca<sup>2+</sup>, K<sup>+</sup>, Na<sup>+</sup>, and Cl<sup>-</sup> ion channels, alter the electrical properties of excitable cells, and affect the functional features of tissues and organ systems. These observations provide a foundational framework for researchers, health professionals and drug developers seeking to understand how botanical compounds impacts the functionality of ion channels.

## Keywords

Plant derivatives • Ion channels • Therapeutic properties

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## 1. Introduction

Ion channels are specialized transmembrane proteins for the passage of ions across biological membranes. These structures are typically depicted as narrow, aqueous pores that allow ions to pass through according to their size and electrical charge. This characteristic denotes the ability of the pore to discriminate between ions based on their physical and electrochemical properties [1]. The movement of calcium, potassium, and sodium ions is essential for various physiological functions such as electrical activity and contraction of striated, smooth, and heart muscles, insulin secretion, T-lymphocyte activation, and generation and propagation of neural signals. These ions can passively diffuse across the membrane through specific ion channels driven by their electrochemical gradients and membrane potential [2].

Ion channels, classified according to their ion selectivity and the stimuli that trigger their opening or

closing, include voltage-gated sodium channels ( $\text{Na}_v$ ), voltage-gated calcium channels ( $\text{Ca}_v$ ), various voltage-gated potassium channels ( $\text{K}_v$ ) such as inward-rectifier potassium channel ( $\text{K}_{ir}$ ), certain chloride channels, and ligand-gated channels such as ATP-sensitive potassium channels ( $\text{K}_{ATP}$ ), acetylcholine-activated potassium channels ( $\text{K}_{ACh}$ ), small-conductance calcium-activated potassium channels (SK), transient receptor potential (TRP) channels, hyperpolarization-activated cyclic nucleotide-gated (HCN) channels, among others. Some receptors, such as ryanodine receptors (RyR) and inositol 1,4,5-trisphosphate ( $\text{IP}_3$ ) receptors, which are located in the membrane of the endoplasmic reticulum, mediate the

release of calcium from intracellular stores into the cytoplasm [3-6]. Table 1 summarizes various types of ions channels and their localization, distribution, common inhibitors, and key function [7-9].

Each ion channel plays a specific role in regulating the membrane potential and synergistically contributes to the generation of action potential in excitable cells. Dysfunction of these channels, resulting from genetic mutations or post-translational modifications, can disturb cell function, leading to various diseases, such as myocardial hypertrophy, arrhythmias, heart failure, epilepsy, ileus, and other channelopathies [10].

**Table 1.** Classification, synonyms, localization, and major characteristics of ion channels

Type of Channel	Synonyms/Abbreviation	Main Ion Conducted	Localization	Key Characteristics / Function	Clinically Relevant Inhibitors / Modulators
<b>Voltage-gated sodium channels</b>	$\text{Na}_v$ (e.g., $\text{Na}_v1.5$ in the heart)	$\text{Na}^+$	Plasmalemmal	Initiate and propagate action potentials (rapid depolarization)	Lidocaine, tetrodotoxin
<b>L-type voltage-gated calcium channels</b>	$\text{Ca}_v1.x$ ( $\text{Ca}_v1.1$ – $\text{Ca}_v1.4$ ); LTCC (L = long, long-lasting, large)	$\text{Ca}^{2+}$	Plasmalemmal (T-tubules, sarcolemma)	Long-lasting $\text{Ca}^{2+}$ influx; excitation-contraction coupling	Nifedipine, verapamil, diltiazem
<b>T-type voltage-gated calcium channels</b>	$\text{Ca}_v3.x$	$\text{Ca}^{2+}$	Plasmalemmal	Low-voltage-activated; pacemaker activity	Mibefradil, ethosuximide
<b>P/Q-type voltage-gated calcium channels</b>	$\text{Ca}_v2.1$	$\text{Ca}^{2+}$	Presynaptic terminals	Neurotransmitter release	$\omega$ -Agatoxin IVA
<b>N-type voltage-gated calcium channels</b>	$\text{Ca}_v2.2$	$\text{Ca}^{2+}$	Neurons	Neurotransmitter release	$\omega$ -Conotoxin GVIA
<b>R-type voltage-gated calcium channels</b>	$\text{Ca}_v2.3$	$\text{Ca}^{2+}$	Neurons, endocrine cells	Synaptic transmission, hormone release	SNX-482
<b>Voltage-gated potassium channels</b>	$\text{K}_v$	$\text{K}^+$	Plasmalemmal	Repolarization, control of excitability	4-aminopyridine tetraethylammonium (TEA)
<b>Inwardly rectifying potassium channels</b>	$\text{K}_{ir}$	$\text{K}^+$	Plasmalemmal	Maintain resting potential	Barium ions ( $\text{Ba}^{2+}$ )
<b>ATP-sensitive potassium channels</b>	$\text{K}_{ATP}$	$\text{K}^+$	Plasmalemmal (heart, pancreas)	Couple metabolism to excitability	Glibenclamide, diazoxide
<b>Acetylcholine-activated potassium channels</b>	$\text{K}_{ACh}$	$\text{K}^+$	Plasmalemmal	Parasympathetic regulation of heart rate	Muscarinic antagonists






<b>Small-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels</b>	SK	K <sup>+</sup>	Plasmalemmal	Ca <sup>2+</sup> -dependent hyperpolarization	Apamin
<b>Big-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels</b>	BK (Maxi-K, KCa1.1)	K <sup>+</sup>	Plasmalemmal, smooth muscle	Link Ca <sup>2+</sup> to membrane potential; regulate vascular tone	Paxilline, iberiotoxin
<b>Two-pore domain K<sup>+</sup> channels</b>	K2P (TREK, TASK)	K <sup>+</sup>	Plasmalemmal	Background (leak) K <sup>+</sup> current; set resting potential	Volatile anesthetics, bupivacaine
<b>Chloride channels</b>	ClC, CFTR, CaCC	Cl <sup>-</sup>	Plasmalemmal	Volume regulation, secretion, excitability	Niflumic acid, CFTR modulators
<b>Ca<sup>2+</sup>-activated chloride channels</b>	TMEM16A (ANO1)	Cl <sup>-</sup>	Plasmalemmal	Smooth muscle contraction, epithelial secretion	Niflumic acid
<b>Ligand-gated chloride channels</b>	GABA <sub>A</sub> , glycine receptors	Cl <sup>-</sup>	Plasmalemmal (neurons)	Inhibitory neurotransmission	benzodiazepines, picrotoxin
<b>Proton channels</b>	HVCN1	H <sup>+</sup>	Plasmalemmal	Regulate pH, NADPH oxidase activity	Zinc ions (Zn <sup>2+</sup> )
<b>Transient receptor potential channels</b>	TRP (TRPC, TRPV, TRPM, etc.)	Ca <sup>2+</sup> , Na <sup>+</sup>	Plasmalemmal	Sensory transduction, temperature, osmotic stress	2-APB, ruthenium red
<b>Mechanosensitive / stretch-activated channels</b>	Piezo1, Piezo2, TREK-1	Cations (Na <sup>+</sup> , Ca <sup>2+</sup> , K <sup>+</sup> )	Plasmalemmal	Mechano-transduction, pressure sensing	GsMTx4 (spider toxin)
<b>Hyperpolarization-activated cyclic nucleotide-gated channels</b>	HCN	Na <sup>+</sup> , K <sup>+</sup>	Plasmalemmal (SA node, neurons)	Pacemaker (If) currents	Ivabradine
<b>Pannexin and connexin channels</b>	Pannexin, Connexin (gap junctions)	Ions, ATP	Plasmalemmal	Intercellular communication, paracrine signaling	Carbenoxolone
<b>Ryanodine receptors</b>	RyR1–RyR3	Ca <sup>2+</sup>	Intracellular (sarcoplasmic/endoplasmic reticulum)	Release of Ca <sup>2+</sup> from SR/ER stores	Ryanodine, caffeine
<b>Inositol 1,4,5-trisphosphate receptors</b>	IP <sub>3</sub> R	Ca <sup>2+</sup>	Intracellular (endoplasmic reticulum)	IP <sub>3</sub> -mediated Ca <sup>2+</sup> release	Xestospongine C, 2-APB
<b>Store-operated calcium channels</b>	Orai1/STIM1 complex (SOCE)	Ca <sup>2+</sup>	Plasmalemmal-ER junction	Refill intracellular Ca <sup>2+</sup> stores after depletion	SKF-96365, YM-58483
<b>Mitochondrial calcium uniporter</b>	MCU	Ca <sup>2+</sup>	Intracellular (mitochondrial membrane)	Controls mitochondrial Ca <sup>2+</sup> uptake	Ru360

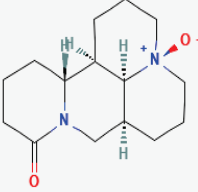
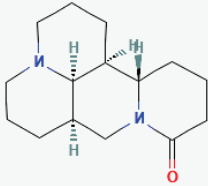
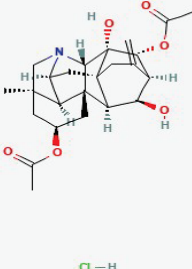
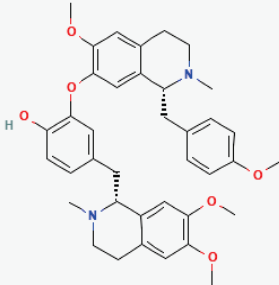
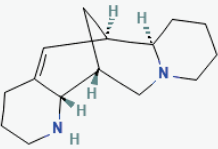
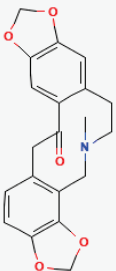
Many synthetic drugs can treat conditions like convulsion and cardiac arrhythmia by modulating the function of ion channels [11]. However, the same mechanism of action can also cause serious side effects such as dangerous arrhythmias and instability in the nervous system, thereby increasing the risk of death.

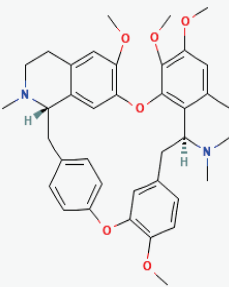
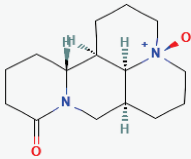
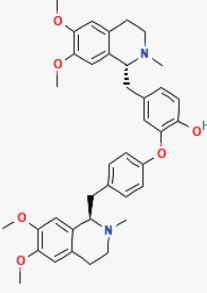
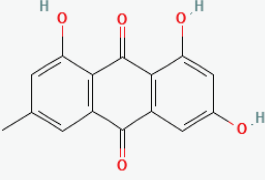
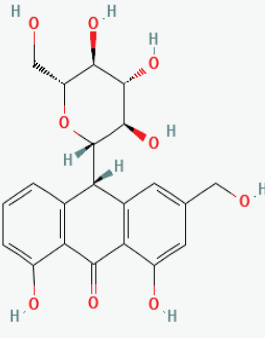
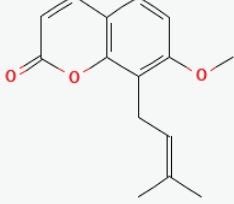
For thousands of years, plants have been recognized as a rich reservoir of biologically active substances for therapeutic use [12]. Their widespread use in traditional medicine is attributed to their affordability and accessibility [13]. The basic mechanisms of action of

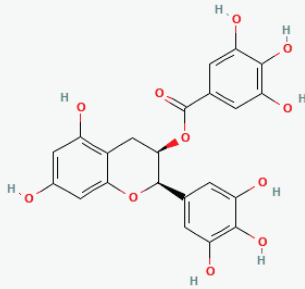
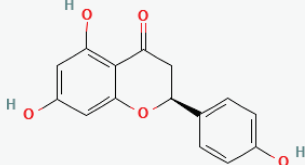
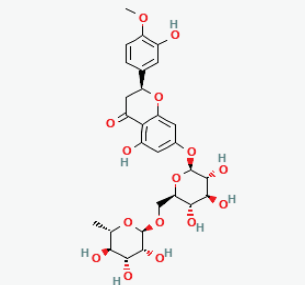
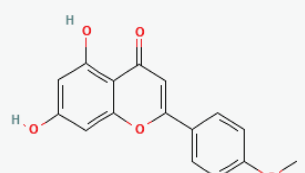
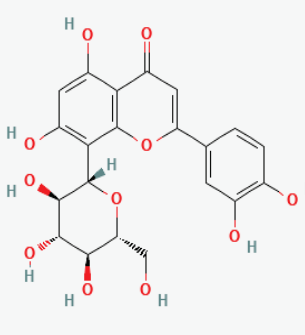
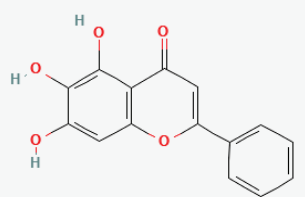
some herbal medicines are increasingly being covered, revealing that many plants and their constituents exert therapeutic and/or side effects through the modulation of specific ion channels. Given the limited number of reviews on this subject, this narrative review aimed to consolidate current knowledge by focusing on the effects of herbal medicines on various ion channels across different body systems. We also described the modulatory mechanisms of specific herbal compounds, with supplementary data summarized in Tables 2-4 as well as in Figures 1-6.

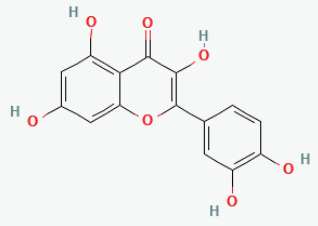
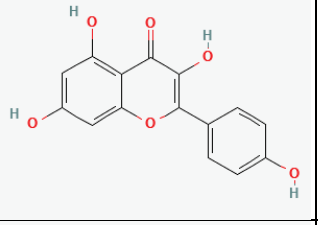
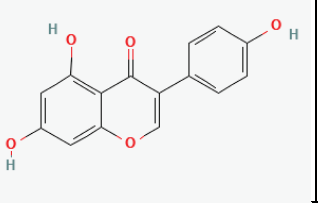
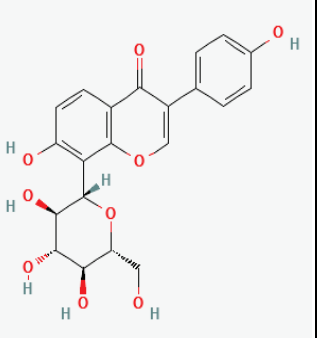
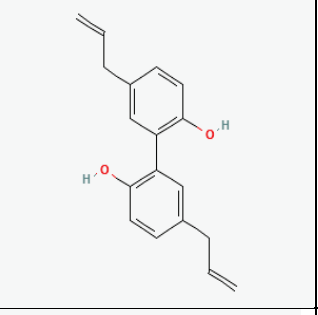
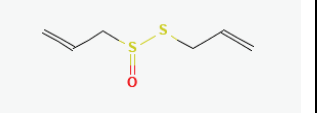
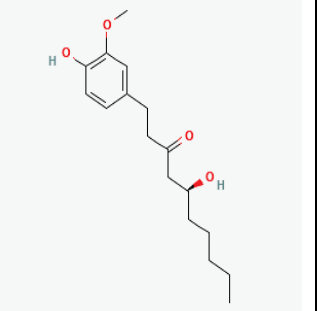
**Table 2.** The properties of compounds and plants.

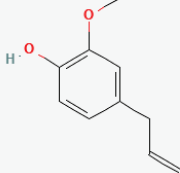
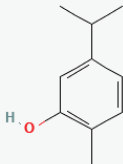
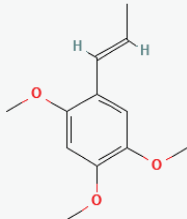
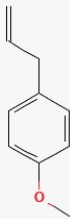
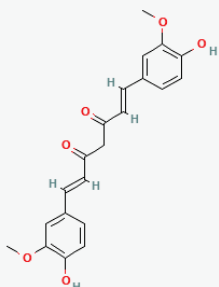
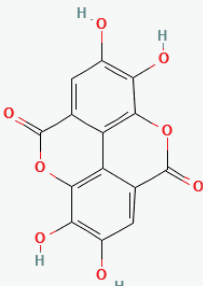
Name	Type of compound	Compound chemical structure or plant picture	Source of compound	Chemical constituents of plants
<b>Salvia miltiorrhiza</b>	-		-	Salvianolic acid Dihydrotanshinone Miltirone Tanshinone I
<b>Lavender</b>	-		-	Linalyl acetate Linalool Tannins Caryophyllene
<b>Melissa officinalis (Lemon Balm)</b>	-		-	Eugenol Tannins Terpenes
<b>Aralia elata</b>	-		-	SilphiosideA Chikusetosaponin Ib Araloside A
<b>Saffron</b>	-		-	Picrocrocin Safranal Carotenoid Crocin

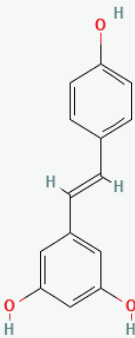
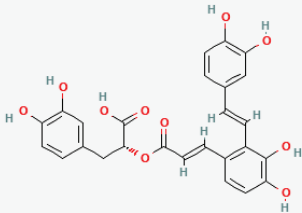
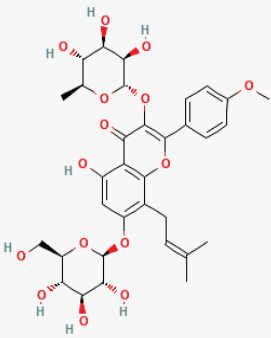
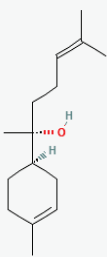
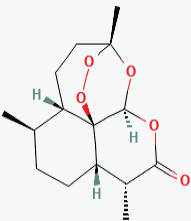
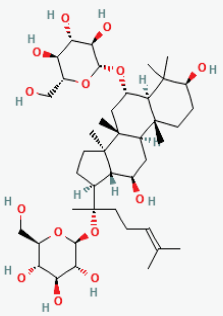
<b>Berberine</b>	Alkaloid		<i>Berberis vulgaris</i>  <i>Berberis aristata</i>  <i>Mahonia aquifolium</i>  <i>Hydrastis canadensis</i>	-
<b>Martine</b>	Alkaloid		<i>Sophora flavescens</i>	-
<b>Guanfu base A</b>	Alkaloid		<i>Aconitum coreanum</i>	-
<b>Neferine</b>	Alkaloid		<i>Nelumbo nucifera</i>	-
<b>Aloperine</b>	Alkaloid		<i>Sophora alopecuroides</i>	-
<b>Protopine</b>	Alkaloid		<i>Opium poppy</i>  <i>Corydalis tubers</i>	-

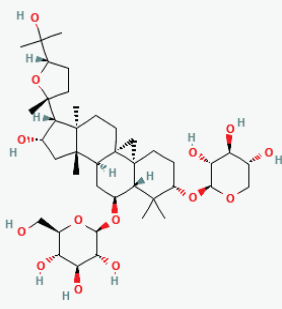
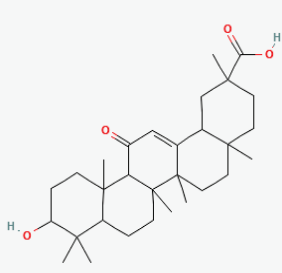
<b>Tetrandrine</b>	Alkaloid		<i>Stephania tetrandra</i>	-
<b>Oxymatrine</b>	Alkaloid		<i>Sophora flavescens</i>	-
<b>Dauricine</b>	Alkaloid		<i>Menispermum dauricum</i> <i>Menispermum canadense</i>	-
<b>Emodin</b>	Anthraquinone glycone		<i>Rhubarb</i> <i>Buckthorn</i>	
<b>Barbaloin</b>	Anthraquinone glycosyl		<i>Aloe species</i>	-
<b>Osthol</b>	Coumarin		<i>Cnidium monnieri</i> <i>Angelica archangelica</i> <i>Angelica pubescens</i>	

<b>Epigallocatechin-3-Gallate</b>	Flavanol		<i>Apple skin</i> <i>Plums</i> <i>Onions</i> <i>Hazelnuts</i> <i>Pecans</i> <i>Tea</i>	
<b>Naringenin</b>	Flavanone		<i>Citrus fruits such as Grapefruit</i>	-
<b>Hesperidin</b>	Flavanone glycoside		<i>Peel of citrus fruits</i>	-
<b>Acacetin</b>	Flavone		<i>Robinia pseudoacacia</i> <i>Turnera diffusa</i> <i>Betula pendula</i>	-
<b>Orientin</b>	Flavone		<i>Adonis vernalis</i> <i>Anadenanthera colubrina</i> <i>Anadenanthera peregrina</i> <i>Phyllostachys nigra</i>	
<b>Baicalein</b>	Flavone		<i>Scutellaria baicalensis</i>  <i>Scutellaria lateriflora</i>	-

<b>Quercetin</b>	Flavonol		Many fruits, vegetables	-
<b>Kaempferol</b>	Flavonol		<i>Kale</i> <i>Beans</i> <i>Tea</i> <i>Spinach</i> <i>Broccoli</i>	
<b>Genistein</b>	Isoflavonoid		<i>Soybeans</i>	
<b>Puerarin</b>	Isoflavonoid		<i>Pueraria lobata</i>  <i>Pueraria phaseoloides</i>	-
<b>Magnolol</b>	Lignan		<i>Magnolia officinalis</i>  <i>Magnolia grandiflora</i>	-
<b>Allicin</b>	Organosulfur compound		<i>Garlic</i> <i>Leeks</i>	-
<b>6-gingerol</b>	Phenolic phytochemical		<i>Ginger</i>	-

<b>Eugenol</b>	Phenylpropanoid		<i>Nutmeg</i> <i>Cinnamon</i> <i>basil</i> <i>bay leaf</i>	-
<b>Carvacrol</b>	Phenylpropanoid		<i>Origanum vulgare</i> <i>Thyme</i> <i>Pepperwort</i> <i>Wild bergamot</i>	-
<b>Asarone</b>	Phenylpropanoid		<i>Acorus</i> <i>Asarum</i>	-
<b>Estragole</b>	Phenylpropene		<i>Anise</i> <i>Fennel</i> <i>Bay</i> <i>Tarragon</i> <i>Basil</i>	-
<b>Curcumin (Turmeric)</b>	Polyphenol		<i>Curcuma longa</i> <i>Ginger</i>	-
<b>Ellagic acid</b>	Polyphenol		Numerous fruits and vegetables	-

<b>Resveratrol</b>	Polyphenol		<i>Grapes</i> <i>Blueberries</i> <i>Raspberries</i> <i>Mulberries</i> <i>Peanuts</i>	-
<b>Salvianic acid A</b>	Polyphenolic acid		<i>Salvia miltiorrhiza</i>	
<b>Icariin</b>	Prenylated flavonol glycoside		<i>Epimedium</i>	-
<b><math>\alpha</math>-(-)-Bisabolol</b>	Sesquiterpene alcohol		<i>Matricaria recutita</i> <i>Myoporum crassifolium</i>	-
<b>Artemisinin</b>	Sesquiterpene lactone		<i>Artemisia annua</i>	-
<b>Ginsenoside</b>	Steroid glycosides and triterpene saponins		<i>Panax (ginseng)</i>	-

Astragaloside IV	Triterpenoid saponin		<i>Astragalus hoantchy</i>  <i>Astragalus lepsensis</i>	-
Glycyrrhizic acid	Triterpenoid saponin		<i>Glycyrrhiza glabra</i>	-

## 2. Search Methods

A comprehensive and systematic search was conducted across seven major scientific databases: Scopus, Springer Link, EMBASE, Science Direct, PubMed, Google Scholar, and Web of Science. The search strategy incorporated the following key terms: "ion channel modulation", "plant-derived ion channel modulators", "phototherapeutic compounds", "bioactive plant constituents", "secondary plant metabolites", and "traditional medicinal plants". Publications up to November 2024 were considered. Studies were eligible for inclusion if they involved preclinical investigations and clinical trials. Botanical substances exhibiting cardiac rhythm stabilization, analgesic, neurological effects, particularly anticonvulsant actions, smooth muscle relaxation, or cholinergic pathway modulation were selected. The selection process followed a rigorous two-phase screening approach, beginning with title and abstract assessment, followed by full-text evaluation of potentially relevant studies. To ensure methodological consistency, the search strategy was guided by the Population, Intervention, Comparator, and Outcome (PICO) framework. To maintain conciseness without compromising completeness, when multiple studies on the same botanical agent reported comparable findings, preference was given to the most recent or the highest-quality study. To minimize the selection bias, two independent reviewers performed the screening. Any disagreements were resolved through consensus discussion, thereby enhancing the reliability and validity of the included studies.

## 3. Results

### 3.1. Plants

#### 3.1.1. *Aralia elata*

The Japanese, Chinese, or Korean angelica tree (*Aralia elata*) is a woody plant species in the family *Araliaceae* [14]. The positive inotropic effect of *Aralia elata* observed in canine myocardium and isolated rat cardiomyocytes is potentially mediated by an increase in the amplitude of the cytosolic calcium transient [22]. It has been also shown to boost the amplitude of L-type calcium channels (LTCC) current in heart of rats with diabetic cardiomyopathy [21], identifying a likely pathway for the elevated intracellular calcium.

#### 3.1.2. Lavender

*Lavandula angustifolia* is an evergreen plant native to the Mediterranean, known for its fragrant flowers and essential oil, which are also used medicinally. Lavender oil has calming effects and may help relax certain muscles. It also shows antibacterial and antifungal properties [15]. Lavender has been shown to modulate transient calcium channel (T-type  $\text{Ca}^{2+}$  channels), contributing to its analgesic and sedative effects [16]. It also inhibits N- and P/Q-types voltage-gated calcium channels in the hippocampus [17,18]. Hashimoto *et al.* [19] further revealed inhibitory effects of linalool, an essential component of lavender oil on nociceptive transient receptor potential cation channel A1 (TRPA1) and voltage-gated  $\text{Ca}^{2+}$  channels in mouse sensory neurons.

### 3.1.3. *Melissa officinalis* L.

*Melissa officinalis* is an aromatic and medicinal plant commonly known as lemon balm, honey balm, or bee balm. *Melissa officinalis* is an excellent source of terpene and polyphenolic compounds, including phenolic acids such as rosmarinus acid, caffeic acid, and protocatechuic acid; flavonoids such as quercitrin, rhamnocitrin, luteolin; and tannins [20]. Lemon balm has sedative and mild hypnotic, hypoglycemic, hepatoprotective, antibacterial, anti-inflammatory, antioxidant, antiviral, antispasmodic, anxiolytic, neuroprotective, cardioprotective, and antiarrhythmic properties [20-22].

The mechanisms underlying the anxiolytic effects of *Melissa officinalis* remain controversial. Some studies reported that *Melissa officinalis* activates the GABAergic system and increases chloride ion conductance, whereas others suggested that its effects are mediated through modulation of the cholinergic system and voltage-dependent calcium channels [23]. Gazola *et al.* [24] demonstrated that *Melissa officinalis* may reduce heart rate *via* stimulation of cardiac muscarinic receptors or inhibition of voltage-dependent calcium channels. The antiarrhythmic effects of this plant have also been attributed to the inhibition of calcium channels and the activation of voltage-dependent potassium channels [22,25].

### 3.1.4. *Panax notoginseng* saponins

The roots of *Panax notoginseng* (Burk.) are used as a source of *Panax notoginseng* saponins, which have been shown to exhibit antiarrhythmic effects [26]. *Panax notoginseng* saponins modulates potassium and calcium ion channels, reducing cardiomyocyte automaticity, slowing cardiac electrical conduction, prolonging action potential duration and effective refractory period, and preventing reentry arrhythmias [27]. Ginsenoside Rg1, as a monomer of *Panax notoginseng* saponins, has been reported to prolong sinus node recovery time, atrioventricular conduction Wenckebach cycle length, and ventricular effective refractory period, as well as to increase the ventricular fibrillation threshold, producing cardiac electrophysiological effects similar to those of amiodarone [28].

### 3.1.5. Saffron

Saffron, *Crocus sativus* L., is a widely used spice with numerous documented pharmacological properties, including neuroprotective, cardiovascular

protective, antinociceptive, antiarrhythmic, anxiolytic and bronchodilator effects [29-31]. Its major bioactive constituents are crocin, crocetin, picrocrocin, and safranal. Electrophysiological studies using the patch-clamp technique have shown that crocetin and safranal significantly inhibit the human ether-a-go-go-related gene (hERG) K<sup>+</sup> current, whereas crocin and picrocrocin do not exert such effects [32]. hERG is coded by KCNH2 gene, which is the pore-forming subunit responsible for rapid delayed-rectifier potassium current (I<sub>Kr</sub>). Crocetin has been reported to inhibit LTCC in adult rat ventricular myocytes, leading to reduced intracellular Ca<sup>2+</sup> levels and decreased contractility [33]. Safranal has antinociceptive activity through partial agonism and selective desensitization of TRPA1 [34]. It also induces vasodilatation by inhibiting calcium influx through LTCC and suppressing the Na<sup>+</sup>/Ca<sup>2+</sup> exchanger [35]. Saffron exert antiarrhythmic effect at least in part through inhibition of calcium channels [36].

### 3.1.6. *Salvia miltiorrhiza*

*Salvia miltiorrhiza* Bunge, a traditional Chinese medicines, is officially listed in the Chinese Pharmacopoeia [37]. The dried roots of *Salvia miltiorrhiza* Bunge are widely used for their therapeutic effects, including promoting blood flow, removing blood stasis, alleviating intestinal gurgling, dissolving calculi, relieving abdominal fullness, and reducing swelling [38]. The mechanism underlying *Salvia miltiorrhiza*-induced contraction remains uncertain; however, it appears not to be mediated by muscarinic receptors, sodium channels, or calcium channels, but rather through the increase in cytosolic Ca<sup>2+</sup> concentration and the activation of the Ca<sup>2+</sup>-calmodulin pathway [39]. It also has antiepileptic effects through increasing presynaptic Ca<sup>2+</sup> influx [40].

### 3.1.7. Turmeric

Turmeric, *Curcuma longa* L., a member of the *Zingiberaceae* family, is a widely used culinary spice with numerous pharmacological properties. Curcumin, its principal bioactive constituent, has neuroprotective, antioxidant, anticancer, anti-inflammatory, hepatoprotective, immunomodulatory, cardioprotective, antidepressant, antifertility, antiallergic, antimicrobial, and antidermatophytic properties [41]. Curcumin suppresses early and delayed afterdepolarization like arrhythmogenic activities, shortens action potential duration, and inhibits late sodium current (I<sub>NaL</sub>), transient sodium current (I<sub>NaT</sub>), L-type calcium current (I<sub>CaL</sub>), and I<sub>Kr</sub> [42]. Curcumin also

show the antiepileptic (anticonvulsant) effects [43], which may be mediated by modulation of neuronal sodium channels and downregulation of  $\text{Na}_v1.1$  expression in the cortex [43,44]. Evidence from animal studies suggests that its analgesic effects involve activation of both cannabinoid and opioid systems, potentially through the release of endogenous endocannabinoids and opioids [45]. Curcumin inhibits glutamate release from rat prefrontocortical synaptosomes by suppressing presynaptic voltage-gated calcium channels of N- and P/Q-type [46]. Curcumin appears to mediate antidepressant-like effects through ion channel modulation, including inhibition of  $\text{Ca}^{2+}$  release-activated  $\text{Ca}^{2+}$  (CRAC) channels [47]. It also reduces Kv currents in rabbit coronary arterial smooth muscle cells [48]. Additionally, curcumin alleviates diabetic neuropathic pain by modulating voltage-gated sodium channels and increasing sodium levels in dorsal root ganglion neurons [49]. Curcumin also exhibits gastroprotective effects *via* activation of the nitric oxide (NO) / cyclic guanosine monophosphate (cGMP) /  $\text{K}_{\text{ATP}}$  channel pathway [50]. Furthermore, it functions as a multi-ion channel blocker, with a preferential inhibitory effect on the late sodium current, suggesting potential antiarrhythmic benefits [42]. In addition, curcumin has been shown to have inhibitory, excitatory, and modulatory effects on TRP channels [51].

### 3.1.8. Salviatic acid A

Danshensu, dried root of *Salvia miltiorrhiza*, is widely used in traditional medicine for the treatment of coronary artery disease [52]. Salviatic acid A reduces the myocardial ischemia injury possibly by preventing  $\text{I}_{\text{CaL}}$  current and decreasing myocardial contractility [53].

## 3.2. Plant derivatives

### 3.2.1. 6-gingerol

6-gingerol, one of the most abundant and bioactive constituents of *Zingiber officinale* Roscoe, ginger, exhibits anticancer, anti-inflammatory, antioxidant and cardiometabolic protective effects [54]. Using patch-clamp technique and Ion Optix system, it has been demonstrated that 6-gingerol inhibits L-type calcium channels at a concentration-dependent manner in ischemic cardio-myocytes, reducing  $\text{Ca}^{2+}$  influx, stabilizing intracellular  $\text{Ca}^{2+}$  homeostasis, attenuating oxidative stress, and ultimately decreasing myocardial infarct size, and improving cardiac function [55].

*Zingiber officinale* and its active components have analgesic effect through serotonergic pathways, nitric oxide (NO) signaling,  $\text{K}_{\text{ATP}}$  channel, and modulation calcitonin gene related peptide (CGRP) [56]. 6-gingerol acts as an anticonvulsant by interacting with the amino-terminal domain, glutamate-binding site, and ion channel of the NR2B containing N-methyl-D-aspartate (NMDA) receptor, thereby restoring the balance between GABAergic and glutamatergic in the epileptic brain [57].

### 3.2.2. 8-gingerol

8-gingerol has been reported to exert antioxidant, anti-inflammatory, and cardiovascular protective effect [58]. 8-gingerol regulates the mitogen-activated protein kinase (MAPK) signaling pathway, inhibits LTCCs, decreases calcium overload, alleviates oxidative stress damage, and prevents cardiomyocyte apoptosis. These actions collectively contribute to its protective effects against myocardial ischemic injury [59].

### 3.2.3. Acacetin

The bioactive flavonoid acacetin, isolated from *Saussurea involucrata* exhibits considerable antiarrhythmic effects [60]. Pharmacological studies indicate that its mechanism involves the selective blockade of  $\text{K}^+$  channels, with negligible effects on  $\text{Na}^+$  channels or LTCC. A study in a controlled canine model also demonstrated that acacetin inhibited experimentally induced atrial fibrillation. This effect occurred without QT interval prolongation, suggesting a low risk for pro-arrhythmic side effects and a desirable safety profile [61].

### 3.2.4. Aconitine

Aconitine is a C19 norterpenoid alkaloid produced by some medicinal plants such as *Aconitum carmichaelii* Debx. Although it has potential applications to treat cancer, pain, inflammation and immune related diseases [62], its use is limited by a considerable toxicity. By binding to voltage-gated sodium channels, aconitine like mesaconitine induces the persistent activation of sodium channels, preventing its normal inactivation and becoming refractory to next excitation. The consequent uncontrolled sodium influx results in toxic effects on the nervous system, heart, and muscles. Because of this specific cardiotoxicity, aconitine is used for the arrhythmia induction in experimental studies [63,64].

### 3.2.5. Allicin

Allitridum (Allicin), the primary bioactive compound found in garlic (*Allium* genus, *Liliaceae* family), exhibits significant therapeutic potential in managing and preventing cardiovascular diseases [65]. The specific mutation, lysine-proline-glutamine deletion ( $\Delta$ KPQ), in sodium voltage-gated channel alpha subunit 5 (SCN5A) causes a well-known long QT syndrome type 3 (LQT3), an inherited cardiac arrhythmia disorder. A study showed that allicin decreases the late sodium current associated with the  $\Delta$ KPQ-SCN5A mutation, likely by enhancing steady-state and intermediate-state inactivation of the channel, thereby reducing persistent leak of sodium ions [65]. It has been also shown that in rat cerebrocortical nerve terminals, allicin depresses glutamate release and decreases N- and P/Q-type channel activity and suppresses  $Ca^{2+}$  influx and protein kinase C (PKC) activity [66]. Furthermore, allicin also attenuates transient outward potassium currents in mouse ventricular myocytes [67].

### 3.2.6. Aloperine

Aloperine, quinolizidine alkaloid isolated from *Sophora alopecuroides* L., exhibits potential therapeutic effects in cardiovascular diseases including hypertension, ventricular remodeling, myocardial ischemia, and arrhythmias [68]. Studies indicate that aloperine targets key cardiac ion channels. At a concentration of 100  $\mu$ M, it did not affect cloned  $K_v4.3$  channels but increased the  $I_{Kr}$  current [68,69]. Furthermore, in rat ventricular myocytes, aloperine significantly reduced the peak density of voltage-gated sodium current in a concentration-dependent manner [70].

### 3.2.7. Artemisinin

Artemisinin, a compound derived from the Qinghao plant, *Artemisia annua* L., has been suggested as an antiarrhythmic substance. In a study artemisinin attenuates  $I_{Na}$  by modulating the voltage dependence of the  $Na^+$  channel similar to the class I antiarrhythmic agents [71].

### 3.2.8. Asarone

Asarone isomers are phenylpropene secondary metabolites naturally occurring in species such as *Acorus calamus* L., *Guatteria gaumeri* Greenman and *Aniba hostmanniana* Nees. Chemically, they are classified into propenyl isomers (trans- $\alpha$  and cis- $\beta$  asarone) and an allylic isomer ( $\gamma$ -asarone) [72]. The antiepileptic

mechanism of these compounds is mediated through the inhibition of voltage-gated calcium and sodium channels [73].

### 3.2.9. Astragaloside IV

Astragaloside IV is a saponin purified from *Astragalus membranous* (Fisch) Bunge, which is widely used in the treatment of diabetes. It has been reported to possess various anti-inflammatory, antioxidant, antiviral and immunomodulatory effects [74]. Astragaloside IV prolongs the action potential duration in guinea pig ventricular myocytes, which can be attributed to its inhibitory effect on the  $K^+$  current. In addition, astragaloside IV suppresses  $I_{CaL}$  to affect  $Ca^{2+}$  signaling [74].

### 3.2.10. Baicalein

Baicalein is the principal active ingredient of *Scutellaria baicalensis* Georgi. Several preclinical studies reported cardioprotective effects of baicalein. However, its low oral bioavailability greatly limits clinical efficacy [75]. It has been shown baicalein can alleviate myocardial ischemic injury by inhibiting  $I_{CaL}$  and reducing intracellular calcium level [75].

### 3.2.11. Barbaloin

*Aloe vera* contains the bioactive compound barbaloin, which exhibits anti-inflammatory, antiarrhythmic, antitumor and antibacterial properties [76]. In rabbit ventricular myocytes, barbaloin has been shown to eliminate early and delayed after-depolarization [77]. Moreover, barbaloin inhibits  $I_{CaL}$  and peak sodium current in a dose-dependent manner, thereby reduces ventricular arrhythmias [78].

### 3.2.12. Berberine

Berberine is an isoquinoline alkaloid derived from the roots, stems, and bark of plants including barberry, Chinese goldthread, goldenseal, tree turmeric, and Oregon grape. Extracts from berberis have been reported to exhibit antihypertensive and antiarrhythmic effects [79-81]. Antiarrhythmic and anticonvulsant effects of those plants have also been attributed to berberine [82,83]. Berberine prolongs the action potential duration and the effective refractory period of myocardial cells, thereby inhibiting the occurrence of atrioventricular reentrant tachycardia through the suppression of potassium and calcium ion channels [82] and HCN cation channel [84,85]. In a rat model, berberine alleviate

cisplatin-induced peripheral neuropathy by modulating inflammation signal *via* TRPV1[86].

### 3.2.13. Carvacrol

Carvacrol, a natural monocyclic monoterpene (2-methyl-5-isopropyl phenol) and a major component of thyme oil, possesses several biological activities, including antioxidant properties [87,88]. It has been shown that carvacrol can reduce the action potential upstroke velocity, action potential duration and conduction velocity in heart of rabbit and human by inhibiting  $\text{Ca}^{2+}$  and  $\text{Na}^{+}$  current and blocking transient receptor potential cation channel subfamily M member 7 (TRPM7) channels [87]. TRPM7 belongs to the TRP family of ion channels, mainly conducts divalent cations such as  $\text{Mg}^{2+}$ ,  $\text{Ca}^{2+}$ , and  $\text{Zn}^{2+}$  (Table 1).

### 3.2.14. Dauricine

Dauricine, a bisbenzyltetrahydroisoquinoline alkaloid extracted from the roots of *Menispermum dauricum* DC, exhibits notable antiarrhythmic effects [89]. Dauricine prolongs the atrial effective refractory period, action potential duration, and significantly blocks  $\text{Na}^{+}$ ,  $\text{K}^{+}$ , and  $\text{Ca}^{2+}$  currents in myocardial tissues, thereby demonstrating its antiarrhythmic properties [90].

### 3.2.15. Ellagic acid

Ellagic acid, a natural polyphenolic component present in berries, has been demonstrated to exhibit antioxidant, anticancer and anti-inflammatory properties [91]. Ellagic acid can affect ionic and mechanical properties of rat ventricular myocytes. Ellagic acid through the activation of nitric oxide synthase - guanylate cyclase - cGMP pathways lead to reduction of  $\text{I}_{\text{CaL}}$  and results in negative inotropic effects. Therefore, ellagic acid may be useful in the pathophysiological conditions such as hypertension and coronary artery diseases [91].

### 3.2.16. Elatoside C

Elatoside C, a natural saponin isolated from *Aralia chinensis* L., has antioxidative activity [92]. The elatoside C has been shown to protect the heart against ischemia/reperfusion injury (I/R) by modulating intracellular  $\text{Ca}^{2+}$  homeostasis through regulation of calcium channels. Specifically, elatoside C improves abnormal calcium handling during I/R injury by enhancing the activity of sarcoplasmic reticulum  $\text{Ca}^{2+}$ -ATPase (SERCA2), reducing endoplasmic

reticulum (ER)  $\text{Ca}^{2+}$  depletion, and decreasing the expression of ER stress protein markers [93].

### 3.2.17. Emodin

Emodin, a natural anthraquinone derivative extracted from traditional medicinal herbs such as *Rheum officinale* and *Polygonum cuspidatum*, exhibits diverse pharmacological properties, including anticancer, anti-inflammatory, antioxidant, and antibacterial effects [94]. Moreover, the vasorelaxant effect of emodin is mediated by the activation of large-conductance calcium- and voltage-activated potassium (BK) channels [95].

### 3.2.18. Epigallocatechin-3-gallate

Epigallocatechin-3-gallate, a polyphenol abundant in green and white tea and present at lower concentrations in black tea, has been reported to exert protective effects against heart injury and related cardiovascular pathology [96-98]. In neonatal rat ventricular myocytes, epigallocatechin-3-gallate has been shown to activate the sodium current. At a concentration of 10  $\mu\text{M}$ , it slows down the inactivation and accelerates the recovery from inactivation [99]. Epigallocatechin-3-gallate has been found to inhibit the  $\text{I}_{\text{CaL}}$  and  $\text{I}_{\text{Kr}}$ , slow delayed-rectifier potassium current ( $\text{I}_{\text{Ks}}$ ), transient outward potassium channel (Ito), and KATP currents, leading to a decrease in the action potential duration [100,101]. It also interacts with the TRPA1 in several physiological systems. In the cardiovascular system, epigallocatechin-3-gallate activates TRPA1 on perivascular sensory nerves, promoting calcium influx and the release of CGRP, which leads to neurogenic vasodilation and reductions in blood pressure [102]. Within the nervous system, oxidized epigallocatechin-3-gallate derivatives activate TRPA1 in dorsal root ganglion neurons, driving  $\text{Ca}^{2+}$  entry and sensory-neuropeptide release, indicating a role in nociceptive signaling [103].

### 3.2.19. Eugenol

Clove and basil essential oils contain eugenol, a phenylpropene compound obtained through fractionation [104]. Eugenol has been shown to modulate the excitability of rat sciatic nerve and superior cervical ganglion neurons by inhibiting  $\text{Na}^{+}$  channels [104]. Eugenol increases intracellular  $\text{Ca}^{2+}$  levels by activating TRPA1 that may contribute to the desensitization of pain sensation and analgesic effects in trigeminal ganglion neurons [105,106].

### 3.2.20. Genistein

Genistein, an isoflavone primarily extracted from soybeans, is a well-known phytoestrogen with various biological activities, including antiproliferative, anti-inflammatory, and antioxidant effects [107]. Through a protein tyrosine kinase-dependent mechanism, genistein has been shown to inhibit BK channels, suppress cell cycle progression, and ultimately reduce low-density lipoprotein (LDL)-induced proliferation of vascular smooth muscle cells [107]. Genistein has been found to amplify TRPC5-mediated  $\text{Ca}^{2+}$  influx in both TRPC5-overexpressing HEK cells and primary bovine aortic endothelial cells [108].

### 3.2.21. Ginsenoside

Ginsenosides, the active components of the natural medicinal herb ginseng, exhibit antiarrhythmic effects by inhibiting  $\text{Ca}^{2+}$  channels and modulating intracellular  $\text{Ca}^{2+}$  signaling [109]. Specific ginsenosides such as Re, Rb1, and Rg2 have been shown to block LTCC [110]. Using the patch-clamp technique, it was demonstrated that ginsenoside Re increases  $I_{\text{Ks}}$ , reduces  $I_{\text{CaL}}$  and  $\text{Ca}^{2+}$  overload, and protects the heart from I/R injury, through the cGMP-dependent pathway [111]. In rabbit ventricular myocytes subjected to I/R injury, ginsenoside Rb1 was found to block both  $I_{\text{NaL}}$  and  $I_{\text{CaL}}$  currents, decrease action potential amplitude, prevent delayed afterdepolarizations induced by elevated calcium, and protect cardiomyocytes from calcium overload [112]. Furthermore, ginsenosides have been reported to exhibit antiepileptic effects by reducing calcium influx [113].

### 3.2.22. Glycyrrhizic acid

Licorice, derived from the root and rhizome of *Glycyrrhiza uralensis* Fisch., *Glycyrrhiza inflata* Bat., or *Glycyrrhiza glabra* L., contains glycyrrhizic acid as its primary active component [114]. The antiarrhythmic effects of glycyrrhizic acid are attributed to its ability to block sodium and calcium ion channels, to inhibit pacemaker cell automaticity, to slow conduction speed, and to prolong action potential duration and the effective refractory period [115].

### 3.2.23. Guanfu base A

The roots of *Aconitum coreanum* are used to extract guanfu base A, an alkaloid from the *Ranunculaceae* family with antiarrhythmic effects [116]. Guanfu base A acts primarily as a multichannel blocker, including inhibition of sodium channels. It reduces action

potential amplitude and  $V_{\text{max}}$ , and prolongs action potential duration and effective refractory period, converts unidirectional conduction block to bidirectional conduction block, and decreases the incidence of premature contraction and atrioventricular reentry [117]. It has been shown that guanfu base A has antiarrhythmic effects through inhibiting  $I_{\text{Ks}}$  and  $I_{\text{Ca}}$  in the heart [118].

### 3.2.24. Hesperidin and Hesperetin

Hesperidin and its aglycone, hesperetin, are flavanones primarily found in the young fruit of *Citrus* species (*Rutaceae*) and are increasingly consumed in Western diets [119]. Hesperetin is a potent blocker of  $\text{Na}^+$  channels, binding to these channels in their open and inactivated states [119]. Hesperidin also inhibits glutamate release and provides neuroprotection against kainic acid-induced excitotoxicity in the rats hippocampus by inhibiting  $\text{Ca}^{2+}$  channels [120]. In addition, it has been shown that hesperidin can improve ileus by regulating of  $\text{Ca}^{2+}$  channels [121]. Hesperidin treatment and TRPV1 channel inhibition offer a potential therapeutic opportunity for preventive and therapeutic strategies in the treatment of migraine [122]. The modulatory effects of hesperidin on TRP channels was shown in several studies [123,124].

### 3.2.25. Icariin

Flavonoid glucosides, such as icariin, have been isolated from *Epimedium* species (*Berberidaceae*). It has been demonstrated that icariin decreases intracellular calcium overload by inhibiting  $I_{\text{Na}}$  and  $I_{\text{CaL}}$  in the myocardium, exhibiting antiarrhythmic effects [78,125].

### 3.2.26. Kaempferol

The flavonoid kaempferol is widely present in food plants such as cabbage, broccoli, kale, beans, tea and tomatoes [44]. Experimental studies have shown that kaempferol enhances endothelium dependent relaxation in the goat and porcine coronary artery, primarily through the activation of calcium-activated  $\text{K}^+$  channels [44,126,127]. It also exerts analgesic and anti-inflammatory effects *via* TRPV1 in a rat model. TRPV1 inhibition in the peripheral nerve endings reduce calcium and sodium influx, inhibiting nociceptive sensitization [128-131].

### 3.2.27. Magnolol

Magnolol, a polyphenolic compound isolated from Houpu, the bark of *Magnolia officinalis*, has

demonstrated its neuroprotective effects both *in vitro* and *in vivo* [132]. Magnolol acts as a dual inhibitor of voltage-gated sodium and potassium channels, which may partly explain its neuroprotective actions [132]. Magnolol also blocks  $\text{Ca}^{2+}$  channels, thereby inhibiting  $\text{IP}_3$  receptor type 1 and regulating  $\text{IP}_3$ -mediated  $\text{Ca}^{2+}$  release from intracellular stores. Additionally, it activates calmodulin (CaM), leading to the suppression of SK channels. Furthermore, magnolol downregulates PKC, which promotes the opening of  $\text{BKCa}\alpha 1$  and  $\text{BKCa}\beta 3$  channels while closing  $\text{BKCa}\beta 4$  channels. These combined mechanisms contribute to the modulation of intestinal ion secretion [133]. In rats, Zhang *et al.* [134] showed that magnolol inhibits colonic motility by down regulating voltage-gated LTCC in the colonic smooth muscle cells. It has been demonstrated that magnolol inhibits transient receptor potential cation channel C4 (TRPC4) to reduce calcium load and cause smooth muscle relaxation [135].

### 3.2.28. Matrine

Matrine, a quinolizidine alkaloids widely distributed in *Sophora flavescens*, *Sophora alopecuroides*, and other legumes, exhibits antiarrhythmic effect [90]. It prolongs action potential duration and the effective refractory period, decreases heart rate, enhances myocardial contractility, and inhibits ectopic rhythms and atrioventricular reentry by blocking potassium, sodium, and calcium channels [136]. Studies have shown that matrine has concentration-dependent effects on hERG expression [137,138] in rat cardiomyocytes. At low concentrations (1  $\mu\text{mol/l}$ ), matrine promotes hERG expression, whereas at higher concentrations (100  $\mu\text{mol/l}$ ), it suppresses hERG expression, prolongs action potential duration and the effective refractory period of ventricular myocytes, slows spontaneous discharge frequency, and reduces the occurrence of ectopic rhythms [139]. In guinea pig ventricular myocytes, matrine (100  $\mu\text{mol/l}$ ) shortens the ouabain-induced prolonged action potential and prevents the ouabain-induced increase in  $I_{\text{CaL}}$  and  $\text{Ca}^{2+}$  transients. This protective mechanism may involve matrine-mediated inhibition of  $I_{\text{CaL}}$ , thereby reducing calcium overload by competing with ouabain for its binding sites [140]. It has been shown that chronic administration of matrine significantly improves left atrial conduction function, inhibits atrial myofibroblast differentiation, and reduces the duration of atrial fibrillation episodes in rat models. These effects appear to be associated with

increased  $I_{\text{CaL}}$  density and  $\text{Ca}_v1.2$  protein expression in atrial myocyte membranes [141].

### 3.2.29. Nantenine

Alkaloid nantenine is found in plant *Nandina domestica* and several *Corydalis* species. In animal studies, nantenine acts as an antagonist of the  $\alpha$ -adrenergic receptors and the 5-hydroxytryptamine (serotonin) receptor 2A (5-HT<sub>2A</sub>) [142]. Nantenine has also been shown to exert antiepileptic effects by reducing  $\text{Ca}^{2+}$  influx into cells through inhibition of  $\text{Ca}^{2+}$  channels [143].

### 3.2.30. Naringin

Grapefruit juice has a characteristic bitter taste, and its major flavonoid is the flavanone naringin; making it an important dietary source of flavonoids [144]. In mouse hearts, naringin has a negative inotropic effect, which may result from a reduction in both  $I_{\text{Na}}$  and  $I_{\text{CaL}}$  [144].

### 3.2.31. Neferine

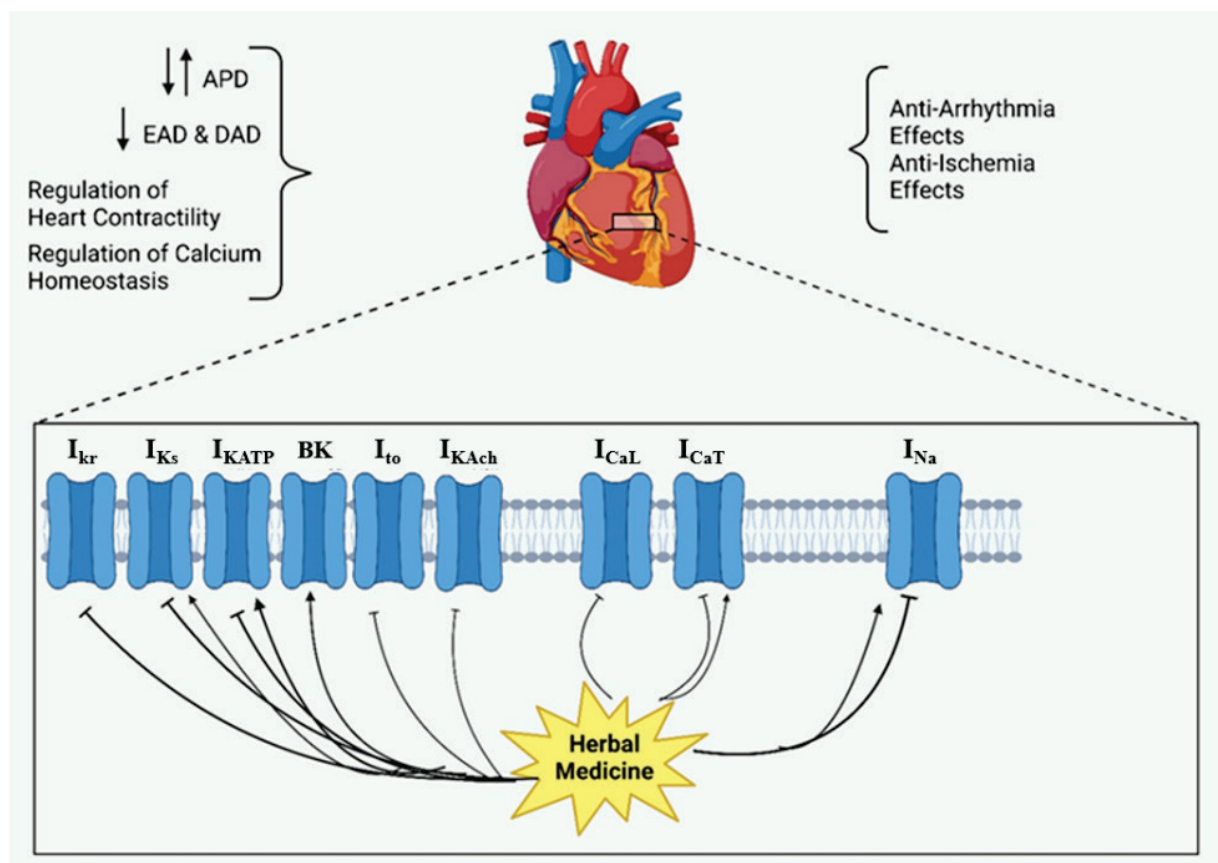
Neferine, an alkaloid extracted from the seeds of *Nymphaeaceae lotus* species, exhibits vasorelaxant, antihypertensive, and antiarrhythmic effects [145]. These properties result from its ability to reduce  $\text{Na}^+$  and  $\text{Ca}^{2+}$  influx while increasing the  $\text{K}^+$  efflux. Neferine has also been shown to prolong QT interval and effectively slow the heart rate [146].

### 3.2.32. Osthole

The coumarin derivative osthole (7-methoxy-8-(3-methyl-2-butenyl) coumarin) is an important bioactive compound found in several medicinal plants and herbs used in traditional medicine. It is clinically utilized and exhibits a wide range of pharmacological and biochemical activities [71]. Osthole has been shown to alleviate epilepsy in mice by increasing  $\text{K}_v1.2$  expression in neurons of the hippocampus CA3 region [143]. It has been shown that osthole alleviates neuropathic pain by suppressing astrocytes activation and associated inflammatory responses *via* the  $\text{PKC}\delta/\text{TRPV4}$  signaling pathway [147]. One study reported that osthole is a potent non-electrophilic agonist of TRPA1 [148].

### 3.2.33. Orientin

The flavonoid monomer orientin is found in several medicinal plants, including *Ocimum sanctum*, bamboo leaves, and *Calendula officinalis*. Orientin has



**Fig. 1.** The effects of herbal medicine or their components on cardiac ion channels. "Arrow" means stimulation, "brake" means inhibition. APD – action potential duration, EAD – early after-depolarization, DAD – delayed after-depolarization.

demonstrated antioxidative, antiapoptotic, antithrombotic and antiarrhythmic effects in experimental models, but it is not an established clinical drug [149,150]. Orientin by blocking LTCC promote cardiac artery dilation [151]. Furthermore, orientin has been reported to inhibit multiple ionic currents in cardiomyocytes including  $I_{Na}$ ,  $I_{to}$ , and  $I_{CaL}$  and to modify their kinetic properties [152].

### 3.2.34. Oxymatrine

Oxymatrine is an alkaloid isolated from the traditional Chinese herb *Sophora flavescens* [153]. Its beneficial antiarrhythmic effect may be related to its ability to shorten action potential duration by reducing  $I_{CaL}$ , increasing  $I_{to}$ , and blocking  $I_{K1}$  [153]. In addition, the antinociceptive effects of oxymatrine mediated through inhibition of voltage-activated  $K^+$  channel [154].

### 3.2.35. Puerarin

Puerarin is a flavonoid compound with antiarrhythmic effect [155], which is extracted from dried root of *Pueraria puerariae*. It has been shown to decrease heart rate, to suppress cardiomyocyte automaticity, and to prolong the effective refractory period and action

potential duration [156]. Puerarin also blocks  $Na^+$  current, which contributes to its antiarrhythmic effect [157]. Puerarin acts as a TRPV4 agonist, induces endothelium-dependent vasodilation in mouse mesenteric arteries, and attenuates blood pressure in high-salt-induced hypertensive mice [158]. Puerarin ameliorates cisplatin-induced ototoxicity and blocks cellular apoptosis by inhibiting activated TRPV1/IP3R1/p65 pathway, blocking the induction of calcium overload and excessive ROS production [159]. On the other hand, some studies revealed the inhibitory effects of puerarin on TRP channels [160,161].

### 3.2.36. Protopine

The isoquinoline alkaloid protopine is obtained from the *Corydalis tubers*. It has been reported to exhibit antiarrhythmic, antihypertensive, negative inotropic and other cardiovascular protective effects [162]. It has been demonstrated that protopine inhibits  $I_{CaL}$  in a concentration-dependent manner, with no apparent effect on transient calcium current ( $I_{CaT}$ ) in isolated guinea pig ventricular myocytes [163].

### 3.2.37. Quercetin

Quercetin, a flavonoid commonly found in fruits, vegetables, and leaves [164], exhibits cardioprotective, antihypertensive and other biological activities [165,166]. In rats, quercetin acts as an LTCC inhibitor, producing cardioprotective effects by reducing  $\text{Ca}^{2+}$  influx and myocardial contractility [167]. Additionally, isoquercetin has been shown to exert antiepileptic effects by modulating the GABA<sub>A</sub> channel and  $\text{Cl}^-$  current [143]. It has also been shown that quercetin increase colonic contraction by stimulating calcium channels [168].

### 3.2.38. Quinidine

Quinidine, an alkaloid derived from the bark of the cinchona tree, has been used to treat both atrial and ventricular arrhythmias [10]. It has been reported to block  $I_{Ks}$ ,  $I_{to}$ , and ultrarapid delayed-rectifier potassium ( $I_{Kur}$ ) currents, and these actions contribute to its antiarrhythmic effects [169,170].

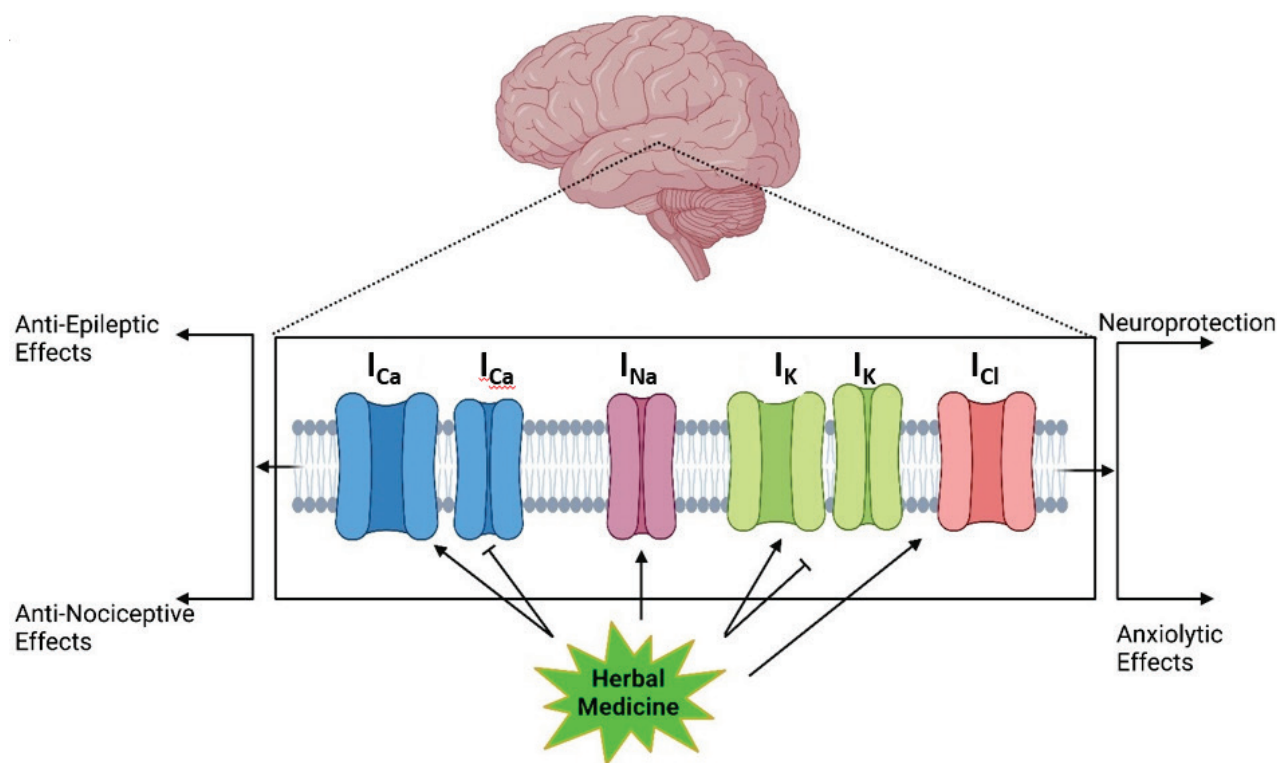
### 3.2.39. Resveratrol

Resveratrol, extracted from dry roots of

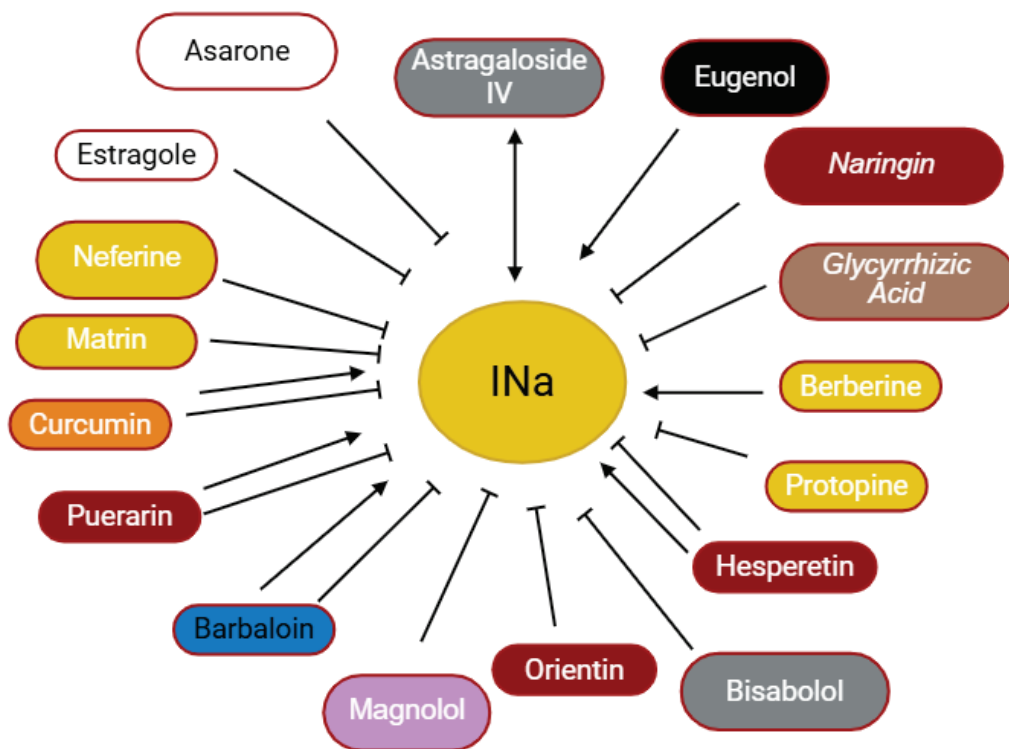
*Polygonum cuspidatum* Sieb et Zucc. (Polygonaceae) has antiarrhythmic properties [171]. Resveratrol regulates potassium, sodium and calcium ion channels,  $K_{ACh}$ , and hHCN4, which is highly expressed in sinoatrial node pacemaker cells, where it generates the funny current ( $I_f$ ), thereby slowing heart rate, prolonging the effective refractory period of cardiomyocytes, and preventing early and delayed after depolarization [172,173]. It has been shown that resveratrol modulates diabetes-induced neuropathic pain, apoptosis, and oxidative neurotoxicity in mice through TRPV4 channel inhibition [174].

### 3.2.40. Tetrandrine

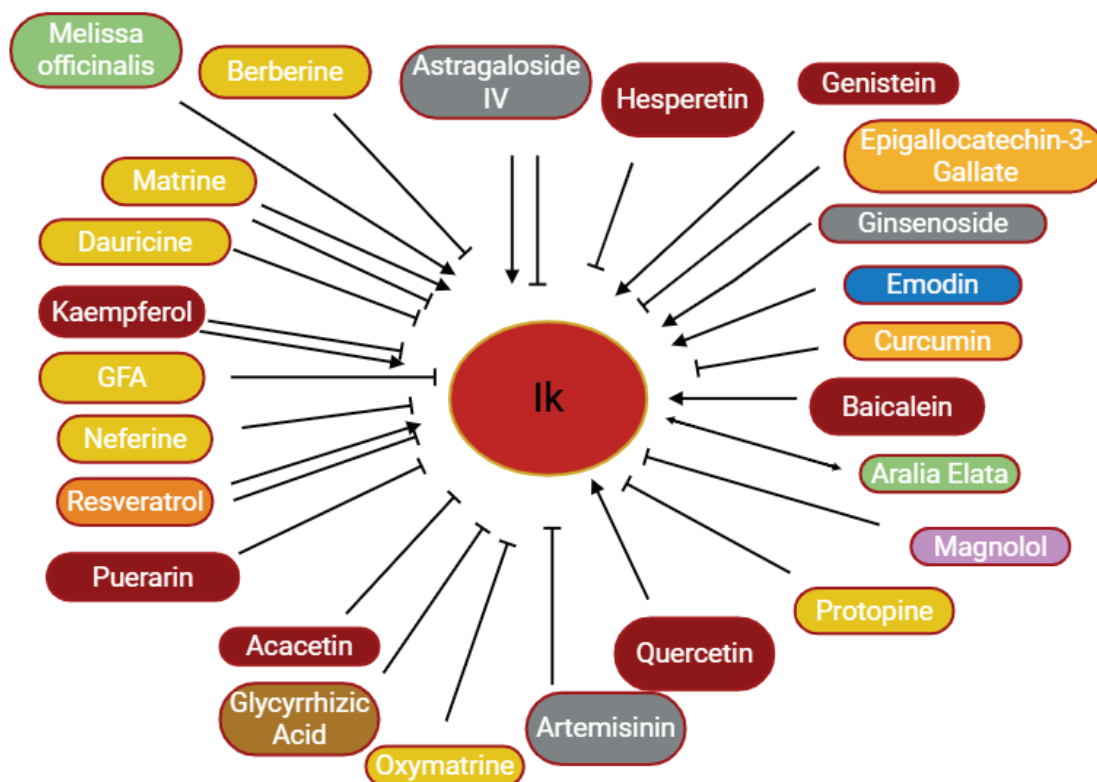
Tetrandrine is a dibenzyl isoquinoline alkaloid derived from the rhizomes of *Trichosanthes* Merr. Chun. and from roots of *Stephania discolor*, *Stephania tetrandra* and *Aristolochia heterophylla*. It is recognized as an antiarrhythmic agent due to its ability to inhibit calcium, potassium, and sodium channels [175]. Tetrandrine can decrease the heart rate, inhibit atrioventricular conduction, and prolong cardiomyocytes effective refractory period [176].



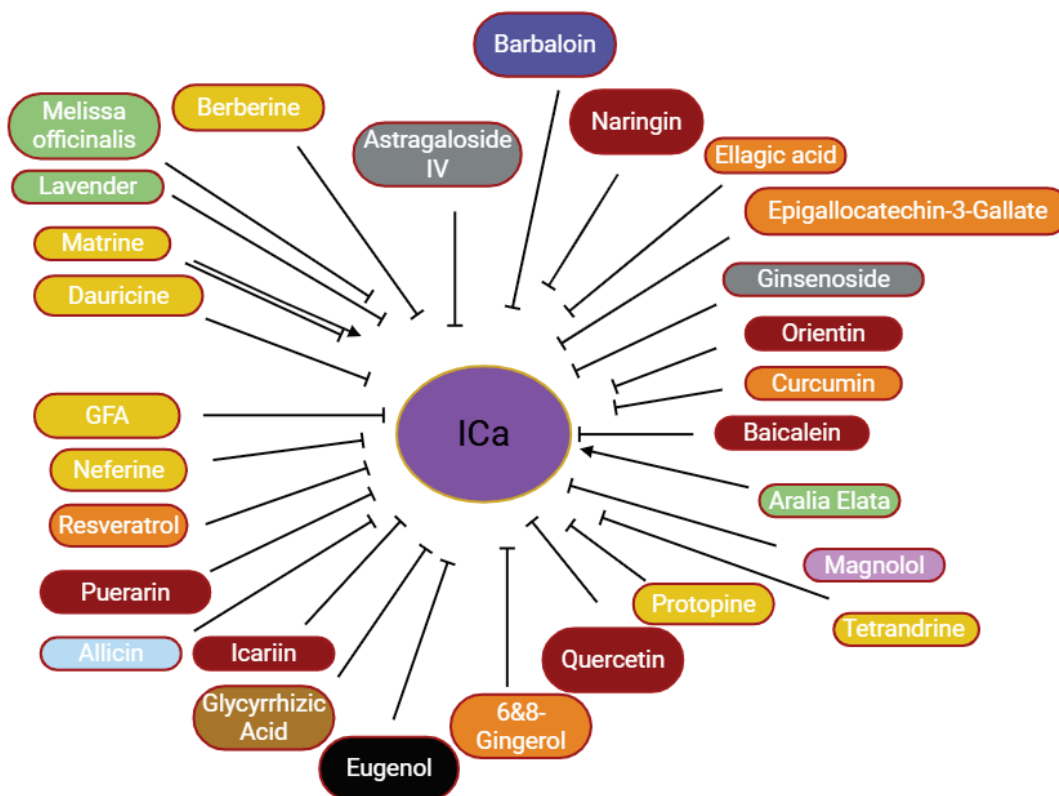
**Fig. 2.** The effects of herbal medicine or their components on brain ion channels. "Arrow" means stimulation, "brake" means inhibition.



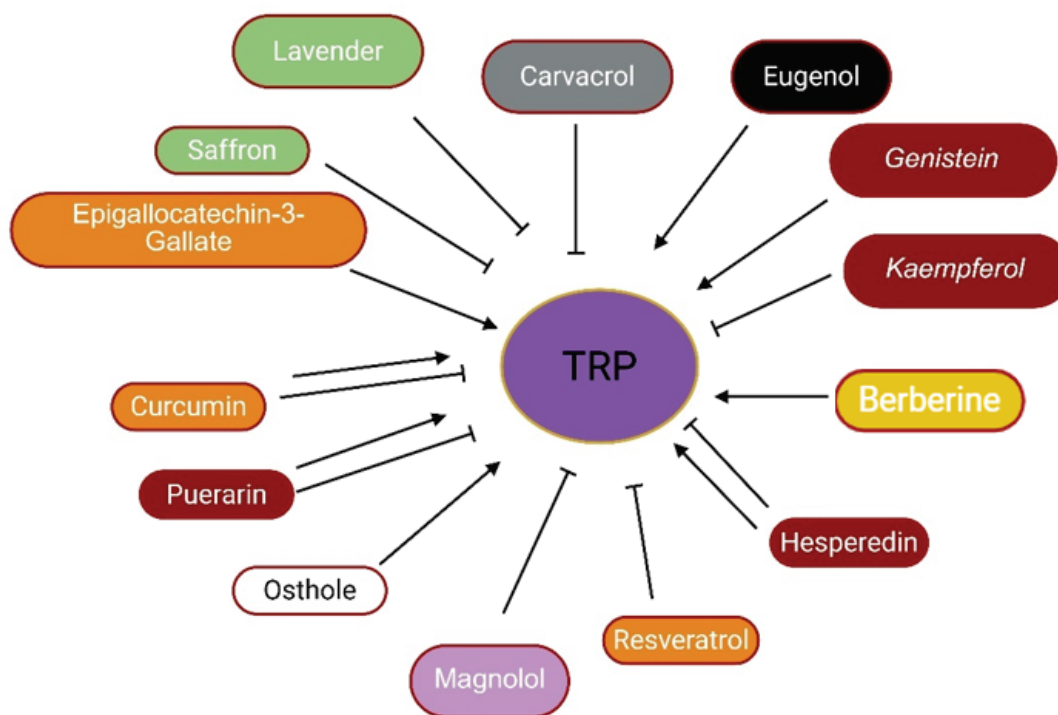
**Fig. 3.** The effects of some herbal medicine or their components on sodium current. "Arrow" means stimulation, "brake" means inhibition. Yellow boxes: Alkaloids, Red boxes: Flavonoids, Black box: Benzene, Blue box: Quinone, Gray boxes: Terpenoids, Orange boxes: Polyphenols, Brown box: Saponin, Pink box: Lignan, White boxes: Phenyl propene.



**Fig. 4.** The effects of some herbal medicine or their components on potassium current. "Arrow" means stimulation, "brake" means inhibition. Yellow boxes: Alkaloids, Red boxes: Flavonoids, Green boxes: Plants, Blue box: Quinone, Gray boxes: Terpenoids, Orange boxes: Polyphenols, Brown box: Saponin, Pink box: Lignan.



**Fig. 5.** The effects of some herbal medicine or their components on calcium current. "Arrow" means stimulation, "brake" means inhibition. Yellow boxes: Alkaloids, Red boxes: Flavonoids, Green boxes: Plants, Blue box: Quinone, Gray boxes: Terpenoids, Orange boxes: Polyphenols, Brown box: Saponin, Pink box: Lignan, Black box: Benzene, Light blue box: Sulfur compounds.



**Fig. 6.** The effects of some herbal medicine or herbal components on TRP channels. "Arrow" means stimulation, "brake" means inhibition. Red boxes: Flavonoids, Black box: Benzene, Gray box: Terpenoids, Orange boxes: Polyphenols, Pink box: Lignan, White box: Coumarin, Green boxes: plants, yellow box: Alkaloid.

**Table 3.** The effects of herbal compounds on cardiac ion channels

Compound	Effect on ion channel conductivity	Main effect	Mechanisms of action	Species	Dose/Concentration	Ref
<b>6-gingerol</b>	ICaL ↓	Antiarrhythmic effect	Increased action potential duration	Rat	300 μmol/l	[55]
<b>8-gingerol</b>	LTCC ↓	Antiarrhythmic effect	Myocardial anti-ischemic effects	Rat	10 and 20 mg/kg	[59]
<b>Acacetin</b>	IK <sub>Kr</sub> , IK <sub>r</sub> , KACH, Ito and SK ↓ Ito ↓	Antiarrhythmic effect Improved atrial fibrillation	Increased action potential duration Increased refractory period of the fibrillating atrium	Dog Human Rabbit	5-10 μM 2.5, 5 and 10 mg/kg	[60, 180, 193]
<b>Aloperine</b>	IK <sub>r</sub> ↑ INa ↓	Antiarrhythmic effect	Increased action potential duration	Rat	10 mg/kg 3, 10 and 30 μM	[68, 70]
<b>Aralia elata</b>	ICaL ↑ Ito ↔	Improvement of cardiomyopathy	Increased left ventricular systolic pressure Increased contractility	Rat Dog	19.6 mg/kg 30 and 60 mg/kg	[14, 194]
<b>Artemisinin</b>	IK <sub>1</sub> and Ito ↓ HCN ↓	Antiarrhythmic effect	Increased action potential duration	Dog Frog	5 and 50 μmol/l	[181, 195, 196]
<b>Astragaloside IV</b>	IK <sub>1</sub> and IKATP ↑ IK ↓ ICaL ↓ CaSR ↓	Antiarrhythmic effect Decreased infarct size	Increased action potential duration Reduced apoptosis	Guinea pig Rat	1×10 <sup>-6</sup> M 16, 32 and 64 μM	[74, 197-200]
<b>Baicalein</b>	ICaL ↓ BK ↑	Protection of against myocardial infarction injury	Dampening of intracellular calcium Reduced amplitude of voltage-dependent calcium channels currents	Mouse	30 and 60 mg/kg 100 μM	[75, 182]
<b>Barbaloin</b>	ICaL ↓ INa ↓	Antiarrhythmic effect	Reduced delayed and early after-depolarization	Rabbit	100 and 200 μmol/l	[77]
<b>Berberine</b>	IK <sub>r</sub> , IK <sub>s</sub> , IKATP and IK <sub>1</sub> ↓ ICaL and ICaT ↓ hHCN4 ↓	Antiarrhythmic effect	Increased action potential duration Decreased rate of pacemaker and diastolic depolarization	Rat Guinea pig	3-100 μmol/l 10 and 20 mg/kg i.p. 10 and 30 μmol/l	[82, 178, 179, 201-203]
<b>Carvacrol</b>	TRPM7 ↓ IK <sub>v</sub> ↑	Antiarrhythmic effect	Decrease in conduction velocity	Rabbit Human	100 μM	[87, 204]
<b>Curcumin (Turmeric)</b>	INa, ICaL and IK <sub>r</sub> ↓	Prevention of ischemia/reperfusion-induced arrhythmias	Shortened action potential duration Suppressed early and delayed afterdepolarization	Rabbit	30 μmol/l	[42]
<b>Dauricine</b>	IK <sub>r</sub> and IK <sub>s</sub> ↓ ICaL ↓	Reduced calcium concentration and Inhibition of Ca <sup>2+</sup> -ATPase activity	Increased action potential duration Increased atrial effective refractory period	Guinea pig Rabbit	1, 3, 10, 30, 100 μmol/l	[90, 205, 206]
<b>Elatoside C</b>	SERCA2 ↑ mPTPs ↓	Increases in left ventricular systolic pressure, ± dP/dt <sub>max</sub> , and heart rate	Attenuating calcium overload	Rat		[93]
<b>Ellagic acid</b>	ICaL ↓	Protection against coronary artery diseases	Negative inotropic effect	Rat	23 nM	[91]

<b>Emodin</b>	BK and IK1 ↑	Improved hypertension	Aorta relaxation Reduced QT interval	Rat		[95, 207]
<b>Epigallocatechin-3-gallate</b>	ICaL ↓ IKr, IKs, Ito and IKATP ↓ TRPA1 ↑	Antiarrhythmic effect	Reduced action potential duration	Rat	10 μM 30 μM	[100, 101]
<b>Genistein</b>	BK and IKur ↓ IKATP ↑	Improved atherosclerosis and hypertension	Vascular relaxation	Rat Human	50 μmol/l 30 μmol/l 5 mg/kg	[107, 208-210]
<b>Ginsenoside</b>	LTCC ↔ IKs ↑ INa ↓ IK1 and IKv ↓	Antiarrhythmic effect	Reduced action potential duration Reduced amplitude	Guinea pig Rabbit	5, 10 and 20 mM	[111, 112]
<b>Glycyrrhizic acid</b>	INa ↓ ICaL ↓ Ito, IKr and IKs ↓	Antiarrhythmic effect	Reduced conduction speed Increased action potential duration Increased effective refractory period	Rat Guinea pig Ventricular myocytes Xenopus oocytes	10 mg/kg 3.5 mg/kg	[114, 211]
<b>Guanfu base A</b>	IKs and IKr ↓ Kir and Ito ↔ LTCC ↓	Antiarrhythmic effect	Reduced action potential amplitude Increased action potential duration	Human Dog Monkey, Rabbit	100, 400, 1000, 2500 μmol/l 25, 125, 250, 1000 μmol/l	[118, 196, 212, 213]
<b>Hesperetin</b>	INa ↓ IKs ↓	Antiarrhythmic effect	Decrease in conduction velocity	Human Rabbit Dog		[119, 214]
<b>Icariin</b>	LTCC ↓	Antiarrhythmic effect	Reduced delayed after depolarization	Rabbit	5 and 10 μM	[125]
<b>Kaempferol</b>	BK ↑ IKv ↓	Attenuated hypertension	Vascular relaxation	Rat Goat Pig	10-7-10-4 M 3 × 10-6 M	[44, 215]
<b>Matrine</b>	IKr ↓ IKM3 ↓ KCNB1 and KCNJ2 ↑ LTCC ↑ INa ↓	Antiarrhythmic effect Reduced atrial fibrillation	Increased action potential duration Reduced action potential amplitude	Rat Guinea pig	1 and 100 μmol/l 15, 30, 45 mg/kg i.v. 50, 100, 200 mg/kg p.o.	[139, 177]
<b>Melissa officinalis</b>	LTCC ↓ IK ↑	Antiarrhythmic effect	Modulation of ECG and heart electrical system	Rat	50, 100 and 200mg/kg	[21, 25]
<b>Naringin</b>	ICaL ↓ INa ↓	Negative inotropic effects	Regulation of heart contractility	Mouse Rat	30-100 μM 10-30 μM	[127, 144]
<b>Neferine</b>	IKr ↓ ICaL ↓ INa ↓	Antiarrhythmic effect	Increased action potential duration Increased effective refractory period Reduced action potential amplitude	Rat In vitro	10 and 30 μmol/l	[216, 217]
<b>Orientin</b>	ICaL ↓ INa ↓ Ito ↓	Antiarrhythmic effect	Reduced action potential duration	Mouse	40 mg/kg	[152]

<b>Panax notoginseng saponins</b>	KCNN3 ↓ SK ↓	Antiarrhythmic effect Improved atrial fibrillation	Increased action potential duration Reduced calcium release from sarcoplasmic reticulum	Rat	150 mg/kg i.p.	[218, 219]
<b>Protopine</b>	ICaL ↓ ICaT ↔ IK1 and IK ↓ INa ↓	Antiarrhythmic effect	Increased action potential duration	Guinea pig	25, 50 and 100 μM	[163]
<b>Puerarin</b>	IKs, IK1 and Kir ↓ LTCC ↓ INa ↓	Antiarrhythmic effect Improved atrial fibrillation	Increased action potential duration	Rat Mice	0.01, 0.1 and 1.0 mmol/l 1.2 mmol/l	[196, 220-222]
<b>Quercetin</b>	ICaL ↓ KCNQ ↑	Anti-ischemic effects	Regulation of calcium homeostasis	Rat	60 mg/kg	[167, 183]
<b>Quinidine</b>	INa ↓ IKATP and IKr ↓	Antiarrhythmic effect	Increased action potential duration	Hamster Rat Guinea pig	9.7 μM	[223, 224]
<b>Resveratrol</b>	IKr and IKACH ↓ IKATP ↑ ICaL ↓ HCN4 ↓	Inhibition of cardiac contractility Reduced heart rate	Increased action potential duration Increased effective refractory period Inhibition of delayed and early after-depolarization	Rat Guinea pig	50, 100 and 500 μM 14.02 μmol/l 1, 50, 100 μmol/l	[196, 225-229]
<b>Salvianic acid A</b>	ICaL ↓	Antiarrhythmic effect	Decreased myocardial contractility	Rat	14.7 μM	[53]
<b>Tetrandrine</b>	BK ↓ LTCC and TTCC ↓	Antiarrhythmic effect	Decreased action potential duration Reduced Ca <sup>2+</sup> influx into the cell from sarcolemma Decreased Ca <sup>2+</sup> uptake into the sarcoplasmic reticulum	Rabbit Rat	7.5 and 15 μmol/l 30 μmol/l	[230-233]

#### 4. Discussion

Ion channels play essential roles in regulating electrical properties and synchronized contraction of the heart. They are also essential for the function of central and peripheral nervous systems and the transmission of electrical signals among neurons. The digestive system also relies heavily on ion channels to regulate gastrointestinal motility, secretion and nutrient absorption. Therefore, disturbances in the normal activity of these channels, whether genetic or acquired, can impair the function of multiple organ systems.

Medicinal plants, as natural healing resources, have been widely used throughout history to manage various disorders and are commonly used as alternative medicines today. This review summarizes available information regarding the effects of medicinal plants and/or their active constituents on ion channels in different physiological systems.

In the cardiovascular system, the activation of

voltage-dependent sodium channel initiates action potentials by allowing Na<sup>+</sup> influx into cardiac myocytes. Potassium channels are responsible for repolarization, while calcium ions are involved in pacemaker activity, action potential formation, heart rate regulation, and myocardial contraction. Several medicinal plants or their phytochemicals affect multiple cardiac ion channels or different subunits of the same channel. For example, matrine inhibits sodium channel conductance as well as I<sub>CaL</sub> and I<sub>K</sub> [141,177]. Barbaloin blocks I<sub>Na</sub> and I<sub>Ca</sub> [77]. Berberine inhibits I<sub>Kr</sub>, I<sub>Ks</sub>, I<sub>K1</sub>, hHCN4 currents and both L-type and T-type calcium channels [82,178,179], contributing to its antiarrhythmic effects. Several herbal compounds exert their cardioprotective or antiarrhythmic effects by modulating potassium channels, such as puerarin [157], acacetin [180], artemisinin [181], and protopine [163] through channel blockade or *Melissa officinalis* [25] and ginsenoside [112] through potassium channels activation. In addition, components such as 6-gingerol [55], icariin [125], and 8-gingerol [59] show

**Table 4.** The effects of herbal medicine on brain and gastrointestinal ion channels

Compound	Effect on ion channels	Brain effects	GI effects	Species	Dose	Ref
<b>Allicin</b>	NTCC and P/QTCC ↓	Neuroprotective	-	Rat	20 μM	[66]
<b>Asarone</b>	ICa ↓ NaV1.2 ↓	Antiepileptic effect	-	Mouse	20 mg/kg	[73]
<b>Curcumin (Turmeric)</b>	INa and IKATP ↓	Nociceptive effect Antidepressant effect	Protection against gastric ulcer	Rat	100 mg/kg	[46, 49, 50]
<b>Estragole</b>	INa ↓	Anxiolytic effect	-	Rat	30 mg/kg	[186]
<b>Eugenol</b>	INa and ICa ↓ ICl ↓ TRPA1 ↑	Analgesic and antiepileptic effects	Antidiarrheal effect	Rat	2 mM 100 mg/kg	[104, 191, 234-236]
<b>Ginsenoside</b>	ICaL ↓ IKATP ↓	Antiepileptic effect Improvement of Alzheimer disease	Improved small intestine motility	Rat Mouse	20 μmol/l	[113, 188, 190]
<b>Hesperidin</b>	NTCC ↓	Neuroprotection Anticonvulsive effects	Improved ileus	Rat	10 and 50 mg/kg 500 mg/kg	[120, 121, 143, 237]
<b>Lavender</b>	ICaT ↓ TRPA1 ↓	Anxiolytic Antinociceptive	-	Mouse <i>in vitro</i>	30 μmol/l	[15, 17, 18]
<b>Magnolol</b>	INa, ICaL and IK ↓ TRPC4 ↓	Neuroprotection	Antidiarrhea Inhibits colonic motility	Rat Mouse	100, 300 and 500 mg/kg	[132-134]
<b>Melissa officinalis</b>	ICl ↑ ICa ↓	Anxiolytic activity	Reduced ileum contraction	Mouse	25 mg/kg	[23]
<b>Naringenin</b>	NaV1.8 ↓ BK ↑	Analgesic effect Improve motor neuron function	-	Rat <i>in vitro</i>	100 and 10 μM	[184, 238]
<b>Oxymartine</b>	IK ↓	Antinociceptive effects	-	Mouse	150 mg/kg	[154]
<b>Quercetin</b>	GABA <sub>A</sub> -Cl <sup>-</sup> channels ↓ Acid-sensing ion channels ↓ INa ↓ ICaL and IK ↑	Antiepileptic Improved cognitive deficits	Increased colon contractility	Rat Guinea pig	2 μM 30 μM	[143, 168, 239-241]
<b>Salvia miltiorrhiza</b>	Ca <sup>2+</sup> /calmodulin pathway ↑		Increased ileum tonic contraction	Rat Mouse	40 μmol/ml	[39, 242, 243]
<b>α(-)-Bisabolol</b>	INa ↓ IcaT ↓ IK ↑	Antinociceptive effects	-	Mouse	0.5, 1, 5 and 10 mM 1 μg 300 μg	[185, 244, 245]

antiarrhythmic effects, while baicalein [182], ellagic acid [91], and quercetin [183] protect against myocardial injury by blocking calcium channels. However, some agents, such as *Aralia elata* [14], increase calcium channel conductance and promote cardiomyopathy. The effects of plants or their effective compounds on cardiac ion channels were summarized in Table 2.

Based on the findings of various studies, it can be suggested that different herbal medicines or their derivatives exert their effects on neurological disorders through decreasing or increasing the conductance of specific ion channels (Table 3). For example, curcumin and naringenin exert anti-nociceptive and analgesic effects by inhibiting sodium channels [46,184]. Alpha-bisabolol show similar effects by blocking sodium and potassium channels [185], whereas lavender mediates its analgesic action through modulation of calcium channels and interaction with NMDA receptors [18]. The anxiolytic effects of estragol has been attributed to its sodium-channel blocking effects [186]. In addition, several herbal agents, such as asarone, nantenin and ginsenoside exhibit anti-epileptic properties through attenuating calcium channel conductance [73,187,188]. Components such as berberine [86], linalool [19], safranal [34], kaempferol [128,131], osthole [147] and resveratrol [174] exhibit analgesic effects, and magnolol [135] causes smooth muscle relaxation by inhibiting TRP channels. However, capsaicin [189], eugenol [105,106] and epigallocatechin-3-gallate [102,103] activate TRP channels and temporary enhance nociceptive signaling, vasodilation and blood pressure reduction.

Despite the extensive use of plant-derived compounds for treating gastrointestinal disorders in traditional medicine, knowledge of their mechanisms, particularly their effects on ion channels remains limited. However, it is reported that some herbal agents such as magnolol inhibit colonic motility by downregulation of LTCC [134]. Ginsenoside and eugenol exhibit antiarrhythmic effects through potassium channels modulation [190] and calcium channels inhibition [191], respectively, in the gastrointestinal smooth muscles, thereby contributing to the regulation of gastric motility and alleviation of indigestion symptoms [190].

### Gaps in Research and Foresight

Despite the substantial amount of information available in this field, our understanding of how

medicinal plants influence ion channels remains limited, much like seeing only the tip of a floating iceberg. Given the diversity of subunits within each ion channel type, the effects of plant-derived components on each specific subtype must be investigated in detail. Furthermore, considering the distribution and diversity of ion channels subunits, which vary among tissues, these compounds may elicit distinct outcomes depending on the tissue type involved. These require comprehensive, large-scale studies to determine how various phytochemicals modulate individual channel subtypes and how these interactions translate to physiological effects on organ function and excitable cells. In addition, the existing findings are derived from experimental studies and due to the lack of clinical validation, the capacity to use laboratory results in real-world clinical settings is limited. For example, many herbal polyphenols, such as flavonoids, are characterized by inherently low oral bioavailability, largely due to poor intestinal absorption, extensive first-pass metabolism, and rapid systemic clearance [192]. As a result, the plasma concentrations achievable in humans following dietary intake or oral supplementation are typically far lower than the concentrations commonly used in *in vitro* assays, which frequently range from  $\geq 10$ –100  $\mu\text{M}$ . This discrepancy is important to acknowledge when interpreting mechanistic findings, as some of the ion channel modulatory effects reported at high micromolar concentrations may not be replicable under physiological *in vivo* conditions. Nevertheless, such data remain valuable for identifying potential molecular targets and for understanding the pharmacodynamic potential of these compounds under conditions of enhanced delivery or structural optimization.

Therefore, further studies are essential to elucidate the molecular mechanisms and tissue-specific actions of medicinal plants. Consequently, comprehensive clinical and pharmacological studies must be conducted to confirm the bioavailability, efficacy, doses and safety of these compounds for clinical uses. Such studies will be invaluable for clarifying the broader implications of phytochemicals in complementary medicine and their clinical potential in treating a range of disorders, such as cardiovascular, neurological, and gastrointestinal diseases.

### Conclusion

Overall, this review highlights how specific

phytochemicals in medicinal plants interact with a variety of ion channels. Certain plants or their active constituents can influence multiple channel types or subunits, whereas others act more selectively. Given the diversity of ion channel subunits, their tissue-specific distribution, and their distinct activation mechanisms under physiological conditions, it is clear that herbal medicines can produce a wide array of tissue-dependent physiological and pharmacological effects. Consequently, the use of herbal agents may lead to different outcomes depending on the

target tissue and the specific channels involved.

### Conflict of Interest

There is no conflict of interest.

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### Abbreviations

5-HT2A	5-hydroxytryptamine (serotonin) receptor 2A
APD	Action potential duration
BK	Large-conductance Ca <sup>2+</sup> -activated potassium channel
CaM	Calmodulin
CaSR	Calcium-sensing receptor
cGMP	Cyclic guanosine monophosphate
CGRP	Calcitonin gene-related peptide
CRAC	Ca <sup>2+</sup> release-activated Ca <sup>2+</sup> channel
DAD	Delayed after depolarization
EAD	Early after depolarization
ECG	Electrocardiogram
GABA	Gamma-aminobutyric acid
HCN	Hyperpolarization-activated cyclic nucleotide-gated channel
hERG	Human ether-à-go-go related gene
I/R	Ischemia/reperfusion
I <sub>Ca</sub>	Calcium current
I <sub>CaL</sub>	L-type calcium current
I <sub>CaT</sub>	T-type calcium current
I <sub>Cl</sub>	Chloride current
I <sub>f</sub>	Funny current (pacemaker current)
I <sub>K</sub>	Potassium current
I <sub>K1</sub>	Inward rectifier potassium current
I <sub>KACh</sub>	Potassium current via K <sub>ACh</sub> channel
I <sub>KATP</sub>	Potassium current via K <sub>ATP</sub> channel
I <sub>Kr</sub>	Rapid delayed-rectifier potassium current
I <sub>Ks</sub>	Slow delayed-rectifier potassium current
I <sub>KM3</sub>	M3-receptor-mediated potassium current.
I <sub>Kur</sub>	Ultrarapid delayed-rectifier potassium current
I <sub>KV</sub>	Voltage-gated potassium channel
I <sub>Na</sub>	Sodium current
I <sub>NaL</sub>	Late sodium current
I <sub>NaT</sub>	Transient sodium current
I <sub>to</sub>	Transient outward potassium current
K <sub>ACh</sub>	Acetylcholine-activated (muscarinic) potassium channel
K <sub>ATP</sub>	ATP-sensitive potassium channel

KCNH2	Potassium voltage-gated channel subfamily H member 2
KCNN3	Potassium calcium-activated channel subfamily N member 3
KCNQ	Voltage-gated Kv7 potassium channel
K <sub>ir</sub>	Inward-rectifier potassium channel
K <sub>v</sub>	Voltage-gated potassium channel
LDL	Low-density lipoprotein
L-type Ca <sup>2+</sup> channel (LTCC)	Large voltage-gated calcium channel
MAPK	Mitogen-activated protein kinase
mPTPs	Mitochondrial permeability transition pores
N-type Ca <sup>2+</sup> channel (NTCC)	Neuronal voltage-gated calcium channel
NMDA	N-methyl-D-aspartate
NR2B	N-methyl D-aspartate receptor subtype 2B
PKC	Protein kinase C
P/Q-type Ca <sup>2+</sup> channel (P/QTCC)	P/Q-type voltage-gated calcium channel
ROS	Reactive oxygen species
SCN5A	Sodium channel protein type 5 subunit alpha
SERCA2	Sarcoplasmic/endoplasmic reticulum calcium ATPase 2
SK	Small-conductance Ca <sup>2+</sup> -activated K <sup>+</sup> channel
T-type Ca <sup>2+</sup> channel (TTCC)	Transient voltage-gated calcium channel
TRP	Transient receptor potential channel
TRPA1	Transient receptor potential cation channel A1
TRPC4	Transient receptor potential cation channel C4

## References

1. Kefauver J, Ward A, Patapoutian A. Discoveries in structure and physiology of mechanically activated ion channels. *Nature* 2020;587:567-576. <https://doi.org/10.1038/s41586-020-2933-1>
2. András V, Tomek J, Nagy N, Virág L, Passini E, Rodriguez B, Baczkó I. Cardiac transmembrane ion channels and action potentials: cellular physiology and arrhythmogenic behavior. *Physiol Rev* 2021;101:1083-1176. <https://doi.org/10.1152/physrev.00024.2019>
3. Voigt N, Friedrich A, Bock M, Wettwer E, Christ T, Knaut M, Strasser RH, Ravens U, Dobrev D. Differential phosphorylation-dependent regulation of constitutively active and muscarinic receptor-activated IK,ACh channels in patients with chronic atrial fibrillation. *Cardiovasc Res* 2007;74:426-437. <https://doi.org/10.1016/j.cardiores.2007.02.009>
4. Voigt N, Maguy A, Yeh Y-H, Qi X, Ravens U, Dobrev D, Nattel S. Changes in IK,ACh single-channel activity with atrial tachycardia remodelling in canine atrial cardiomyocytes. *Cardiovasc Res* 2008;77:35-43. <https://doi.org/10.1093/cvr/cvm051>
5. da Costa R, Passos GF, Quintão NLM, Fernandes ES, Maia J, Campos MM, Calixto JB. Taxane-induced neurotoxicity: Pathophysiology and therapeutic perspectives. *Br J Pharmacol* 2020;177:3127-3146. <https://doi.org/10.1111/bph.15086>
6. Joukar S. A comparative review on heart ion channels, action potentials and electrocardiogram in rodents and human: extrapolation of experimental insights to clinic. *Lab Anim Res* 2021;37:25. <https://doi.org/10.1186/s42826-021-00102-3>
7. Lipscombe D. L-type calcium channels: highs and new lows. *Circ Res* 2002;90:933-935. <https://doi.org/10.1161/01.RES.0000019740.52306.92>
8. Palmisano VF, Anguita-Ortiz N, Faraji S, Nogueira JJ. Voltage-gated ion channels: structure, pharmacology and photopharmacology. *Chemphyschem* 2024;25:e202400162. <https://doi.org/10.1002/cphc.202400162>
9. Vandenberg JI, Perry MD, Perrin MJ, Mann SA, Ke Y, Hill AP. hERG K<sup>+</sup> channels: structure, function, and clinical significance. *Physiol Rev* 2012; 92:1393-1478. <https://doi.org/10.1152/physrev.00036.2011>

10. Yan Z, Zhong L, Zhu W, Chung SK, Hou P. Chinese herbal medicine for the treatment of cardiovascular diseases - Targeting cardiac ion channels. *Pharmacol Res* 2023;192:106765. <https://doi.org/10.1016/j.phrs.2023.106765>
11. Waszkielewicz AM, Gunia A, Szkaradek N, Sloczynska K, Krupinska S, Marona H. Ion channels as drug targets in central nervous system disorders. *Curr Med Chem* 2013;20:1241-1285. <https://doi.org/10.2174/0929867311320100005>
12. Rajizadeh MA, Najafipour H, Bejeshk MA. An updated comprehensive review of plants and herbal compounds with antiasthmatic effect. *Evid Based Complement Alternat Med* 2024;2024:5373117. <https://doi.org/10.1155/2024/5373117>
13. Rajizadeh MA, Bejeshk MA, Aminizadeh A, Yari A, Rostamabadi F, Bagheri F, Najafipour H, Nematollahi MH, Amirhosravi A, Mehrabani M. Inhalation of spray-dried extract of *Salvia rosmarinus* Spenn alleviates lung inflammatory, oxidative, and remodeling changes in asthmatic rats. *Pharmacology* 2024;109:10-21. <https://doi.org/10.1159/000534392>
14. Wang M, Xu X, Xu H, Wen F, Zhang X, Sun H, Yao F, Sun G, Sun X. Effect of the total saponins of *Aralia elata* (Miq) Seem on cardiac contractile function and intracellular calcium cycling regulation. *J Ethnopharmacol* 2014;155:240-247. <https://doi.org/10.1016/j.jep.2014.04.016>, <https://doi.org/10.1016/j.jep.2014.05.024>
15. López V, Nielsen B, Solas M, Ramírez MJ, Jäger AK. Exploring pharmacological mechanisms of lavender (*Lavandula angustifolia*) essential oil on central nervous system targets. *Front Pharmacol* 2017;8:280. <https://doi.org/10.3389/fphar.2017.00280>
16. El Alaoui C, Chemin J, Fechtali T, Lory P. Modulation of T-type Ca<sup>2+</sup> channels by Lavender and Rosemary extracts. *PLoS One* 2017;12:e0186864. <https://doi.org/10.1371/journal.pone.0186864>
17. Schuwald AM, Nöldner M, Wilmes T, Klugbauer N, Leuner K, Müller WE. Lavender oil-potent anxiolytic properties via modulating voltage dependent calcium channels. *PLoS One* 2013;8:e59998. <https://doi.org/10.1371/journal.pone.0059998>
18. Naufal AH, Virgiriina RP, Fatchiyah F. Molecular interaction of lavender (*Lavandula angustifolia* Mill) essential oil compounds as potential anxiolytic against  $\alpha_2\delta$  subunit voltage-gated calcium channel. *Berkala Penelitian Hayati (J Biol Res)* 2023;29:1-11. <https://doi.org/10.23869/bphjbr.29.1.20231>
19. Hashimoto M, Takahashi K, Ohta T. Inhibitory effects of linalool, an essential oil component of lavender, on nociceptive TRPA1 and voltage-gated Ca<sup>2+</sup> channels in mouse sensory neurons. *Biochem Biophys Rep* 2023;34:101468. <https://doi.org/10.1016/j.bbrep.2023.101468>
20. Draginic N, Jakovljevic V, Andjic M, Jeremic J, Srejovic I, Rankovic M, Tomovic M, Nikolic Turnic T, Svistunov A, Bolevich S. *Melissa officinalis* L. as a nutritional strategy for cardioprotection. *Front Physiol* 2021;12:661778. <https://doi.org/10.3389/fphys.2021.661778>
21. Joukar S, Asadipour H, Sheibani M, Najafipour H, Dabiri S. The effects of *Melissa officinalis* (lemon balm) pretreatment on the resistance of the heart to myocardial injury. *Pharm Biol* 2016;54:1005-1013. <https://doi.org/10.3109/13880209.2015.1091845>
22. Joukar S, Zarisfi Z, Sepehri G, Bashiri A. Efficacy of *Melissa officinalis* in suppressing ventricular arrhythmias following ischemia-reperfusion of the heart: a comparison with amiodarone. *Med Princ Pract* 2014;23:340-345. <https://doi.org/10.1159/000363452>
23. Stojanović NM, Mladenović MZ, Maslovarić A, Stojiljković NI, Randjelović PJ, Radulović NS. Lemon balm (*Melissa officinalis* L.) essential oil and citronellal modulate anxiety-related symptoms - In vitro and in vivo studies. *J Ethnopharmacol* 2022;284:114788. <https://doi.org/10.1016/j.jep.2021.114788>
24. Gazola R, Machado D, Ruggiero C, Singi G, Alexandre MM. *Lippia alba*, *Melissa officinalis* and *Cymbopogon citratus*: effects of the aqueous extracts on the isolated hearts of rats. *Pharmacol Res* 2004;50:477-480. <https://doi.org/10.1016/j.phrs.2004.01.012>
25. Joukar S, Asadipour H. Evaluation of *Melissa officinalis* (Lemon Balm) effects on heart electrical system. *Res Cardiovasc Med* 2015;4:e27013. <https://doi.org/10.4103/2251-9572.218779>
26. Xu Z-y, Xu Y, Xie X-f, Tian Y, Sui J-h, Sun Y, Lin D-s, Gao X, Peng C, Fan Y-j. Anti-platelet aggregation of *Panax notoginseng* triol saponins by regulating GP1BA for ischemic stroke therapy. *Chin Med* 2021;16:12. <https://doi.org/10.1186/s13020-021-00424-3>

27. Wu Y-H, Wu Y-R, Li B, Yan Z-Y. Cryptotanshinone: A review of its pharmacology activities and molecular mechanisms. *Fitoterapia* 2020;145:104633. <https://doi.org/10.1016/j.fitote.2020.104633>
28. Yang X, Xiong X, Wang H, Wang J. Protective effects of Panax notoginseng saponins on cardiovascular diseases: a comprehensive overview of experimental studies. *Evid Based Complement Alternat Med* 2014;2014:204840. <https://doi.org/10.1155/2014/204840>
29. Maggi MA, Bisti S, Picco C. Saffron: Chemical composition and neuroprotective activity. *Molecules* 2020;25:5618. <https://doi.org/10.3390/molecules25235618>
30. Joukar S, Ghasemipour-Afshar E, Sheibani M, Naghsh N, Bashiri A. Protective effects of saffron (*Crocus sativus*) against lethal ventricular arrhythmias induced by heart reperfusion in rat: a potential anti-arrhythmic agent. *Pharm Biol* 2013;51:836-843. <https://doi.org/10.3109/13880209.2013.767362>
31. Joukar S, Najafipour H, Khaksari M, Sepehri G, Shahrokh N, Dabiri S, Gholamhoseinian A, Hasanzadeh S. The effect of saffron consumption on biochemical and histopathological heart indices of rats with myocardial infarction. *Cardiovasc Toxicol* 2010;10:66-71. <https://doi.org/10.1007/s12012-010-9063-1>
32. Jin W, Xue Y, Liang Y, Zhang Y, Zhang J, Chu X, Wang H, Guan S. Inhibitory effects of four active components in saffron on human ether-a-go-go-related gene (hERG) K<sup>+</sup> currents. *Gen Physiol Biophys* 2020;39:491-498. [https://doi.org/10.4149/gpb\\_2020025](https://doi.org/10.4149/gpb_2020025)
33. Zhao Z, Zheng B, Li J, Wei Z, Chu S, Han X, Chu L, Wang H, Chu X. Influence of crocetin, a natural carotenoid dicarboxylic acid in saffron, on L-type Ca<sup>2+</sup> current, intracellular Ca<sup>2+</sup> handling and contraction of isolated rat cardiomyocytes. *Biol Pharm Bull* 2020;43:1367-1374. <https://doi.org/10.1248/bpb.b20-00298>
34. Li Puma S, Landini L, Macedo Jr SJ, Seravalli V, Marone IM, Coppi E, Patacchini R, Geppetti P, Materazzi S, Nassini R. TRPA1 mediates the antinociceptive properties of the constituent of *Crocus sativus* L., safranal. *J Cell Mol Med* 2019;23:1976-1986. <https://doi.org/10.1111/jemm.14099>
35. Al-Saigh NN, Abdalla S. Safranal induces vasorelaxation by inhibiting Ca<sup>2+</sup> influx and Na<sup>+</sup>/Ca<sup>2+</sup> exchanger in isolated rat aortic rings. *Molecules* 2022;27:4228. <https://doi.org/10.3390/molecules27134228>
36. Beik A, Joukar S, Najafipour H. A review on plants and herbal components with antiarrhythmic activities and their interaction with current cardiac drugs. *J Tradit Complement Med* 2020;10:275-287. <https://doi.org/10.1016/j.jtcme.2020.03.002>
37. Pharmacopoeia of the People's Republic of China. The State Pharmacopoeia Commission of PR China. Beijing: China Medical Science 2010;3.
38. Wang X, Morris-Natschke SL, Lee KH. New developments in the chemistry and biology of the bioactive constituents of Tanshen. *Med Res Rev* 2007;27:133-148. <https://doi.org/10.1002/med.20077>
39. Tsai C-C, Huang S-C, Liu J-K, Wang H-C, Tsai T-R, Tsai P-J, Liu C-W, Chang L-C. *Salvia miltiorrhiza* causes tonic contraction in rat ileum through Ca<sup>2+</sup>-calmodulin pathway. *J Ethnopharmacol* 2012;142:694-699. <https://doi.org/10.1016/j.jep.2012.05.041>
40. Jia C, Han S, Wei L, Dang X, Niu Q, Chen M, Cao B, Liu Y, Jiao H. Protective effect of compound Danshen (*Salvia miltiorrhiza*) dripping pills alone and in combination with carbamazepine on kainic acid-induced temporal lobe epilepsy and cognitive impairment in rats. *Pharm Biol* 2018;56:217-224. <https://doi.org/10.1080/13880209.2018.1432665>
41. Fuloria S, Mehta J, Chandel A, Sekar M, Rani NNIM, Begum MY, Subramaniyan V, Chidambaram K, Thangavelu L, Nordin R. A comprehensive review on the therapeutic potential of *Curcuma longa* Linn. in relation to its major active constituent curcumin. *Front Pharmacol* 2022;13:820806. <https://doi.org/10.3389/fphar.2022.820806>
42. Song L, Zhang Z-F, Hu L-K, Zhang P-H, Cao Z-Z, Liu Z-P, Zhang P-P, Ma J-H. Curcumin, a multi-ion channel blocker that preferentially blocks late Na<sup>+</sup> current and prevents I/R-induced arrhythmias. *Front Physiol* 2020;11:978. <https://doi.org/10.3389/fphys.2020.00978>
43. Choo BK, Shaikh MF. Mechanism of *Curcuma longa* and its neuroactive components for the management of epileptic seizures: A systematic review. *Curr Neuropharmacol* 2021;19:1496-1518. <https://doi.org/10.2174/1570159X19666210517120413>

44. Kumar V, Prakash C, Singh R, Sharma D. Curcumin's antiepileptic effect, and alterations in Nav1.1 and Nav1.6 expression in iron-induced epilepsy. *Epilepsy Res* 2019;150:7-16. <https://doi.org/10.1016/j.eplepsyres.2018.12.007>
45. Aguiar DD, Gonzaga ACR, Teófilo ALH, Miranda FA, de Castro Perez A, Duarte IDG, Romero TRL. Curcumin induces peripheral antinociception by opioidergic and cannabinoidergic mechanism: Pharmacological evidence. *Life Sci* 2022;293:120279. <https://doi.org/10.1016/j.lfs.2021.120279>
46. Lin TY, Lu CW, Wang C-C, Wang Y-C, Wang S-J. Curcumin inhibits glutamate release in nerve terminals from rat prefrontal cortex: possible relevance to its antidepressant mechanism. *Prog Neuropsychopharmacol Biol Psychiatry* 2011;35:1785-1793. <https://doi.org/10.1016/j.pnpbp.2011.06.012>
47. Shin DH, Seo EY, Pang B, Nam JH, Kim HS, Kim WK, Kim SJ. Inhibition of Ca<sup>2+</sup>-release-activated Ca<sup>2+</sup> channel (CRAC) and K<sup>+</sup> channels by curcumin in Jurkat-T cells. *J Pharmacol Sci* 2011;115:144-154. <https://doi.org/10.1254/jphs.10209FP>
48. Son YK, Choi I-W, Park WS. The inhibitory effect of curcumin on voltage-dependent K<sup>+</sup> channels in rabbit coronary arterial smooth muscle cells. *Biochem Biophys Res Commun* 2013;430:307-312. <https://doi.org/10.1016/j.bbrc.2012.10.132>
49. Meng B, Shen L-l, Shi X-t, Gong Y-s, Fan X-f, Li J, Cao H. Effects of curcumin on TTX-R sodium currents of dorsal root ganglion neurons in type 2 diabetic rats with diabetic neuropathic pain. *Neurosci Lett* 2015;605:59-64. <https://doi.org/10.1016/j.neulet.2015.08.011>
50. Díaz-Triste NE, González-García MP, Jiménez-Andrade JM, Castañeda-Hernández G, Chávez-Piña AE. Pharmacological evidence for the participation of NO-cGMP-KATP pathway in the gastric protective effect of curcumin against indomethacin-induced gastric injury in the rat. *Eur J Pharmacol* 2014;730:102-106. <https://doi.org/10.1016/j.ejphar.2014.02.030>
51. Nalli M, Ortar G, Moriello AS, Di Marzo V, De Petrocellis L. Effects of curcumin and curcumin analogues on TRP channels. *Fitoterapia* 2017;122:126-131. <https://doi.org/10.1016/j.fitote.2017.09.007>
52. Adamcová M, Štěrba M, Šimůnek T, Potáčková A, Popelová O, Geršl V. Myocardial regulatory proteins and heart failure. *Eur J Heart Fail* 2006;8:333-342. <https://doi.org/10.1016/j.ejheart.2005.09.007>
53. Song Q, Chu X, Zhang X, Bao Y, Zhang Y, Guo H, Liu Y, Liu H, Zhang J, Zhang Y. Mechanisms underlying the cardioprotective effect of Salvianic acid A against isoproterenol-induced myocardial ischemia injury in rats: possible involvement of L-type calcium channels and myocardial contractility. *J Ethnopharmacol* 2016;189:157-164. <https://doi.org/10.1016/j.jep.2016.05.038>
54. Zhang S, DiMango E, Zhu Y, Saroya TK, Emala CW, Sang S. Pharmacokinetics of gingerols, shogaols, and their metabolites in asthma patients. *J Agric Food Chem* 2022;70:9674-9683. <https://doi.org/10.1021/acs.jafc.2c03150>
55. Han X, Zhang Y, Liang Y, Zhang J, Li M, Zhao Z, Zhang X, Xue Y, Zhang Y, Xiao J. 6-Gingerol, an active pungent component of ginger, inhibits L-type Ca<sup>2+</sup> current, contractility, and Ca<sup>2+</sup> transients in isolated rat ventricular myocytes. *Food Sci Nutr* 2019;7:1344-1352. <https://doi.org/10.1002/fsn3.968>
56. Kim S, Cheon C, Kim B, Kim W. The effect of ginger and its sub-components on pain. *Plants* 2022;11:2296. <https://doi.org/10.3390/plants11172296>
57. Gawel K, Kukula-Koch W, Banono NS, Nieoczym D, Targowska-Duda KM, Czernicka L, Parada-Turska J, Esguerra CV. 6-Gingerol, a major constituent of *Zingiber officinale* rhizoma, exerts anticonvulsant activity in the pentylenetetrazole-induced seizure model in larval zebrafish. *Int J Mol Sci* 2021;22:7745. <https://doi.org/10.3390/ijms22147745>
58. Mukkavilli R, Yang C, Singh Tanwar R, Ghareeb A, Luthra L, Aneja R. Absorption, metabolic stability, and pharmacokinetics of ginger phytochemicals. *Molecules* 2017;22:553. <https://doi.org/10.3390/molecules22040553>
59. Xue Y, Zhang M, Zheng B, Zhang Y, Chu X, Liu Y, Li Z, Han X, Chu L. [8]-Gingerol exerts anti-myocardial ischemic effects in rats via modulation of the MAPK signaling pathway and L-type Ca<sup>2+</sup> channels. *Pharmacol Res Perspect* 2021;9:e00852. <https://doi.org/10.1002/prp2.852>
60. Li G-R, Wang H-B, Qin G-W, Jin M-W, Tang Q, Sun H-Y, Du X-L, Deng X-L, Zhang X-H, Chen J-B. Acacetin, a natural flavone, selectively inhibits human atrial repolarization potassium currents and prevents atrial fibrillation in dogs. *Circulation* 2008;117:2449-2457. <https://doi.org/10.1161/CIRCULATIONAHA.108.769554>

61. Liu H, Wang Y-J, Yang L, Zhou M, Jin M-W, Xiao G-S, Wang Y, Sun H-Y, Li G-R. Synthesis of a highly water-soluble acacetin prodrug for treating experimental atrial fibrillation in beagle dogs. *Sci Rep* 2016;6:25743. <https://doi.org/10.1038/srep25743>
62. Gao Y, Fan H, Nie A, Yang K, Xing H, Gao Z, Yang L, Wang Z, Zhang L. Aconitine: A review of its pharmacokinetics, pharmacology, toxicology and detoxification. *J Ethnopharmacol* 2022;293:115270. <https://doi.org/10.1016/j.jep.2022.115270>
63. Chan TY. Aconite poisoning. *Clin Toxicol* 2009;47:279-285. <https://doi.org/10.1080/15563650902904407>
64. Binayi F, Joukar S, Najafipour H, Karimi A, Abdollahi F, Masumi Y. The effects of nandrolone decanoate along with prolonged low-intensity exercise on susceptibility to ventricular arrhythmias. *Cardiovasc Toxicol* 2016;16:23-33. <https://doi.org/10.1007/s12012-015-9313-3>
65. Chen Y, Huang Y, Bai J, Liu C, Ma S, Li J, Lu X, Fu Z, Fang L, Li Y. Effects of allicin on late sodium current caused by  $\Delta$ KPQ-SCN5A mutation in HEK293 cells. *Front Physiol* 2021;12:636485. <https://doi.org/10.3389/fphys.2021.636485>
66. Lu C-W, Hung C-F, Lin T-Y, Hsieh TY, Wang SJ. Allicin inhibits glutamate release from rat cerebral cortex nerve terminals through suppressing  $\text{Ca}^{2+}$  influx and protein kinase C activity. *J Med Food* 2019;22:696-702. <https://doi.org/10.1089/jmf.2018.4337>
67. Cao H, Huang C, Wang X. Allicin inhibits transient outward potassium currents in mouse ventricular myocytes. *Exp Ther Med* 2016;11:1896-1900. <https://doi.org/10.3892/etm.2016.3116>
68. Hu Z, Li J, Liu Q, Manville RW, Abbott GW. The plant-derived alkaloid aloperine prevents ischemia/reperfusion injury-induced sudden cardiac death. *FASEB J* 2023;37:e22999. <https://doi.org/10.1096/fj.202300253R>
69. Lamothe SM, Guo J, Li W, Yang T, Zhang S. The human ether-a-go-go-related gene (hERG) potassium channel represents an unusual target for protease-mediated damage. *J Biol Chem* 2016;291:20387-20401. <https://doi.org/10.1074/jbc.M116.743138>
70. Li M-T, Du Y-Y, Zhong F, Wang J-R, Gu Y-W, Zhang Y, Huang X-T, Deng Y-Z, Xu Z-X. Inhibitory effects of aloperine on voltage-gated  $\text{Na}^+$  channels in rat ventricular myocytes. *Naunyn-Schmiedeberg's Arch Pharmacol* 2021;394:1579-1588. <https://doi.org/10.1007/s00210-021-02076-4>
71. Song H, Ao Z, Song Y, Li X, Xu X, Cheng C, Shi M, Liu L, Wu J, Liu Y. Effects of artemisinin on peak sodium current in ventricular myocytes. *Cardiology Cardiovasc Med* 2020;4:111-117. <https://doi.org/10.26502/fccm.92920109>
72. Uebel T, Hermes L, Hauptenthal S, Müller L, Esselen M.  $\alpha$ -Asarone,  $\beta$ -asarone, and  $\gamma$ -asarone: Current status of toxicological evaluation. *J Appl Toxicol* 2021;41:1166-1179. <https://doi.org/10.1002/jat.4112>
73. Yuan X, Li Z, Wang X-T, Li X-Y, Hua H, Li X-C, Tang R-X, Liu X-M. Roles and mechanisms of traditional Chinese medicine and its active ingredients in treating epilepsy. *Zhongguo Zhong yao za zhi = Zhongguo Zhongyao Zazhi = China Journal of Chinese Materia Medica* 2019;44:9-18.
74. Zhao M, Zhao J, He G, Sun X, Huang X, Hao L. Effects of astragaloside IV on action potentials and ionic currents in guinea-pig ventricular myocytes. *Biol Pharm Bull* 2013;36:515-521. <https://doi.org/10.1248/bpb.b12-00655>
75. Li J, Yang Y, Wang H, Ma D, Wang H, Chu L, Zhang Y, Gao Y. Baicalein ameliorates myocardial ischemia through reduction of oxidative stress, inflammation and apoptosis via TLR4/MyD88/MAPKS/NF- $\kappa$ B pathway and regulation of  $\text{Ca}^{2+}$  homeostasis by L-type  $\text{Ca}^{2+}$  channels. *Front Pharmacol* 2022;13:842723. <https://doi.org/10.3389/fphar.2022.842723>
76. Patel DK, Patel K, Tahilyani V. Barbaloin: a concise report of its pharmacological and analytical aspects. *Asian Pac J Trop Biomed*. 2012 Oct;2(10):835-8. [https://doi.org/10.1016/S2221-1691\(12\)60239-1](https://doi.org/10.1016/S2221-1691(12)60239-1)
77. Cao Z-Z, Tian Y-J, Hao J, Zhang P-H, Liu Z-P, Jiang W-Z, Zeng M-L, Zhang P-P, Ma J-H. Barbaloin inhibits ventricular arrhythmias in rabbits by modulating voltage-gated ion channels. *Acta Pharmacol Sin* 2018;39:357-370. <https://doi.org/10.1038/aps.2017.93>
78. Zhang X, Gao Y, Zhou Y, Liu Z, Liu R. Pharmacological mechanism of natural drugs and their active ingredients in the treatment of arrhythmia via calcium channel regulation. *Biomed Pharmacother* 2023;160:114413. <https://doi.org/10.1016/j.biopha.2023.114413>

79. Mahdavi N, Joukar S, Najafipour H, Asadi-Shekaari M. The promising effect of barberry (Zereshk) extract against experimental pulmonary microvascular remodeling and hypertension: A comparison with sildenafil. *Pharm Biol* 2016;54:509-515. <https://doi.org/10.3109/13880209.2015.1050676>
80. Joukar S, Mahdavi N. Alterations of blood pressure and ECG following two-week consumption of *Berberis integerrima* fruit extract. *Int Schol Res Notices* 2014;2014:209683. <https://doi.org/10.1155/2014/209683>
81. Hu Y, Zhang P, Wang X. Berberine exerts neuroprotective effects in Alzheimer's disease by switching microglia M1/M2 polarization through PI3K-AKT signaling. *Physiol Res* 2025;74:129-140. <https://doi.org/10.33549/physiolres.935410>
82. Cai Y, Xin Q, Lu J, Miao Y, Lin Q, Cong W, Chen K. A new therapeutic candidate for cardiovascular diseases: berberine. *Front Pharmacol* 2021;12:631100. <https://doi.org/10.3389/fphar.2021.631100>
83. Zhang B, Wang L, Ji X, Zhang S, Sik A, Liu K, Jin M. Anti-inflammation associated protective mechanism of berberine and its derivatives on attenuating pentylenetetrazole-induced seizures in zebrafish. *J Neuroimmune Pharmacol* 2020;15:309-325. <https://doi.org/10.1007/s11481-019-09902-w>
84. Zhou Z-W, Zheng H-C, Zhao L-F, Li W, Hou J-W, Yu Y, Miao P-Z, Zhu J-M. Effect of berberine on acetylcholine-induced atrial fibrillation in rabbit. *Am J Transl Res* 2015;7:1450.
85. Mirhadi E, Rezaee M, Malaekheh-Nikouei B. Nano strategies for berberine delivery, a natural alkaloid of *Berberis*. *Biomed Pharmacother* 2018;104:465-473. <https://doi.org/10.1016/j.biopha.2018.05.067>
86. Meng J, Qiu S, Zhang L, You M, Xing H, Zhu J. Berberine alleviate cisplatin-induced peripheral neuropathy by modulating inflammation signal via TRPV1. *Front Pharmacol* 2022;12:774795. <https://doi.org/10.3389/fphar.2021.774795>
87. Almanaitytė M, Jurevičius J, Mačianskienė R. Effect of carvacrol, TRP channels modulator, on cardiac electrical activity. *Biomed Res Int* 2020;2020:6456805. <https://doi.org/10.1155/2020/6456805>
88. Gültekin B, Çetinkaya Karabekir S, Ayan IÇ, Savaş HB, Cüce G, Kalkan SS. Effect of carvacrol on diabetes-induced oxidative stress, fibrosis and apoptosis in testicular tissues of adult rats. *Physiol Res* 2025;74:459-469. <https://doi.org/10.33549/physiolres.935573>
89. Jin X, Jiang Y, Xue G, Yuan Y, Zhu H, Zhan L, Zhuang Y, Huang Q, Shi L, Zhao Y. Increase of late sodium current contributes to enhanced susceptibility to atrial fibrillation in diabetic mice. *Eur J Pharmacol* 2019;857:172444. <https://doi.org/10.1016/j.ejphar.2019.172444>
90. Xia J, Guo D, Zhang Y, Zhou Z, Zeng F, Hu C. Inhibitory effects of dauricine on potassium currents in guinea pig ventricular myocytes. *Acta Pharmacol Sin* 2000;21:60-64.
91. Olgar Y, Ozturk N, Usta C, Puddu PE, Ozdemir S. Ellagic acid reduces L-type Ca<sup>2+</sup> current and contractility through modulation of NO-GC-cGMP pathways in rat ventricular myocytes. *J Cardiovasc Pharmacol* 2014;64:567-573. <https://doi.org/10.1097/FJC.0000000000000153>
92. Luo Y, Meng X, Zhou P, Lu S, Qin M, Xu X, Sun G, Sun X. Elatoside C protects against ox-LDL-induced HUVECs injury by FoxO1-mediated autophagy induction. *Biochim Biophys Acta Mol Basis Dis* 2017;1863:1654-1665. <https://doi.org/10.1016/j.bbadis.2017.01.017>
93. Wang M, Sun G-B, Zhang J-Y, Luo Y, Yu Y-L, Xu X-D, Meng X-B, Lin W-B, Sun X-B. Elatoside C protects the heart from ischaemia/reperfusion injury through the modulation of oxidative stress and intracellular Ca<sup>2+</sup> homeostasis. *Int J Cardiol* 2015;185:167-176. <https://doi.org/10.1016/j.ijcard.2015.03.140>
94. Guo Y, Zhang R, Li W. Emodin in cardiovascular disease: The role and therapeutic potential. *Front Pharmacol* 2022;13:1070567. <https://doi.org/10.3389/fphar.2022.1070567>
95. Zhang C, Xiao M, Cao N, Zhang L, He Q, Wang J, Wang R, Wang L, Zhao L, Si J. Emodin activates BK channel in vascular smooth muscle cells and relaxes the interlobar renal artery of rat. *Biomed Pharmacother* 2022;153:113452. <https://doi.org/10.1016/j.biopha.2022.113452>
96. Joukar S, Bashiri H, Dabiri S, Ghotbi P, Sarveazad A, Divsalar K, Joukar F, Abbaszadeh M. Cardiovascular effects of black tea and nicotine alone or in combination against experimental induced heart injury. *J Physiol Biochem* 2012;68:271-279. <https://doi.org/10.1007/s13105-011-0141-z>
97. Joukar S, Shahouzehi B, Najafipour H, Gholamhoseinian A, Joukar F. Ameliorative effect of black tea on nicotine induced cardiovascular pathogenesis in rat. *EXCLI J* 2012;11:309-317.

98. Tran HH-V, Mansoor M, Butt SRR, Satnarine T, Ratna P, Sarker A, Ramesh AS, Munoz C, Jamil D, Mohammed L. Impact of green tea consumption on the prevalence of cardiovascular outcomes: A systematic review. *Cureus* 2023;15:e49775. <https://doi.org/10.7759/cureus.49775>
99. Wu AZ-Y, Loh S-H, Cheng T-H, Lu H-H, Lin C-I. Antiarrhythmic effects of (-)-epicatechin-3-gallate, a novel sodium channel agonist in cultured neonatal rat ventricular myocytes. *Biochem Pharmacol* 2013;85:69-80. <https://doi.org/10.1016/j.bcp.2012.10.003>
100. Amarouch M-Y, Kurt H, Delemotte L, Abriel H. Biophysical characterization of epigallocatechin-3-gallate effect on the cardiac sodium channel Nav1.5. *Molecules* 2020;25:902. <https://doi.org/10.3390/molecules25040902>
101. Chang J-H, Chang S-L, Hong P-D, Chen P-N, Hsu C-H, Lu Y-Y, Chen Y-C. Epigallocatechin-3-gallate modulates arrhythmogenic activity and calcium homeostasis of left atrium. *Int J Cardiol* 2017;236:174-180. <https://doi.org/10.1016/j.ijcard.2017.01.090>
102. Peixoto-Neves D, Soni H, Adebisi A. CGRPergic nerve TRPA1 channels contribute to epigallocatechin gallate-induced neurogenic vasodilation. *ACS Chem Neurosci* 2018;10:216-220. <https://doi.org/10.1021/acschemneuro.8b00493>
103. Takahashi S, Kurogi M, Saitoh O. The diversity in sensitivity of TRPA1 and TRPV1 of various animals to polyphenols. *Biomed Res* 2021;42:43-51. <https://doi.org/10.2220/biomedres.42.43>
104. Moreira-Lobo DC, Linhares-Siqueira ED, Cruz GM, Cruz JS, Carvalho-de-Souza JL, Lahlou S, Coelho-de-Souza AN, Barbosa R, Magalhães PJ, Leal-Cardoso JH. Eugenol modifies the excitability of rat sciatic nerve and superior cervical ganglion neurons. *Neurosci Lett* 2010;472:220-224. <https://doi.org/10.1016/j.neulet.2010.02.009>
105. Chung G, Im S, Kim Y, Jung S, Rhyu M-R, Oh S. Activation of transient receptor potential ankyrin 1 by eugenol. *Neuroscience* 2014;261:153-160. <https://doi.org/10.1016/j.neuroscience.2013.12.047>
106. Ye H, Lin Q, Mei Q, Liu Q, Cao S. Study on mechanism of transdermal administration of eugenol for pain treatment by network pharmacology and molecular docking technology. *Heliyon* 2024;10:e2972. <https://doi.org/10.1016/j.heliyon.2024.e29722>
107. Bai B, Lu N, Zhang W, Lin J, Zhao T, Zhou S, Khasanova E, Zhang L. Inhibitory effects of genistein on vascular smooth muscle cell proliferation induced by Ox-LDL: role of BKCa channels. *Anal Cell Pathol* 2020;2020:8895449. <https://doi.org/10.1155/2020/8895449>
108. Wong CO, Huang Y, Yao X. Genistein potentiates activity of the cation channel TRPC5 independently of tyrosine kinases. *Br J Pharmacol* 2010;159:1486-1496. <https://doi.org/10.1111/j.1476-5381.2010.00636.x>
109. Lv X-F, Wen R-Q, Liu K, Zhao X-K, Pan C-L, Gao X, Wu X, Zhi X-D, Ren C-Z, Chen Q-L. Role and molecular mechanism of traditional Chinese medicine in preventing cardiotoxicity associated with chemoradiotherapy. *Front Cardiovasc Med* 2022;9:1047700. <https://doi.org/10.3389/fcvm.2022.1047700>
110. Wang Z, Zu T, Huang X, Jiang X, Jia G, Xu J, Cui Z, Zhu F, Zhang J, Li J. Comprehensive investigation of the content and the origin of matrine-type alkaloids in Chinese honeys. *Food Chem* 2023;402:134254. <https://doi.org/10.1016/j.foodchem.2022.134254>
111. Bai CX, Takahashi K, Masumiya H, Sawanobori T, Furukawa T. Nitric oxide-dependent modulation of the delayed rectifier K<sup>+</sup> current and the L-type Ca<sup>2+</sup> current by ginsenoside Re, an ingredient of Panax ginseng, in guinea-pig cardiomyocytes. *Br J Pharmacol* 2004;142:567-575. <https://doi.org/10.1038/sj.bjp.0705814>
112. Liu Z, Song L, Zhang P, Cao Z, Hao J, Tian Y, Luo A, Zhang P, Ma J. Ginsenoside Rb1 exerts antiarrhythmic effects by inhibiting I<sub>Na</sub> and I<sub>CaL</sub> in rabbit ventricular myocytes. *Sci Rep* 2019;9:20425. <https://doi.org/10.1038/s41598-019-57010-9>
113. Kim S, Rhim H. Ginsenosides inhibit NMDA receptor-mediated epileptic discharges in cultured hippocampal neurons. *Arch Pharm Res* 2004;27:524-530. <https://doi.org/10.1007/BF02980126>
114. Singh K, Zaw AM, Sekar R, Palak A, Allam AA, Ajarem J, Chow BK. Glycyrrhizic acid reduces heart rate and blood pressure by a dual mechanism. *Molecules* 2016;21:1291. <https://doi.org/10.3390/molecules21101291>
115. Yang J, Ye K, Zhang R, Fan X, Xiong R, Zhang S, Liu Q, Lin M, Wang B, Tan X. The characteristics and molecular targets of antiarrhythmic natural products. *Biomed Pharmacother* 2023;168:115762. <https://doi.org/10.1016/j.biopha.2023.115762>

116. Narayana Moorthy NSH, J Ramos M, A Fernandes P. Human ether-a-go-go-related gene channel blockers and its structural analysis for drug design. *Curr Drug Targets* 2013;14:102-113. <https://doi.org/10.2174/138945013804806460>, <https://doi.org/10.2174/1389450111314010011>
117. Xiong F, Liu K, Liu S, Chen J, Liu J, Wang H, Gu N. Safety, heart specificity, and therapeutic effect evaluation of Guanfu base A-loaded solid nanolipids in treating arrhythmia. *Drug Deliv Transl Res* 2018;8:1471-1482. <https://doi.org/10.1007/s13346-018-0542-4>
118. Sun J, Peng Y, Wu H, Zhang X, Zhong Y, Xiao Y, Zhang F, Qi H, Shang L, Zhu J. Guanfu base A, an antiarrhythmic alkaloid of *Aconitum coreanum*, is a CYP2D6 inhibitor of human, monkey, and dog isoforms. *Drug Metab Dispos* 2015;43:713-724. <https://doi.org/10.1124/dmd.114.060905>
119. Wang H, Wang H-F, Zhang H, Wang C, Chen Y-F, Ma R, Xiang J-Z, Du X-L, Tang Q. Inhibitory effects of hesperetin on Nav1.5 channels stably expressed in HEK 293 cells and on the voltage-gated cardiac sodium current in human atrial myocytes. *Acta Pharmacol Sin* 2016;37:1563-1573. <https://doi.org/10.1038/aps.2016.97>
120. Chang CY, Lin TY, Lu CW, Huang SK, Wang YC, Chou SSP, Wang SJ. Hesperidin inhibits glutamate release and exerts neuroprotection against excitotoxicity induced by kainic acid in the hippocampus of rats. *Neurotoxicology* 2015;50:157-169. <https://doi.org/10.1016/j.neuro.2015.08.014>
121. Xiong Y-J, Chu H-W, Lin Y, Han F, Li Y-C, Wang A-G, Wang F-J, Chen D-P, Wang J-Y. Hesperidin alleviates rat postoperative ileus through anti-inflammation and stimulation of Ca<sup>2+</sup>-dependent myosin phosphorylation. *Acta Pharmacol Sin* 2016;37:1091-1100. <https://doi.org/10.1038/aps.2016.56>
122. Ahlatcı A, Yıldızhan K, Keleş ÖF, Bayir MH, Çınar R. The effect of hesperidin on trigeminal nerve damage in an NTG-induced migraine model: the role of the TRPV1 channel. *Mol Biol Rep* 2026;53:25. <https://doi.org/10.1007/s11033-025-11194-8>
123. Cortez GB, Bertozzi MM, Dionisio AM, Piva M, Morelli NR, Carvalho TT, Casagrande R, Verri WA, Borghi SM. Role of TRPV1<sup>+</sup> and TRPA1<sup>+</sup> nociceptive neurons in delayed-onset muscle soreness: inhibition by hesperidin methyl chalcone. *Inflammopharmacology* 2025;33:2815-2832. <https://doi.org/10.1007/s10787-025-01762-6>
124. Bayir MH, Yıldızhan K, Altındağ F. Effect of hesperidin on sciatic nerve damage in STZ-induced diabetic neuropathy: modulation of TRPM2 channel. *Neurotox Res* 2023;41:638-647. <https://doi.org/10.1007/s12640-023-00657-0>
125. Jiang W, Zeng M, Cao Z, Liu Z, Hao J, Zhang P, Tian Y, Zhang P, Ma J. Icariin, a novel blocker of sodium and calcium channels, eliminates early and delayed afterdepolarizations, as well as triggered activity, in rabbit cardiomyocytes. *Front Physiol* 2017;8:342. <https://doi.org/10.3389/fphys.2017.00342>
126. Kumar T, Sharma M, Rana A, Lingaraju MC, Parida S, Kumar D, Singh TU. Effect of different potassium channel blockers on kaempferol-induced relaxation in the isolated coronary artery of goat. *J Pharmacognosy Phytochem* 2019;8:2944-2947.
127. Yang Y, Qi J, Zhang M, Chen P, Liu Y, Sun X, Chu L. The cardioprotective effects and mechanisms of naringenin in myocardial ischemia based on network pharmacology and experiment verification. *Front Pharmacol* 2022;13:954555. <https://doi.org/10.3389/fphar.2022.954555>
128. Jara-Oseguera A, Simon SA, Rosenbaum T. TRPV1: on the road to pain relief. *Curr Mol Pharmacol* 2008;1:255-269. <https://doi.org/10.2174/1874467210801030255>
129. Cui M, Gosu V, Basith S, Hong S, Choi S. Polymodal transient receptor potential vanilloid type 1 nociceptor: structure, modulators, and therapeutic applications. *Adv Protein Chem Struct Biol* 2016;104:81-125. <https://doi.org/10.1016/bs.apcsb.2015.11.005>
130. Zarei MM, Abdolmaleki Z, Shahidi S. Bioflavonoid exerts analgesic and anti-inflammatory effects via transient receptor potential 1 channel in a rat model. *Arq Neuropsiquiatr* 2022;80:900-907. <https://doi.org/10.1055/s-0042-1755321>
131. Slepíčka J, Paleček J. Glial Activation enhances spinal TRPV1 receptor sensitivity in a paclitaxel model of neuropathic pain. *Physiol Res* 2025;74:677-699. <https://doi.org/10.33549/physiolres.935599>
132. Gong C-L, Wong K-L, Cheng K-S, Kuo C-S, Chao C-C, Tsai M-F, Leung Y-M. Inhibitory effects of magnolol on voltage-gated Na<sup>+</sup> and K<sup>+</sup> channels of NG108-15 cells. *Eur J Pharmacol* 2012;682:73-78. <https://doi.org/10.1016/j.ejphar.2012.02.013>

133. Deng Y, Han X, Tang S, Xiao W, Tan Z, Zhou C, Wang M, Kang J. Magnolol and honokiol regulate the calcium-activated potassium channels signaling pathway in enterotoxigenic *Escherichia coli*-induced diarrhea mice. *Eur J Pharmacol* 2015;755:66-73. <https://doi.org/10.1016/j.ejphar.2015.03.002>
134. Zhang M, Zang K-H, Luo J-L, Leung F-P, Huang Y, Lin C-Y, Yang Z-J, Lu A-P, Tang X-D, Xu H-X. Magnolol inhibits colonic motility through down-regulation of voltage-sensitive L-type  $Ca^{2+}$  channels of colonic smooth muscle cells in rats. *Phytomedicine* 2013;20:1272-1279. <https://doi.org/10.1016/j.phymed.2013.07.008>
135. Niu L, Wang J, Shen F, Gao J, Jiang M, Bai G. Magnolol and honokiol target TRPC4 to regulate extracellular calcium influx and relax intestinal smooth muscle. *J Ethnopharmacol* 2022;290:115105. <https://doi.org/10.1016/j.jep.2022.115105>
136. Woo C-Y, Baek JY, Kim A-R, Hong CH, Yoon JE, Kim HS, Yoo HJ, Park T-S, Kc R, Lee K-U. Inhibition of ceramide accumulation in podocytes by myriocin prevents diabetic nephropathy. *Diabetes Metab J* 2020;44:581-591. <https://doi.org/10.4093/dmj.2019.0063>
137. Saponara S, Fusi F, Iovinelli D, Ahmed A, Trezza A, Spiga O, Sgaragli G, Valoti M. Flavonoids and hERG channels: Friends or foes? *Eur J Pharmacol* 2021;899:174030. <https://doi.org/10.1016/j.ejphar.2021.174030>
138. Wilson SL, Dempsey CE, Hancox JC, Marrison NV. Identification of a proton sensor that regulates conductance and open time of single hERG channels. *Sci Rep* 2019;9:19825. <https://doi.org/10.1038/s41598-019-56081-y>
139. Zhou J, Ma W, Wang X, Liu H, Miao Y, Wang J, Du P, Chen Y, Zhang Y, Liu Z. Matrine suppresses reactive oxygen species (ROS)-mediated MKKs/p38-induced inflammation in oxidized low-density lipoprotein (ox-LDL)-stimulated macrophages. *Med Sci Monit* 2019;25:4130-4136. <https://doi.org/10.12659/MSM.917151>
140. Zhou Y, Xu W, Han R, Zhou J, Pan Z, Rong H, Li J, Xu C, Qiao G, Lu Y. Matrine inhibits pacing induced atrial fibrillation by modulating IKM3 and I $Ca$ -L. *Int J Biol Sci* 2012;8:150-158. <https://doi.org/10.7150/ijbs.8.150>
141. Shrestha S, Sunaga H, Hanaoka H, Yamaguchi A, Kuwahara S, Umbarawan Y, Nakajima K, Machida T, Murakami M, Saito A, Tsushima Y, Kurabayashi M, Iso T. Circulating FABP4 is eliminated by the kidney via glomerular filtration followed by megalin-mediated reabsorption. *Sci Rep* 2018;8:16451. <https://doi.org/10.1038/s41598-018-34902-w>
142. Patel DK, Patel K. Therapeutic potential and pharmacological activities of (+)-nantenine in medicine: An aporphine class phytocomponent of *Nandina domestica* Thunberg. *Infect Disord Drug Targets* 2024;24:73-80. <https://doi.org/10.2174/0118715265244269231010090316>
143. Zhu HL, Wan JB, Wang YT, Li BC, Xiang C, He J, Li P. Medicinal compounds with antiepileptic/ anticonvulsant activities. *Epilepsia* 2014;55:3-16. <https://doi.org/10.1111/epi.12463>
144. Alvarez-Collazo J, López-Medina AI, Rodríguez AA, Alvarez JL. Mechanism of the negative inotropic effect of naringin in mouse heart. *J Pharm Pharmacognosy Res* 2014;2:148-157. [https://doi.org/10.56499/jppres14.046\\_2.5.148](https://doi.org/10.56499/jppres14.046_2.5.148)
145. Wicha P, Onsa-Ard A, Chaichompoo W, Suksamrarn A, Tocharus C. Vasorelaxant and antihypertensive effects of neferine in rats: an in vitro and in vivo study. *Planta Med* 2020;86:496-504. <https://doi.org/10.1055/a-1123-7852>
146. Xu T, Singh D, Liu J, Li H, Peng S, Rizzolo LJ, Wang S-B. Neferine is not inducer but blocker for macroautophagic flux targeting on lysosome malfunction. *Biochem Biophys Res Commun* 2018;495:1516-1521. <https://doi.org/10.1016/j.bbrc.2017.11.169>
147. Xu X, Song S, Zhao X, Qu Y, Li D, Shen L, Liu J, Yue S. Osthole alleviates neuropathic pain by suppressing astrocytes activation and associated inflammatory responses via the PKC $\delta$ /TRPV4 signaling pathway. *Int Immunopharmacol* 2025;165:115453. <https://doi.org/10.1016/j.intimp.2025.115453>
148. Torres KV, Pantke S, Rudolf D, Eberhardt MM, Leffler A. The coumarin osthole is a non-electrophilic agonist of TRPA1. *Neurosci Lett* 2022;789:136878. <https://doi.org/10.1016/j.neulet.2022.136878>
149. Ong W-Y, Herr DR, Sun GY, Lin T-N. Anti-inflammatory effects of phytochemical components of *Clinacanthus nutans*. *Molecules* 2022;27:3607. <https://doi.org/10.3390/molecules27113607>
150. Tian F, Tong M, Li Z, Huang W, Jin Y, Cao Q, Zhou X, Tong G. The effects of orientin on proliferation and apoptosis of T24 human bladder carcinoma cells occurs through the inhibition of nuclear factor-kappaB and the hedgehog signaling pathway. *Med Sci Monit* 2019;25:9547-9954. <https://doi.org/10.12659/MSM.919203>

151. Trettel G, Bertocini CRA, Lima-Landman MT. The mechanisms of calcium mobilization by procyanidins, flavonols and flavonoids from *Cecropia glaziovii* Sneth in pulmonary endothelial cell cultures endorse its popular use as vasodilator phytomedicine. *Biomed Pharmacother* 2021;144:112231. <https://doi.org/10.1016/j.biopha.2021.112231>
152. Li F, Zong J, Zhang H, Zhang P, Xu L, Liang K, Yang L, Yong H, Qian W. Orientin reduces myocardial infarction size via eNOS/NO signaling and thus mitigates adverse cardiac remodeling. *Front Pharmacol* 2017;8:926. <https://doi.org/10.3389/fphar.2017.00926>
153. Cao YG, Jing S, Li L, Gao JQ, Shen ZY, Liu Y, Xing Y, Wu ML, Wang Y, Xu CQ, Sun HL. Antiarrhythmic effects and ionic mechanisms of oxymatrine from *Sophora flavescens*. *Phytother Res* 2010;24:1844-1849. <https://doi.org/10.1002/ptr.3206>
154. Wang Y, Yuan J, Yuan X, Wang W, Pei X, Zhao Q, Cao H, Xu M, Liu Z. Observation of antinociceptive effects of oxymatrine and its effect on delayed rectifier K<sup>+</sup> currents (I<sub>k</sub>) in PC12 cells. *Neurochem Res* 2012;37:2143-2149. <https://doi.org/10.1007/s11064-012-0836-8>
155. Shuyong W. Research progress on cardiovascular protective effect and mechanism of puerarin. *Chin J Tradit Chin Med* 2015;40:2278-2284.
156. Scholz EP, Zitron E, Katus HA, Karle CA. Cardiovascular ion channels as a molecular target of flavonoids. *Cardiovasc Ther* 2010;28:e46-e52. <https://doi.org/10.1111/j.1755-5922.2010.00212.x>
157. Zhang G-Q, Hao X-M, Dai D-Z, Fu Y, Zhou P-A, Wu C-H. Puerarin blocks Na<sup>+</sup> current in rat ventricular myocytes. *Acta Pharmacol Sin* 2003;24:1212-1216.
158. Zhou T, Wang Z, Guo M, Zhang K, Geng L, Mao A, Yang Y, Yu F. Puerarin induces mouse mesenteric vasodilation and ameliorates hypertension involving endothelial TRPV4 channels. *Food Funct* 2020;11:10137-10148. <https://doi.org/10.1039/D0FO02356F>
159. Lin Y, Liang R, Xie K, Ma T, Zhang J, Xu T, Wang A, Liu S. Puerarin inhibits cisplatin-induced ototoxicity in mice through regulation of TRPV1-dependent calcium overload. *Biochem Pharmacol* 2024;220:115962. <https://doi.org/10.1016/j.bcp.2023.115962>
160. Wu Y, Chen J, Wang R. Puerarin suppresses TRPV1, calcitonin gene-related peptide and substance P to prevent paclitaxel-induced peripheral neuropathic pain in rats. *Neuroreport* 2019;30:288-294. <https://doi.org/10.1097/WNR.0000000000001199>
161. Zeng X, Feng Q, Zhao F, Sun C, Zhou T, Yang J, Zhan X. Puerarin inhibits TRPM3/miR-204 to promote MC3T3-E1 cells proliferation, differentiation and mineralization. *Phytother Res* 2018;32:996-1003. <https://doi.org/10.1002/ptr.6034>
162. Huang W, Kong L, Cao Y, Yan L. Identification and quantification, metabolism and pharmacokinetics, pharmacological activities, and botanical preparations of protopine: A review. *Molecules* 2021;27:215. <https://doi.org/10.3390/molecules27010215>
163. Song LS, Ren GJ, Chen ZL, Chen ZH, Zhou ZN, Cheng H. Electrophysiological effects of protopine in cardiac myocytes: inhibition of multiple cation channel currents. *Br J Pharmacol* 2000;129:893-900. <https://doi.org/10.1038/sj.bjp.0703132>
164. Zhang Y-M, Zhang Z-Y, Wang R-X. Protective mechanisms of quercetin against myocardial ischemia reperfusion injury. *Front Physiol* 2020;11:956. <https://doi.org/10.3389/fphys.2020.00956>
165. Rajabi S, Najafipour H, Jafarnejad-Farsangi S, Joukar S, Beik A, Askaripour M, Jafari E, Safi Z. Quercetin, perillyl alcohol, and berberine ameliorate right ventricular disorders in experimental pulmonary arterial hypertension: effects on miR-204, miR-27a, fibrotic, apoptotic, and inflammatory factors. *J Cardiovasc Pharmacol* 2021;77:777-786. <https://doi.org/10.1097/FJC.0000000000001015>
166. Patel RV, Mistry BM, Shinde SK, Syed R, Singh V, Shin H-S. Therapeutic potential of quercetin as a cardiovascular agent. *Eur J Med Chem* 2018;155:889-904. <https://doi.org/10.1016/j.ejmech.2018.06.053>
167. Liang Y, Zhang Y, Liu M, Han X, Zhang J, Zhang X, Chu L. Protective effect of quercetin against myocardial ischemia as a Ca<sup>2+</sup> channel inhibitor: Involvement of inhibiting contractility and Ca<sup>2+</sup> influx via L-type Ca<sup>2+</sup> channels. *Arch Pharm Res* 2020;43:808-820. <https://doi.org/10.1007/s12272-020-01261-y>
168. Huang W-F, Ouyang S, Li S-Y, Lin Y-F, Ouyang H, Zhang H, Lu C-J. Effect of quercetin on colon contractility and L-type Ca<sup>2+</sup> channels in colon smooth muscle of guinea-pig. *Acta Physiol Sin* 2009;61:567-576.

169. Tsujimae K, Suzuki S, Yamada M, Kurachi Y. Comparison of kinetic properties of quinidine and dofetilide block of HERG channels. *Eur J Pharmacol* 2004;493:29-40. <https://doi.org/10.1016/j.ejphar.2004.04.015>
170. Zhang R, Jie L-J, Wu W-Y, Wang Z-Q, Sun H-Y, Xiao G-S, Wang Y, Li Y-G, Li G-R. Comparative study of carvedilol and quinidine for inhibiting hKv4.3 channel stably expressed in HEK 293 cells. *Eur J Pharmacol* 2019;853:74-83. <https://doi.org/10.1016/j.ejphar.2019.03.029>
171. Hernandez-Cascales J. Resveratrol enhances the inotropic effect but inhibits the proarrhythmic effect of sympathomimetic agents in rat myocardium. *PeerJ* 2017;5:e3113. <https://doi.org/10.7717/peerj.3113>
172. Kazemirad H, Kazerani HR. Cardioprotective effects of resveratrol following myocardial ischemia and reperfusion. *Mol Biol Rep* 2020;47:5843-5850. <https://doi.org/10.1007/s11033-020-05653-7>
173. Barangi S, Hayes AW, Karimi G. The more effective treatment of atrial fibrillation applying the natural compounds; as NADPH oxidase and ion channel inhibitors. *Crit Rev Food Sci Nutr* 2018;58:1230-1241. <https://doi.org/10.1080/10408398.2017.1379000>
174. Osmanhoğlu HÖ, Nazıroğlu M. Resveratrol modulates diabetes-induced neuropathic pain, apoptosis, and oxidative neurotoxicity in mice through TRPV4 channel inhibition. *Mol Neurobiol* 2024;61:7269-7286. <https://doi.org/10.1007/s12035-024-04311-4>
175. Yu XC, Wu S, Chen CF, Pang K, Wong T. Antihypertensive and anti-arrhythmic effects of an extract of radix *Stephaniae tetrandrae* in the rat. *J Pharm Pharmacol* 2004;56:115-122. <https://doi.org/10.1211/0022357022458>
176. Li Q, Chang L, Su D-M, Ma X. Effects of tetrandrine on proliferation and activation of cardiac fibroblasts. *Beijing Da Xue Xue Bao Yi Xue Ban (Journal of Peking University Health Sciences)* 2018;50:331-334.
177. Zhou Y, Xu W, Han R, Zhou J, Pan Z, Rong H, Li J, Xu C, Qiao G, Lu Y. Matrine inhibits pacing induced atrial fibrillation by modulating IKM3 and ICa-L. *Int J Biol Sci* 2011;8:150. <https://doi.org/10.7150/ijbs.8.150>
178. Wei T, Liang Z, Jin Y, Zhang L. Effect of berberine, liensinine and neferine on hERG channel expression. *Zhong yao za zhi = Zhongyao Zazhi (Journal of Chinese Materia Medica)* 2013;38:239-244.
179. Wu S-N, Yu H-S, Jan C-R, Li H-F, Yu C-L. Inhibitory effects of berberine on voltage-and calcium-activated potassium currents in human myeloma cells. *Life Sci* 1998;62:2283-2294. [https://doi.org/10.1016/S0024-3205\(98\)00209-4](https://doi.org/10.1016/S0024-3205(98)00209-4)
180. Chen K-H, Liu H, Sun H-Y, Jin M-W, Xiao G-S, Wang Y, Li G-R. The natural flavone acacetin blocks small conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels stably expressed in HEK 293 cells. *Front Pharmacol* 2017;8:716. <https://doi.org/10.3389/fphar.2017.00716>
181. Yang B-F, Xu C-Q, Li Y-R, Du Z-M, Zhou J, Sun J-P. Inhibitory effect of artemisinin on cloned inward rectifier potassium channels. *Chin J Pharmacol Toxicol* 1999;13:245-248.
182. Lin Y-L, Dai Z-K, Lin R-J, Chu K-S, Chen J, Wu J-R, Wu B-N. Baicalin, a flavonoid from *Scutellaria baicalensis* Georgi, activates large-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels via cyclic nucleotide-dependent protein kinases in mesenteric artery. *Phytomedicine* 2010;17:760-770. <https://doi.org/10.1016/j.phymed.2010.01.003>
183. Redford KE, Abbott GW. The ubiquitous flavonoid quercetin is an atypical KCNQ potassium channel activator. *Commun Biol* 2020;3:356. <https://doi.org/10.1038/s42003-020-1089-8>
184. Hsu H-T, Tseng Y-T, Lo Y-C, Wu S-N. Ability of naringenin, a bioflavonoid, to activate M-type potassium current in motor neuron-like cells and to increase BKCa-channel activity in HEK293T cells transfected with  $\alpha$ -hSlo subunit. *BMC Neurosci* 2014;15:135. <https://doi.org/10.1186/s12868-014-0135-1>
185. Ortiz MI, Cariño-Cortés R, Muñoz-Pérez VM, Salas-Casas A, Castañeda-Hernández G. Role of the NO-cGMP-K<sup>+</sup> channels pathway in the peripheral antinociception induced by  $\alpha$ -bisabolol. *Can J Physiol Pharmacol* 2021;99:1048-1056. <https://doi.org/10.1139/cjpp-2020-0744>
186. Silva-Alves K, Ferreira-da-Silva F, Peixoto-Neves D, Viana-Cardoso K, Moreira-Júnior L, Oquendo M, Oliveira-Abreu K, Albuquerque A, Coelho-de-Souza A, Leal-Cardoso J. Estragole blocks neuronal excitability by direct inhibition of Na<sup>+</sup> channels. *Braz J Med Biol Res* 2013;46:1056-1063. <https://doi.org/10.1590/1414-431X20133191>
187. Patel DK, Patel K. Therapeutic potential and pharmacological activities of (+)-nantenine in medicine: an aporphine class phytocomponent of *Nandina domestica* Thunberg. *Infect Disord Drug Targets* 2024;24:e201023222495. <https://doi.org/10.2174/0118715265244269231010090316>

188. Quan Q-K, Li X, Yuan H-F, Wang Y, Liu W-L. Ginsenoside Rg1 inhibits high-voltage-activated calcium channel currents in hippocampal neurons of beta-amyloid peptide-exposed rat brain slices. *Chin J Integr Med* 2016;1-6. <https://doi.org/10.1007/s11655-015-2301-4>
189. Frias B, Merighi A. Capsaicin, nociception and pain. *Molecules* 2016;21:797. <https://doi.org/10.3390/molecules21060797>
190. Hong NR, Park HS, Ahn TS, Kim HJ, Ha K-T, Kim BJ. Ginsenoside Re inhibits pacemaker potentials via adenosine triphosphate-sensitive potassium channels and the cyclic guanosine monophosphate/nitric oxide-dependent pathway in cultured interstitial cells of Cajal from mouse small intestine. *J Ginseng Res* 2015;39:314-321. <https://doi.org/10.1016/j.jgr.2015.02.004>
191. Yao Z, Namkung W, Ko E, Park J, Tradtrantip L. Fractionation of a herbal antidiarrheal medicine reveals eugenol as an inhibitor of Ca<sup>2+</sup>-activated Cl<sup>-</sup> channel TMEM16A. *PLoS One* 2012;7:e38030. <https://doi.org/10.1371/journal.pone.0038030>
192. Thilakarathna SH, Rupasinghe HV. Flavonoid bioavailability and attempts for bioavailability enhancement. *Nutrients* 2013;5:3367-3387. <https://doi.org/10.3390/nu5093367>
193. Wu H-J, Wu W, Sun H-Y, Qin G-W, Wang H-B, Wang P, Yalamanchili HK, Wang J, Tse H-F, Lau C-P. Acacetin causes a frequency- and use-dependent blockade of hKv1.5 channels by binding to the S6 domain. *J Mol Cell Cardiol* 2011;51:966-973. <https://doi.org/10.1016/j.yjmcc.2011.08.022>
194. Xi S, Zhou G, Zhang X, Zhang W, Cai L, Zhao C. Protective effect of total aralosides of *Aralia elata* (Miq) Seem (TASAES) against diabetic cardiomyopathy in rats during the early stage, and possible mechanisms. *Exp Mol Med* 2009;41:538-547. <https://doi.org/10.3858/emm.2009.41.8.059>
195. Yang B-F, Li Y-R, Xu C, Luo D-L, Li B-X, Wang H, Zhou J. Mechanisms of artemisinin antiarrhythmic action. *Chin J Pharmacol Toxicol* 1999;13:169-175.
196. He J, Li S, Ding Y, Tong Y, Li X. Research progress on natural products' therapeutic effects on atrial fibrillation by regulating ion channels. *Cardiovasc Ther* 2022;2022:4559809. <https://doi.org/10.1155/2022/4559809>
197. Han X-H, Ping L, Zhang Y-Y, Zhang N, Chen F-R, Cai J-F. Astragaloside IV regulates expression of ATP-sensitive potassium channel subunits after ischemia-reperfusion in rat ventricular cardiomyocytes. *J Tradit Chin Med* 2011;31:321-326. [https://doi.org/10.1016/S0254-6272\(12\)60012-0](https://doi.org/10.1016/S0254-6272(12)60012-0)
198. Lu M, Wang H, Wang J, Zhang J, Yang J, Liang L, Maslov LN. Astragaloside IV protects against cardiac hypertrophy via inhibiting the Ca<sup>2+</sup>/CaN signaling pathway. *Planta Med* 2014;80:63-69. <https://doi.org/10.1055/s-0033-1360129>
199. Zhao M, Shao D, Yu L, Sun X, Wang Y, Hu H, Feng R, Gao Q, Guo F, Hao L. Electrophysiological effect and the gating mechanism of astragaloside IV on I-type Ca<sup>2+</sup> channels of guinea-pig ventricular myocytes. *Eur J Pharmacol* 2015;760:27-35. <https://doi.org/10.1016/j.ejphar.2015.03.082>
200. Yin B, Hou X-w, Lu M-l. Astragaloside IV attenuates myocardial ischemia/reperfusion injury in rats via inhibition of calcium-sensing receptor-mediated apoptotic signaling pathways. *Acta Pharmacol Sin* 2019;40:599-607. <https://doi.org/10.1038/s41401-018-0082-y>
201. Huang W, Zhang Z, Xu Y. Study of the effects and mechanisms of berberine on slow-response action potentials. *J Electrocardiol* 1990;23:231-234. [https://doi.org/10.1016/0022-0736\(90\)90161-T](https://doi.org/10.1016/0022-0736(90)90161-T)
202. Wang Y-X, Zheng Y-M, Zhou X-B. Inhibitory effects of berberine on ATP-sensitive K<sup>+</sup> channels in cardiac myocytes. *Eur J Pharmacol* 1996;316:307-315. [https://doi.org/10.1016/S0014-2999\(96\)00663-2](https://doi.org/10.1016/S0014-2999(96)00663-2)
203. Chang W, Li K, Guan F, Yao F, Yu Y, Zhang M, Hatch GM, Chen L. Berberine pretreatment confers cardioprotection against ischemia-reperfusion injury in a rat model of type 2 diabetes. *J Cardiovasc Pharmacol Ther* 2016;21:486-494. <https://doi.org/10.1177/1074248415627873>
204. Testai L, Chericoni S, Martelli A, Flamini G, Breschi MC, Calderone V. Voltage-operated potassium (Kv) channels contribute to endothelium-dependent vasorelaxation of carvedilol on rat aorta. *J Pharm Pharmacol* 2016;68:1177-1183. <https://doi.org/10.1111/jphp.12585>
205. Zhao J, Lian Y, Lu C, Jing L, Yuan H, Peng S. Inhibitory effects of a bisbenzylisoquinoline alkaloid dauricine on hERG potassium channels. *J Ethnopharmacol* 2012;141:685-691. <https://doi.org/10.1016/j.jep.2011.08.054>
206. Liu Q-N, Zhang L, Gong P-L, Yang X-Y, Zeng F-D. Inhibitory effects of dauricine on early afterdepolarizations and L-type calcium current. *Can J Physiol Pharmacol* 2009;87:954-962. <https://doi.org/10.1139/Y09-090>

207. Bai Y, Su Z, Sun H, Zhao W, Chen X, Hang P, Zhu W, Du Z. Aloe-emodin relieves high-fat diet induced QT prolongation via MiR-1 inhibition and IK1 up-regulation in rats. *Cell Physiol Biochem* 2018;43:1961-1973. <https://doi.org/10.1159/000484120>
208. Xiao GS, Zhang YH, Wu W, Sun HY, Wang Y, Li GR. Genistein and tyrphostin AG556 decrease ultra-rapidly activating delayed rectifier K<sup>+</sup> current of human atria by inhibiting EGF receptor tyrosine kinase. *Br J Pharmacol* 2017;174:454-467. <https://doi.org/10.1111/bph.13710>
209. Sun L, Zhao T, Ju T, Wang X, Li X, Wang L, Zhang L, Yu G. A combination of intravenous genistein plus Mg<sup>2+</sup> enhances antihypertensive effects in SHR by endothelial protection and BKCa channel inhibition. *Am J Hypertens* 2015;28:1114-1120. <https://doi.org/10.1093/ajh/hpv005>
210. Colareda GA, Ragone MI, Bonazzola P, Consolini AE. The mKATP channels and protein-kinase C are involved in the cardioprotective effects of genistein on estrogen-deficient rat hearts exposed to ischemia/reperfusion: energetic study. *J Cardiovasc Pharmacol* 2020;75:460-474. <https://doi.org/10.1097/FJC.0000000000000816>
211. Tolstikova T, Khvostov M, Bryzgalov A, Belenichev I, Pavlov S. Glycidipine, a promising hypotensive and cardioprotective agent. *Bull Exp Biol Med* 2011;151:597-600. <https://doi.org/10.1007/s10517-011-1391-z>
212. Qi ZL, Wang Z, Li W, Hou JG, Liu Y, Li XD, Li HP, Wang YP. Nephroprotective effects of anthocyanin from the fruits of *Panax ginseng* (GFA) on cisplatin-induced acute kidney injury in mice. *Phytother Res* 2017;31:1400-1409. <https://doi.org/10.1002/ptr.5867>
213. Gao J, Wang T, Yao X, Xie W, Shi X, He S, Zhao T, Wang C, Zhu Y. Clinical evidence-guided network pharmacology analysis reveals a critical contribution of  $\beta$ 1-adrenoreceptor upregulation to bradycardia alleviation by Shenxian-Shengmai. *BMC Complement Altern Med* 2019;19:357. <https://doi.org/10.1186/s12906-019-2769-0>
214. Mohammed ASA, Mohácsi G, Naveed M, Prorok J, Jost N, Virág L, Baczkó I, Topal L, Varró A. Cellular electrophysiological effects of the citrus flavonoid hesperetin in dog and rabbit cardiac ventricular preparations. *Sci Rep* 2024;14:7237. <https://doi.org/10.1038/s41598-024-57828-y>
215. Xu Y, Leung S, Leung G, Man R. Kaempferol enhances endothelium-dependent relaxation in the porcine coronary artery through activation of large-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels. *Br J Pharmacol* 2015;172:3003-3014. <https://doi.org/10.1111/bph.13108>
216. Wan J, Zhao L, Xu C, Zhang S, Zhang Z, Zeng C, Chang M, Xiao J, Wang J. Effects of neferine on the pharmacokinetics of amiodarone in rats. *Biomed Chromatogr* 2011;25:858-866. <https://doi.org/10.1002/bmc.1535>
217. Wang C, Wang H, Xiao J-H, Wang J-L, Xiang J-Z, Tang Q. Inhibitory effects of neferine on Nav1.5 channels expressed in HEK293 cells. *J Huazhong Univ Sci Technol Med Sci* 2016;36:487-493. <https://doi.org/10.1007/s11596-016-1613-8>
218. Zhuo Y, Yuan R, Chen X, He J, Chen Y, Zhang C, Sun K, Yang S, Liu Z, Gao H. Tanshinone I exerts cardiovascular protective effects *in vivo* and *in vitro* through inhibiting necroptosis via Akt/Nrf2 signaling pathway. *Chin Med* 2021;16:48. <https://doi.org/10.1186/s13020-021-00458-7>
219. Chen Y, Li J, Huang Q. Effects of *Panax notoginseng* saponins on rat cardiomyocytes apoptosis induced by angiotensin II *in vitro*. *Zhongguo Zhong Yao Za Zhi (Journal of Chinese Materia Medica)* 2005;30:778-781.
220. Zhang H, Zhang L, Zhang Q, Yang X, Yu J, Shun S, Wu Y, et al. Puerarin: a novel antagonist to inward rectifier potassium channel (IK1). *Mol Cell Biochem* 2011;352:117-123. <https://doi.org/10.1007/s11010-011-0746-0>
221. Xu H, Zhao M, Liang S, Huang Q, Xiao Y, Ye L, Wang Q, He L, Ma L, Zhang H. The effects of puerarin on rat ventricular myocytes and the potential mechanism. *Sci Rep* 2016;6:35475. <https://doi.org/10.1038/srep35475>
222. Guo X, Chen J, Zhang X, Xia Q. Effect of puerarin on L-type calcium channel in isolated rat ventricular myocytes. *Zhongguo Zhong Yao Za Zhi = Zhongguo Zhongyao Zazhi = China Journal of Chinese Materia Medica* 2004;29:248-251.
223. Li Z, Jin X, Wu T, Huang G, Wu K, Lei J, Pan X, Yan N. Structural basis for pore blockade of the human cardiac sodium channel Nav1.5 by the antiarrhythmic drug quinidine. *Angew Chem Int Ed Engl* 2021;60:11474-11480. <https://doi.org/10.1002/anie.202102196>
224. Haworth RA, Goknur AB, Berkhoff HA. Inhibition of ATP-sensitive potassium channels of adult rat heart cells by antiarrhythmic drugs. *Circ Res* 1989;65:1157-1160. <https://doi.org/10.1161/01.RES.65.4.1157>

225. Pan MC, Zhou XW, Liu Y, Wang YN, Qiu XG, Wu SF, Liu Q. Research progress on the molecular mechanisms of toxicology of ethanol-aconitine induced arrhythmia. *Fa Yi Xue Za Zhi (Journal of Forensic Medicine)* 2020;36:115-119.
226. Liu W, Chen P, Deng J, Lv J, Liu J. Resveratrol and polydatin as modulators of Ca<sup>2+</sup> mobilization in the cardiovascular system. *Ann N Y Acad Sci* 2017;1403:82-91. <https://doi.org/10.1111/nyas.13386>
227. Chen Y-R, Yi F-F, Li X-Y, Wang C-Y, Chen L, Yang X-C, Su P-X, Cai J. Resveratrol attenuates ventricular arrhythmias and improves the long-term survival in rats with myocardial infarction. *Cardiovasc Drugs Ther* 2008;22:479-485. <https://doi.org/10.1007/s10557-008-6141-8>
228. Sutanto H, Dobrev D, Heijman J. Resveratrol: an effective pharmacological agent to prevent inflammation-induced atrial fibrillation? *Naunyn-Schmiedeberg's Arch Pharmacol* 2018;391:1163-1167. <https://doi.org/10.1007/s00210-018-1566-5>
229. Baczkó I, Liknes D, Yang W, Hamming KC, Searle G, Jaeger K, Husti Z, et al. Characterization of a novel multifunctional resveratrol derivative for the treatment of atrial fibrillation. *Br J Pharmacol* 2014;171:92-106. <https://doi.org/10.1111/bph.12409>
230. Nieto-Marin P, Jimenez-Jaimez J, Tinaquero D, Alfayate S, Utrilla RG, Perin F, Sarquella-Brugada G, Monserrat L, Brugada J, Tercedor L. Digenic heterozygosity in SCN5A and CACNA1C explains the variable expressivity of the long QT phenotype in a Spanish family. *Rev Esp Cardiol (Engl Ed)* 2019;72:324-332. <https://doi.org/10.1016/j.rec.2018.03.012>
231. Huber I, Wappl E, Herzog A, Mitterdorfer J, Glossmann H, Langer T, Striessnig J. Conserved Ca<sup>2+</sup>-antagonist-binding properties and putative folding structure of a recombinant high-affinity dihydropyridine-binding domain. *Biochem J* 2000;347:829-836. <https://doi.org/10.1042/bj3470829>
232. Wu S-N, Li H-F, Lo Y-C. Characterization of tetrandrine-induced inhibition of large-conductance calcium-activated potassium channels in a human endothelial cell line (HUV-EC-C). *J Pharmacol Exp Ther* 2000;292:188-195. [https://doi.org/10.1016/S0022-3565\(24\)35276-0](https://doi.org/10.1016/S0022-3565(24)35276-0)
233. Wang H-X, Kwan C-Y, Wong T-M. Tetrandrine inhibits electrically induced [Ca<sup>2+</sup>]<sub>i</sub> transient in the isolated single rat cardiomyocyte. *Eur J Pharmacol* 1997;319:115-122. [https://doi.org/10.1016/S0014-2999\(96\)00834-5](https://doi.org/10.1016/S0014-2999(96)00834-5)
234. Huang C-W, Chow JC, Tsai J-J, Wu S-N. Characterizing the effects of eugenol on neuronal ionic currents and hyperexcitability. *Psychopharmacology* 2012;221:575-587. <https://doi.org/10.1007/s00213-011-2603-y>
235. Yao Z, Namkung W, Ko EA, Park J, Tradtrantip L, Verkman A. Fractionation of a herbal antiarrhythmic medicine reveals eugenol as an inhibitor of Ca<sup>2+</sup>-activated Cl<sup>-</sup> channel TMEM16A. *PloS one* 2012;7:e38030. <https://doi.org/10.1371/journal.pone.0038030>
236. Vatanparast J, Khalili S, Naseh M. Dual effects of eugenol on the neuronal excitability: An in vitro study. *Neurotoxicology* 2017;58:84-91. <https://doi.org/10.1016/j.neuro.2016.11.011>
237. Dimpfel W. Different anticonvulsive effects of hesperidin and its aglycone hesperetin on electrical activity in the rat hippocampus in-vitro. *J Pharm Pharmacol* 2006;58:375-379. <https://doi.org/10.1211/jpp.58.3.0012>
238. Zhou Y, Cai S, Moutal A, Yu J, Gómez K, Madura CL, Shan Z, Pham NY, Serafini MJ, Dorame A. The natural flavonoid naringenin elicits analgesia through inhibition of NaV1.8 voltage-gated sodium channels. *ACS Chem Neurosci* 2019;10:4834-4846. <https://doi.org/10.1021/acscchemneuro.9b00547>
239. Mukhopadhyay M, Singh A, Sachchidanand S, Bera AK. Quercetin inhibits acid-sensing ion channels through a putative binding site in the central vestibular region. *Neuroscience* 2017;348:264-272. <https://doi.org/10.1016/j.neuroscience.2017.02.025>
240. Yao Y, Han D, Zhang T, Yang Z. Quercetin improves cognitive deficits in rats with chronic cerebral ischemia and inhibits voltage-dependent sodium channels in hippocampal CA1 pyramidal neurons. *Phytother Res* 2010;24:136-140. <https://doi.org/10.1002/ptr.2902>
241. Cermak R, Kuhn G, Wolfram S. The flavonoid quercetin activates basolateral K<sup>+</sup> channels in rat distal colon epithelium. *Br J Pharmacol* 2002;135:1183-1190. <https://doi.org/10.1038/sj.bjp.0704564>
242. Tan X-Q, Cheng X-L, Yang Y, Yan L, Gu J-L, Li H, Zeng X-R, Cao J-M. Tanshinone II-A sodium sulfonate (DS-201) enhances human BKCa channel activity by selectively targeting the pore-forming  $\alpha$  subunit. *Acta Pharmacol Sin* 2014;35:1351-1363. <https://doi.org/10.1038/aps.2014.85>

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243. Lin TY, Lu CW, Huang S-K, Wang S-J. Tanshinone IIA, a constituent of Danshen, inhibits the release of glutamate in rat cerebrocortical nerve terminals. *J Ethnopharmacol* 2013;147:488-496. <https://doi.org/10.1016/j.jep.2013.03.045>
  244. Aron de Miranda HA, Gonçalves JCR, Cruz JS, Araújo DAM. Evaluation of the sesquiterpene (-)- $\alpha$ -bisabolol as a novel peripheral nervous blocker. *Neurosci Lett* 2010;472:11-15. <https://doi.org/10.1016/j.neulet.2010.01.042>
  245. Gadotti VM, Huang S, Zamponi GW. The terpenes camphene and alpha-bisabolol inhibit inflammatory and neuropathic pain *via* Cav3.2 T-type calcium channels. *Mol Brain* 2021;14:166. <https://doi.org/10.1186/s13041-021-00876-62>.
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