## **SHORT COMMUNICATION**

# Reciprocal Adaptive Response of Human Peripheral Lymphocytes Induced by Bleomycine or Gamma Rays

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## Summary

The adaptive response and reciprocal adaptive response induced *in vitro* by exposure to low doses of gamma rays (0.05 Gy) or bleomycin (0.05 µg/ml) in human peripheral blood lymphocytes were assessed by the frequency of chromosome aberrations. Gamma rays (1.5 Gy) or bleomycin (1.5 µg/ml) were used as the challenge doses. In the experiments, blood samples from 5 healthy donors were investigated. It has been found that low doses of bleomycin and gamma rays induced a reciprocal adaptive response to high doses of gamma rays or bleomycin. Moreover, the results confirmed that the adaptive response did not correlate with the radiosensitivity of the peripheral blood lymphocytes.

## Key words

Adaptive response • Reciprocal adaptive response • Gamma rays • Bleomycin • Chromosome aberration

The ability of cells to become more resistant to high doses of mutagens after a preceding exposure to low doses has been termed the adaptive response (AR). For evaluating this adaptive response various indicators have been employed, e.g. the frequency of chromosomal aberrations, sister chromatid exchanges, micronuclei, gene mutations and survival of cells. Olivieri *et al.* (1984) first reported that the pre-treatment of stimulated human lymphocytes with low level of radioactive thymidine (<sup>3</sup>H-TdR) might cause a significant reduction in chromosomal aberrations after subsequent high doses of X-ray irradiation. Later, it was found that some chemomutagens and carcinogens have a similar ability to

induce AR (Wolff 1996, Wolff et al. 1988, Vijayalaxmi and Burkart 1989, Cortes et al. 1990). The phenomenon of reciprocal AR was observed by Wolff et al. (1988) when the adaptive agent differs from challenging one. As regards to mechanisms of the AR some authors (Wiencke et al. 1986, Wolff et al. 1990, Ikushima et al. 1996) have attributed this to a repair process that might be controlled by enzyme poly(ADP-ribose)polymerase. The present paper deals with the induction of reciprocal AR measured by the frequency of chromosome aberrations in human peripheral blood lymphocytes after in vitro exposure to low doses of bleomycin (BLM) or gamma irradiation.

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Table 1. Frequency of chromosome aberrations in human peripheral lymphocytes adapted by bleomycin or gamma rays

Donor	Treatment group	No. Of metaphases scored	No. o	f chromosome aabberations %	Expected [%] [%]	AR
D1	Control	200	2	1.0		
	1.5 µg BLM/ml	190	70	36.8		
	1.5 Gy	200	40	20.0		
	0.05 µg BLM/ml	200	4	2.0		
	0.05 µg BLM/ml + 1.5 Gy	198	29	14.5	21.0	31.00
	0.05 µg BLM/ml + 1.5 µg BLM/ml	200	41	20.5	37.8	45.80
	0.05 Gy	200	2	1.0		
	0.05 Gy + 1.5 Gy	200	49	24.5	36.6	33.08
	$0.05 \text{ Gy} + 1.5 \mu \text{g BLM./ml}$	200	22	11.0	18.5	40.50
D2	Control	200	3	1.5		
	1.5 µg BLM/ml	200	72	36.0		
	1.5 Gy	200	33	16.5		
	0.05 µg BLM/ml	190	3	1.6		
	$0.05 \ \mu g \ BLM/ml + 1.5 \ Gy$	200	20	10.0	16.6	39.70
	$0.05 \mu g BLM/ml + 1.5 \mu g BLM/ml$	200	42	21.0	36.1	41.80
	0.05 Gy	200	3	1.5		
	0.05 Gy + 1.5 Gy	200	43	21.5	34.5	37.70
	0.05 Gy + 1.5 µg BLM/ml	200	25	12.5	22.5	44.50
D3	Control	200	2	1.0		
	1.5 µg BLM/ml	197	36	18.3		
	1.5 Gy	200	41	20.5		
	0.05 µg BLM/ml	200	3	1.5		
	0.05 µg BLM/ml + 1.5 Gy	200	38	19.0	21.0	9.50 NS
	$0.05 \mu g BLM/ml + 1.5 \mu g BLM/ml$	200	35	17.5	18.8	6.90 NS
	0.05 Gy	200	2	1.0		
	0.05  Gy + 1.5  Gy	200	33	16.5	30.3	45.50
	0.05 Gy + 1.5 µg BLM/ml	192	32	16.0	23.5	31.20
D4	Control	200	3	1.5		
	1.5 µg BLM/ml	200	47	23.5		
	1.5 Gy	200	33	16.5		
	0.05 µg BLM/ml	196	3	1.5		
	0.05 µg BLM/ml + 1.5 Gy	200	20	10.0	16.5	39.44
	0.05 µg BLM/ml + 1.5 µg BLM/ml	200	32	16.0	23.5	31.90
	0.05 Gy	200	3	1.5		
	0.05 Gy + 1.5 Gy 0.05 Gy + 1.5 µg BLM/ml	200 200	24 40	12.0 20.0	29.2 26.5	58.90 24.50
	· · · · · · ·		40	20.0	20.5	24.30
D5	Control	200	4	2.0		
	1.5 µg BLM/ml	200	64	32.0		
	1.5 Gy	200	45	22.5		
	0.05 µg BLM/ml	200	4	2.0	aa -	
	0.05 µg BLM/ml + 1.5 Gy	200	37 52	18.5	22.5	17.80 NS
	0.05 µg BLM/ml + 1.5 µg BLM/ml	200	52.	26.0	32.0	18.80 NS
	0.05 Gy	200	5	2.5	26.5	46.00
	0.05 Gy + 1.5 Gy	198	28	14.1	26.5	46.80
	0.05 Gy + 1.5 µg BLM/ml	200	32	16.0	35.3	54.70

NS - non-significant adaptive response

Whole blood cultures in duplicates from five normal healthy donors (25-40 years old) were set up in 5 ml of RPMI 1640 medium (SEVAC, Prague, Czech Republic) supplemented with 20 % calf serum, (penicillin and streptomycin), antibiotics phytohemagglutinine (HA15, Welcome, UK). Two hours before harvesting and fixation, Colcemid (2.10<sup>-7</sup> M) was added to the cultures. Fixation was performed six hours after the challenge doses in both series of experiments.

Bleomycin (BLM, Nippon Kayaku) served as the adaptive dose in a low concentration (0.05 µg.ml<sup>-1</sup>) in the first series experiments and was used for 24 h after lymphocyte stimulation. The challenge dose of BLM (1.50 µg.ml<sup>-1</sup>) was delivered after 44 h of cultivation.

In the second series of experiments, gamma radiation (0.05 Gy) was used as adaptive dose applied 32 h after stimulation. The challenge dose of gamma rays (1.50 Gy) was delivered after 44 h of cultivation. A Chisostat 60Co apparatus was used as a source of gamma rays. The dose rate was 0.5 Gy per minute.

Chromatid and isochromatid breaks were recorded in both series of experiments. The statistical significance of the reduction of chromosome aberrations was determined using  $\chi^2$ -test with Yates's correction. The expected value of chromosome aberrations was calculated as the sum of individual doses (adaptive + challenge) minus the control value. The adaptive response was expressed as a percentage of the decreased chromosome aberration frequency.

Experimental data concerning the AR induced by low doses of BLM or gamma rays are summarized in Table 1. It can be seen that pre-treatment with the low dose of BLM induced the AR and reciprocal AR in donors D1, D2 and D4, but not in D3 and D5. When the low dose of gamma rays was used as the adaptive dose, the AR was observed in all five donors. The results are in good agreement with the results published earlier on reciprocal AR (Wolff et al. 1988, Vijayalaxmi and Burkart 1989). They indicate that the process of induction of the repair system by low adaptive doses of BLM and gamma rays is probably the same. The absence of AR in donors D3 and D5 after low dose of BLM may be the result of some physiological mechanisms that do not allow the induction of adaptive repair mechanisms in their lymphocytes. The presented results demonstrate an apparent heterogeneity in the AR in the cells of investigated subjects that might partly be determined genetically (Bosi and Olivieri 1989, Kalina and Némethová 1996). The heterogeneity of the AR may also be related to its magnitude and to adaptive factors. Further results have confirmed that the AR did not correlate with the radiosensitivity of peripheral blood lymphocytes.

Recently, it was suggested that the evaluation of AR should be performed in cells of individuals exposed in vivo to low doses of environmental and occupational mutagens (Šrám 1996). Barquinero et al. (1996) observed that occupational exposure to very low doses of ionizing radiation induce the AR which could be detected by a challenge dose of BLM. Similar results on the inducibility of AR in vivo by ionizing radiation in case of the Chernobyl catastrophe have been reported by Tedeschi et al. (1996). An open question, which still remains to be answered, concerns the problem how the AR resulting in refractoriness of cells to mutagens is related to individual radioresistance of patients, e.g. to chemotherapeutic drugs. More information in this direction might also be of importance for clinical medicine.

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#### Reprint requests

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