

Metabolomics and Lipidomics for Studying Metabolic Syndrome: Insights into Cardiovascular Diseases, Type 1 & 2 Diabetes, and Metabolic Dysfunction-Associated Steatotic Liver Disease

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Supplementary Materials

Table S1. Metabolomics and lipidomics cohort studies focused on cardiovascular disease

Table S2. Metabolomics and lipidomics cohort studies focused on type 1 and type 2 diabetes

Table S3. Metabolomics and lipidomics cohort studies focused on metabolic dysfunction-associated steatotic liver disease

Table S1. Metabolomics and lipidomics cohort studies focused on cardiovascular disease

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
Discovery cohort: n = 4,824 (27.8% female) Replication cohort: n = 1,716 (56.3% female)	Participants with 15.8 years follow-up	Serum	NMR	41	Isoleucine Leucine Phenylalanine Glycerol Cholesterol Total lipid concentrations, Glycerides and other Phospholipids, Fatty acids, Fatty acids ratios — see the original paper.	Association with adherence to dietary recommendations provided by the Alternative Healthy Eating Index	[1]
European cohort: n = 352 USA cohort: n = 1,777	European participants (100% Caucasian) with either abdominal aortic aneurysm or sub-aneurysmal aortic dilations, and healthy non-aneurysm subjects US participants (96% Caucasian) with abdominal aortic diameter of 3.0 cm or greater, and subjects with history of dilated aorta with measurements of abdominal aortic diameter less than 3 cm or no prior aortic aneurysm, and no MI, stroke or death over the following 3 years	Plasma	LC-MS/MS	3	Choline Trimethylamine <i>N</i> -oxide Trimethylamine	Association of elevated TMAO with increased abdominal aortic aneurysm incidence	[2]
Low-risk cohort: n = 620 Borderline-risk cohort: n = 110 Intermediate-risk cohort: n = 225 Highrisk cohort: n = 147 (53.3% female)	Participants with LDL levels less than 190 mg/dl and no pre-existing coronary artery disease or myocardial infarction	Plasma	LC-MS/MS	50	Alanine Arginine Aspartic acid CAR 4:0-DC CAR 8:1 CAR 16:0-OH Citrulline Glutamic acid Glutamine Glycine Histidine Phenylalanine Threonine Tryptophan	- Association with the 10-year ASCVD risk score - Identification of metabolic pathways associated with the development of 10-year ASCVD events	[3]
EPIC-Potsdam Study cohort: Common reference subcohort: n = 1262 T2D subcohort: n = 1886 (775 incident cases) CVD subcohort: n = 1671 (551 incident cases) DIVAS study cohort: CVD risk subcohort: n = 113 (on 3 different isoenergetic diets)	General population Patients with estimated moderate CVD risk	Plasma	DMS-MS/MS	282	CE 20:3 DG 16:0 DG 18:0 FA 15:0 FA 20:4 LPC 18:2 MG 15:0 MG 20:4 PC 20:3 PE 20:3 TG 16:0 TG 18:0 TG 18:2 TG 18:3 TG 22:1	- Association with cardiometabolic disease risk and T2D risk - Dietary fat intervention as a potential tool for primary disease prevention	[4]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
Discovery cohort: n = 1,162 (36.3% female) Validation cohorts: n = 2,331 (US, 33.2% female), n = 832 (European, 29.9% female)	Stable participants undergoing elective diagnostic cardiac evaluation	Plasma	LC-MS/MS	5	<i>N</i> ¹ -Methyl-2-pyridone-5-carboxamide <i>N</i> ¹ -Methyl-4-pyridone-3-carboxamide Phenylacetylglutamine Trimethylamine <i>N</i> -oxide Trimethyllysine	Association of terminal breakdown products of excess niacin with residual CVD risk	[5]
Phase I: Discovery cohort: n = 3,613 Validation cohorts: n = 121,733 Phase II: n = 118,120	UK Biobank participants have undergone a wide range of physical measures, provided information on their lifestyle and medical history (follow-up)	Plasma	NMR	111	Multiple markers – see the original paper.	- Association with a healthy lifestyle - Association of healthy lifestyle-associated metabolites with coronary artery disease (CAD)	[6]
Discovery cohort: n = 1,028 Validation cohort: n = 1,670	Discovery cohort: Participants free of coronary heart disease (10 years follow-up)	Plasma	LC-MS/MS	32	LPC 18:1 LPC 18:2 MG 18:2 SM d28:1	- Association of MG 18:2 with coronary heart disease - Association of LPCs with body mass index, C-reactive protein and with less evidence of subclinical CVD	[7]
Discovery cohort: n = 1,833 (57% female) Validation cohorts: n = 1,522 Low walnut intake subcohort: n = 691 High walnut intake subcohort: n = 467	Participants at high cardiovascular risk	Plasma	LC-MS	385	4-Hydroxy-3-methylacetophenone Cyclohexylamine Guanine Isocitric acid <i>N</i> -Acetylaspartic acid Piperine Serine Sorbitol Succinic acid Bilirubin Biliverdin CAR 10:2 LPC 14:0 LPC 16:1 MG 22:1 PC 36:4 PE 36:5 PS 40:6 TG 54:6	Association of walnut consumption with a lower risk of incident T2D and CVD in a Mediterranean population at high cardiovascular risk	[8]
Study cohort: n = 1,057	Participants with symptomatic coronary artery disease	Blood-platelets	LC-MS/MS	767	CAR 10:0 CAR 14:0 CAR 14:1 CAR 16:0 CAR 16:1 FA 18:1 FA 18:2 FA 18:2;2O LPE 18:1 LPE 0:0/18:1 LPE 18:1 LPE 18:1/0:0 LPE 18:2 LPE 0:0/18:2 LPE 18:2 LPE 18:2/0:0 LPE 20:1 LPE 20:1/0:0 LPE 20:3 LPE 0:0/20:3 LPE 20:3 LPE 20:3/0:0 LPE 20:4 LPE 20:5 LPE 22:4 LPE 0:0/22:4 LPE 22:4 LPE 22:4/0:0 LPE 22:5	Association of adverse cardiovascular events with alterations in the platelet lipidome	[9]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
					LPE 22:6 LPS 18:1 LPS 0:0/18:1 PC 34:2;O PE 34:3 PE 16:1_18:2;O PI 36:4 PI 16:0_20:4 PI 38:5 PI 18:1_20:4 TG 48:1 TG 14:0_16:0_18:1 TG 48:2 TG 16:0_14:1_18:1		
Study cohort: n = 1,021 (48.3% female)	Participants with T2D and were followed up for CVD over the subsequent 10 years	Serum	NMR	228	3-Hydroxybutyric acid Acetic acid Creatinine Glycine Lactic acid Leucine Phenylalanine	• Association with 10-year cardiovascular risk in people with type 2 diabetes • Metabolite-based risk score created	[10]
Malmo Diet and Cancer-Cardiovascular cohort: n = 4,067	General population followed up to 23 years and stratified into risk groups	Plasma	DI-MS/MS	184	<i>Sum of lipid subclasses:</i> Ceramide Cholesteryl ester Cholesterol Diacylglycerol Ether-phosphatidylcholine Ether-phosphatidylethanolamine Lysophosphatidylcholine Lysophosphatidylethanolamine Phosphatidylcholine Phosphatidylethanolamine Phosphatidylinositol	• Possible identification of lipidomic risk before disease incidence (CVD and T2D)	[11]
Discovery cohort 1: n = 99 Discovery cohort 2: n = 1,162 Validation cohort: n = 2,140	Sequential stable subjects without evidence of acute coronary syndrome undergoing elective diagnostic coronary angiography for evaluation of CAD with longitudinal (3–5 years) follow-up	Plasma	HILIC-MS/MS LC-MS/MS		Trimethyllysine Trimethylamine <i>N</i> -oxide	• Association with CVD risks	[12]
Study cohort: n = 2,278 (50% female)	Participants were followed up for CVD incident (almost 10 years)	Plasma	LC-MS/MS	790 (37)	Dimethylglycine <i>N</i> -Acetylmethionine (top findings)	• Association with CVD risks	[13]
Study cohort: n = 5,072	Participants with diabetes	Plasma	NMR	44	3-Hydroxybutyric acid Acetic acid Acetoacetic acid Acetone Alanine Citric acid Creatinine Glucose Glutamine Glycine Histidine Isoleucine Lactic acid Leucine Phenylalanine Pyruvic acid Tyrosine Valine	• Association of multiple healthy lifestyle factors with improved circulating metabolites from different pathways	[14]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
Discovery cohort: n = 1,833 (57.6% female) Validation subcohort: n = 1,522	Participants at high risk of CVD (1-year follow-up)	Plasma	LC-MS/MS	382	1-Methylguanine γ-Aminobutyric acid (GABA) Aminoisobutyric acid Asparagine Cortisol Creatine Cytosine Glycodeoxycholic acid Hippuric acid Homoarginine Hypoxanthine Lactic acid Lysine N ¹ -Acetylspermidine N-Acetylaspartic acid N-Acetylorithine N-Carbamoyl-β-alanine Piperine Pyroglutamic acid Sorbitol Sucrose Trimethylbenzene CAR 7:0 CAR 18:2 CAR 18:0 DG 34:3 DG 36:0 LPC 16:1 MG 22:1 PC 34:3 PC 36:4 PC 38:4 PE 32:0 PE 38:6 PE 40:7 SM d34:2 SM d18:1/16:1 TG 50:3 TG 50:4 TG 55:2 TG 56:2	Association of legume consumption with T2D incidence, but not with CVD incidence risk	[15]
Study cohort: n ₁ = 5,991; n ₂ = 3,779 (38.9% female)	Participants with an 8-year follow-up	Plasma	LC-MS/MS	342	CE 24:0 LPI 18:2 PC 38:5 PC O-34:2 PC O-36:1 PC P-40:6 PE 38:6 PI 38:3 SM d42:1	Association with future cardiovascular events and cardiovascular death	[16]
Discovery cohort: n = 1,162 Validation cohort: n = 4,000	Sequential stable subjects undergoing elective diagnostic cardiac evaluation with longitudinal (3 years) follow-up	Plasma	HILIC-MS LC-MS/MS	5 (top-ranked)	Phenylacetylglutamine	Association with cardiovascular disease and death in humans	[17]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
Discovery cohort: n = 1,149 Validation cohort: n = 3,954	Participants with preserved kidney function undergoing elective diagnostic cardiac evaluation with longitudinal follow-up (5 years)	Plasma	GC-MS LC-MS/MS	N/A	<i>p</i> -Cresol sulfate Indoxyl sulfate	Association with CVD risk and overall mortality	[18]
Study cohort: n = 2,627	Participants were invited to attend a health examination for additional tests and collection of 8–12 h fasting blood samples (mean 12.9 years follow-up)	Plasma	HILIC-MS LC-MS/MS	79	Hex2Cer d34:2 Hex2Cer d18:2/16:0 HexCer d36:1 HexCer d18:1/18:0 HexCer d34:1 HexCer d18:1/16:0 HexCer d42:2 HexCer d18:2/24:0 SM d34:1 SM d18:1/16:0 SM d36:1 SM d18:1/18:0 SM d36:2 SM d18:2/18:0 SM d42:1 SM d18:1/24:0	Association with higher CVD risk	[19]
Study cohort: n ₁ =50; n ₂ =4,007	Healthy participants before and after the suppression of intestinal microbiota with oral broad-spectrum antibiotics underwent phosphatidylcholine challenge (ingestion of two hard-boiled eggs and deuterium [<i>d</i>₉]-labeled phosphatidylcholine) Participants undergoing elective diagnostic cardiac catheterization with no history of acute coronary syndrome	Plasma	LC-MS/MS	3	Betaine Choline Trimethylamine <i>N</i> -oxide	Association among intestinal microbiota-dependent metabolism of dietary phosphatidylcholine, TMAO levels, and adverse CVD events	[20]
Discovery cohort: n = 3,867 Validation cohort: n = 3,569	Participants were free of known CVD at baseline	Serum	NMR	N/A	1,5-Anhydrosorbitol 1-Methylhistidine 3-Hydroxybutyric acid 5-Oxoproline Acetaminophen + glucuronide Alanine Aspartic acid Citric acid Glucose Glutamic acid Glutamine Glycerol Glycine Histidine Lactic acid Lysine Mannose Methionine myo-Inositol Dimethylglycine Phenylalanine Glycerol groups of lipids Lipids (CH ₂ -CO) Lipids (CH ₂ -CH ₂ -C=, CH ₂ -CH ₂ -CO) Lipids (CH ₂ -CH ₂ -CH=CH) Lipids (CH ₃ -CH ₂ -R, (CH ₂) _n) Lipids (CH ₃ -CH ₂ -R, CH ₃ -CH ₂ -C=)	Association with atherosclerosis and incident CVD	[21]
Discovery cohort: n = 50 Validation cohort: n = 25	Stable patients undergoing elective cardiac evaluation who subsequently experienced a heart attack, stroke or death over the ensuing three-year period vs. age- and gender-matched subjects who did not	Plasma	LC-MS/MS	18	Betaine Choline Trimethylamine <i>N</i> -oxide	- Identification of markers as predictors of CVD risk - Discovery of a relationship between gut-flora-dependent metabolism of dietary	[22]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
						phosphatidylcholine and CVD pathogenesis	
Discovery cohort: n = 1,157 Validation cohorts: n ₁ = 2,149; n ₂ = 833	Stable subjects undergoing cardiac risk assessment Healthy volunteers (n = 8)	Plasma	GC-MS LC-MS/MS	N/A	Creatinine Erythritol Xylitol	Association with major adverse cardiovascular event	[23]
Discovery cohort: n = 1,157 Validation cohort: n = 2,149	Stable subjects undergoing elective diagnostic cardiac evaluations Healthy volunteers (n = 10)	Plasma	GC-MS LC-MS/MS	N/A	Creatinine Erythritol Xylitol	Association with major adverse cardiovascular event	[24]
Discovery cohort: n = 7,256 Validation cohorts: n ₁ = 2,622; n ₂ = 3,563	Participants were followed up for CVD incident (15 years)	Serum	NMR	68	3-Hydroxybutyric acid Acetic acid Acetoacetatic acid Alanine Citratic acid Glucose Glutamine Glycerol Glycine Histidine Isoleucine Lactic acid Leucine Phenylalanine Pyruvic acid Tyrosine Valine Docosahexaenoic acid (FA 22:6) Linoleic acid (FA 18:2) Monounsaturated FA Omega-3 FA Omega-6 FA Polyunsaturated FA Saturated FA	Association with incident CVD	[25]
Study cohort: n = 4,007	Participants undergoing elective diagnostic cardiac catheterization with no history of an acute coronary syndrome	Plasma	LC-MS/MS	18	Choline Trimethylamine Trimethylamine <i>N</i> -oxide	- Discovery of increased levels of TMAO as a predictor of incident risk for thrombotic events - Association between specific dietary nutrients, gut microbes, platelet function, and thrombosis risk	[26]

Table S2. Metabolomics and lipidomics cohort studies focused on type 1 and type 2 diabetes

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
Study cohort: n = 170 (48% female)	Children with high genetic risk for T1D	Plasma	GC-MS LC-MS/MS	91	γ-Aminobutyric acid (GABA) Glycine Tagatose Arabitol myo-Inositol Adipic acid Cer d38:1 Cer d39:1 LPC 18:3 LPC 20:3 LPC 20:5 SM d41:2	Utilization of multi-omics data for the modeling of complex, multifactorial diseases, like T1D	[27]
Study cohort: n = 152 (47.4% female)	Children with T1D (n=76) and healthy control children (n=76)	Cord blood serum	LC-MS/MS	106	PC 32:1 PC 36:4 PC 38:4 PC 38:5 PC 38:5 PC 38:6 PC 40:4 PC 40:5 PC 40:5 PC 40:7 PC 40:8 PC sum PE 38:4 PE 38:4 PE 40:4	Cord-blood metabolic patterns may be a valuable measure of type 1 diabetes risk	[28]
Study cohort: n = 101 (37.6% female)	Children who progressed to T1D (PT1D; n = 30), children who developed at least one islet autoantibody but did not progress to T1D during the follow-up (P1Ab; n = 33), and their age-matched controls (CTR; n = 38)	Cord blood plasma	LC-MS/MS	232 lipid species	CE 18:2 TG 46:2 TG 46:2 TG 48:1 TG 51:3	- Identification of lipids that can be predictive of the risk of progression to T1D - Comparison of lipidomic profiles of all subcohorts	[29]
Study cohort: n = 120	Children progressed to T1D; children developed at least a single islet autoantibody but did not progress to T1D during the follow-up; matched controls	Plasma	LC-MS/MS	45	CE 20:5 PC 33:0 TG 54:4 TG 18:2_18:1_18:1 TG 56:5	Children who progress to T1D in the follow-up tend to have a distinct and persistently dysregulated lipid profile as compared to those who later progress to islet autoimmunity but not to T1D	[30]
Study cohort: n = 120	Progressors to T1D (n = 40); children tested positive for at least one antibody in a minimum of two consecutive samples but did not progress to clinical T1D during the follow-up (n = 40); control children remained islet autoantibody-negative during the follow-up (n = 40)	Plasma	GC-MS	94	2-Ketoisocaproic acid 3,4-Dihydroxybutanoic acid Aspartic acid Bisphenol A Glutamic acid Glycerol-2-phosphate Levogluconan Malic acid Methionine Pyruvic acid	Association of unique metabolomic profile with T1D	[31]
Study cohort: n = 2,124	Children with high genetic risk for T1D	Plasma	GC-MS LC-MS/MS	357	5-Methoxytryptamine Alanine Glutamic acid Isoleucine Leucine	Studying autoantibodies and metabolomic markers, which are associated with the risk of progression to T1D	[32]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
					Methionine Proline Valine Vitamin E α -Ketoglutaric acid		
Study cohort: n = 166	T1D patients (n = 85) and healthy controls (n = 81). All patients had a stable dose of insulin usage for more than 3 months (dose change <10%)	Serum Urine	LC-MS/MS	54 (serum) 45 (urine)	4-(2-Aminophenyl)-2,4-dioxobutanoic acid 4-Pyridoxic acid 5-Hydroxytryptophan 5-Methoxyindole-3-acetic acid Hypoxanthine Thromboxane B3	Identification of altered metabolic profiles in T1D individuals with different time in range (TIR)	[33]
Study cohort: n = 286	Infants later developed T1D (n=33); infants developed different numbers of islet autoantibodies during the follow-up (n=110); controls matched for sex, HLA-DQB1 genotype, city of birth, and period of birth (n=143)	Cord blood serum	LC-MS/MS	137	PC 32:0 PC 16:0_16:0 PC 32:1 PC 16:0_16:1 PC 34:1 PC 16:0_18:1 PC 34:3 PC 16:0_18:3 PC 36:1 PC 18:0_18:1 PC 38:3 PC 18:0_20:3 SM d34:1 SM d18:1/16:0 SM d36:1 SM d18:1/18:0 SM d38:0 SM d18:0/20:0 SM d38:1 SM d18:1/20:0 SM d42:1 SM d18:0/24:1 SM d42:2 SM d18:1/24:1 SM d42:2 SM d18:0/24:2 SM d42:3 SM d18:2/24:1	Association with high risk for progression to T1D	[34]
Study cohort: n = 343	Children, who later developed type 1 diabetes (n=166), and random control children in the Norwegian Mother, Father, and Child cohort (n=177)	Cord blood plasma	LC-MS/MS	27	Amino adipic acid Indoxyl sulfate Tryptophan	Association with T1D	[35]
Study cohort: n = 655	Children with high genetic risk for T1D	Plasma	GC-MS	139	Ascorbic acid Piperidone	Association with progression to T1D	[36]
Study cohort: n = 141	Children with T1D (n=76) and gender- and age-matched healthy controls (n=65)	Serum	GC-MS	70	1,5-Anhydroglucitol Adenine Fructose Glycerol- α -phosphate Inosine Levogluconan Pyruvic acid Uridine Xylulose	Association with T1D and with the duration of the disease	[37]
Study cohort: n = 11,896	Participants from four prospective population-based cohorts in Finland (follow-up for 7.8–15 years)	Serum	NMR	229	3-Hydroxybutyric acid Acetic acid Acetoacetic acid Citric acid Creatinine Glutamine Glycerol Glycine Histidine Isoleucine Lactic acid Leucine Phenylalanine Pyruvic acid	- Association with risk of developing diabetes - Association with deterioration in post-load glucose and insulin resistance than with future fasting hyperglycemia	[38]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
					Tyrosine Valine		
Study cohort: n = 1,016	General population	Plasma	NMR	49	3-Hydroxybutyric acid Acetic acid Alanine Citric acid Creatine Creatine phosphate Creatinine Cysteine Glutamine CH ₂ CH ₂ CO- CH ₂ N- Isobutyric acid Isopropanol Leucine N-Acetylglutamine O-Phosphoethanolamine Phenylpropionic acid Proline Pyruvic acid	Strong inverse association of healthy lifestyle with incident T2D	[39]
Study cohort: n = 1,138	Participants from four prospective population-based cohorts	Plasma	LC-MS/MS	70	2-Methylbutyrylcarnitine Cortisol Deoxycholic acid Tyrosine γ-Glutamyl-leucine Barogenin CerPE 38:2 LPC 20:2 MG 18:2 PC 42:7 SM d33:1 SM d34:2 SM d36:3 SM d18:2/18:1	Association with incident T2D	[40]
Study cohorts: n ₁ = 1,261; n ₂ = 2,580	Clinically healthy participants (follow-up for 3 years)	Plasma	LC-MS/MS	N/A	2-Hydroxybutyric acid LPC 18:2	Association with insulin resistance and glucose intolerance	[41]
Study cohort: n = 2,282 Incident T2D cohort: n = 800	General population	Serum	FI-MS	163	Glycine Hexose Phenylalanine LPC 18:2 PC O-34:3 PC O-40:6 PC O-42:5 PC O-44:4 PC O-44:5 PC O-32:1 PC 36:1 PC 38:3 PC 40:5 SM d34:2 SM d18:1/16:1	Association with increased or decreased risk of T2D	[42]
Study cohort: n ₁ = 1,813; n ₂ = 451	1,813 participants without any signs of T2D 451 participants with newly diagnosed T2D	Serum	FI-MS LC-MS/MS	134	Alanine/glycine	Association of alanine/glycine ratio with T2D	[43]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
Study cohort: n = 5844 (90% female)	Female and male nurses	Plasma	LC-MS/MS	186	1-Methylnicotinamide 1-Methylguanosine Aminoisobutyric acid Caffeine CAR 2:0 CAR 5:0 CAR 5:0-DC Cortisone Dimethylglycine Guanidoacetic acid N ² ,N ² -Dimethylguanosine N ⁴ -Acetylcytidine N-Acetylspermidine N-Acetyltryptophan N-Carbamoyl-β-alanine Piperine Ribothymidine Tryptophan Biliverdin Cer d34:1 Cer d18:1/16:0 LPE 18:2 PC 34:2 PC P-34:4 PC P-38:4 PE 36:4 PE P-36:2 SM d38:1 SM d18:1/20:0	Association between inflammatory and insulinemic dietary patterns, plasma inflammatory/insulin biomarkers, plasma metabolomics and risk of type 2 diabetes.	[44]
Study cohort: n = 2240	T2D participants, prediabetes participants, and normal glucose tolerance participants	Serum	FI-MS LC-MS/MS	123	Glycine CAR 16:0 LPC 18:2 PC O-36:0	Association with incident T2D	[45]
Study cohort: n = 4,442 (61% female)	Participants without diabetes at baseline	Plasma	LC-MS/MS	6	Betaine Carnitine Choline Crotonobetaine γ-Butyrobetaine Trimethylamine N-oxide	Association with incident T2D	[46]
Study cohort: n = 1571	Healthy participants (follow-up for 14 years)	Plasma	NMR LC-MS	24	1,5-Anhydroglucitol 2-Hydroxybutyric acid 2-Oxoglutaric acid Glycerol Glycine betaine Isoleucine Lactic acid Methionine Pyruvic acid Tyrosine PC 34:2;O TG 48:0 TG 48:1 TG 50:5	Increase of the long-term prediction performance in combination with classical measurements	[47]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
Discovery cohort: n = 3,821 Validation cohort: n = 14,651	Participants with normal glucose regulation	Serum	LC-MS/MS	667 (discovery cohort) 250 (validation cohort)	CE 14:0 LPI 16:1 PC 34:3 PE 38:4 PE 18:0_20:4 TG 48:1 (16:0) TG 48:1 (16:1) TG 48:2 (16:0) TG 48:2 (16:1) TG 48:2 (18:1) TG 48:3 (16:1) TG 50:0 (18:0) TG 50:1 (16:0) TG 50:1 (16:1) TG 50:1 (18:0) TG 50:2 (16:0) TG 50:2 (16:1) TG 50:2 (16:2) TG 50:2 (18:1) TG 50:3 (16:0) TG 50:3 (16:1) TG 50:3 (16:2) TG 51:0 (17:0) TG 51:2 (17:0) TG 51:3 (17:1) TG 53:2 (19:0) TG 53:3 (16:0) TG 54:3 (16:0) TG 54:4 (16:0) TG 54:4 (16:1) TG 54:5 (16:0) TG 54:5 (16:1) TG 54:6 (20:4) TG 54:7 (20:4) TG 54:7 (22:6) TG 55:6 (19:3) TG 56:5(18:1) TG 56:5(22:4) TG 56:6(22:5)	Association of biomarkers and lipid pathway dysregulation with T2D onset	[48]
Study cohort: n = 2,204 (100% female)	Participants with T2D or impaired fasting glucose + normoglycemic control participants	Plasma Urine	LC-MS/MS GC-MS	447	2-Hydroxybutyric acid 1,5-Anhydroglucitol Arabinose Citrulline Dimethylarginine Erythritol Fructose Glucose Isoleucine Lactic acid Leucine Malic acid Mannose N-Acetylglycine Octanoylcarnitine Proline	Association with incident T2D and IFG	[49]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
					Uric acid Valine 10-Heptadecenoic acid (FA 17:1n7) 15-Methylpalmitic acid (FA iso-17:0) 3-Methyl-2-oxobutanoic acid 3-Methyl-2-oxovaleric acid 4-Methyl-2-oxopentanoic acid 5-Dodecenoic acid (FA 12:1n7) Adrenic acid (FA 22:4n6) Arachidonic acid (FA 20:4n6) Cholesterol Heptanoic acid (FA 7:0) Myristic acid (FA 14:0) Myristoleic acid (FA 14:1n5) Palmitoleic acid (FA 16:1n7) SM d34:1 SM d18:1/16:0 Pelargonic acid (FA 9:0) Pentadecanoic acid (FA 15:0)		
Study cohort: n = 1,150	Participants with normal fasting glucose (follow-up for 20 years)	Plasma	LC-MS/MS	N/A	5-Hydroxyindoleacetic acid Glucose Glycine Isocitric acid Phenylalanine Taurine 2-Aminodipic acid 3-Methyladipic acid CE 20:3 DG 36:1 LPC 18:1 LPC 18:2 PC 36:4 SM d42:1 SM d18:1/24:0 TG 48:0 TG 48:1 TG 52:1 TG 54:8 TG 58:11	Association with improved prediction of T2D beyond conventional risk factors	[50]
Discovery cohort: n = 543 Validation cohort: n = 1,044	Non-diabetic participants (follow-up)	Serum	LC-MS/MS GC-MS	568	2-Hydroxybutyric acid Bilirubin Glucose Glutamic acid Glutamine Histidine Isoleucine Mannose Trehalose Valine α-Tocopherol	Association with positive or negative impact on progression to T2D	[51]
Study cohort: n = 1,248	Participants with 6.5 years follow-up	Plasma	DMS-MS/MS GC-MS	N/A	<i>Lipid classes containing species with FA 15:0 and FA 17:0:</i> CE 15:0 CE 17:0 DG 15:0 FA 15:0	Association with incident T2D	[52]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
					FA 17:0 LPC 15:0 LPC 17:0 LPE 17:0 MG 15:0 MG 17:0 PC 15:0 PC 17:0 PE 17:0 PL-OCFA (phospholipid species containing odd-chain fatty acids) TG 15:0 TG 17:0		
Study cohorts: n ₁ = 1,039; n ₂ = 520	Participants with mean follow-ups: 4.61 and 7.57 years	Plasma	LC-MS/MS	166	CE 16:1 LPC 15:0 LPC 18:2 PC 33:3 PC 35:3 PC 40:7 PC 43:6 PC 44:1 SM d34:2 SM d41:2 TG 46:1 (12:0) TG 48:1 (16:0) TG 48:2 (14:0) TG 49:7 (16:0) TG 50:1 (16:0) TG 50:2 (16:0) TG 50:3 (18:1) TG 51:7 (16:0) TG 52:5 (18:2) TG 52:6 (18:2) TG 54:3 (18:0) TG 54:4 (18:2) TG 54:5 (18:2) TG 54:6 (18:2) TG 54:7 (18:3) TG 56:5 (20:4)	Association with incident T2D	[53]
Study cohort: n = 2,939	Participants without diabetes prevalence	Serum	LC-MS/MS	245	3-(4-Hydroxyphenyl)lactic acid Asparagine Erythritol Isoleucine Leucine Trehalose Valine	Association with incident T2D (protective biomarker of diabetes risk)	[54]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
Study cohort: n = 2,103	Participants with a 6-year follow-up	Plasma	LC-MS/MS	34	Carnitine 3-Dehydroxycarnitine 3-Dehydrocarnitine CAR 2:0 CAR 3:0 CAR 3:0-DC CAR 4:0 CAR 5:0 CAR 5:0-OH CAR 5:1 CAR 6:0 CAR 6:0-OH CAR 6:0-DC CAR 7:0-DC CAR 8:0 CAR 8:1 CAR 10:0 CAR 10:0-DC CAR 12:0 CAR 12:0-OH CAR 12:1 CAR 12:0-DC CAR 14:0 CAR 14:0-OH CAR 14:1-OH CAR 16:0 CAR 16:1 CAR 16:2 CAR 18:0 CAR 18:0-OH CAR 18:1 CAR 18:2 CAR 20:0 CAR 20:4	Association with improved predictive ability for type 2 diabetes beyond conventional risk factors	[55]
Study cohort: n = 3,234	Participants were assigned to 1) intensive lifestyle, 2) metformin, or 3) placebo (all followed up for 3.2 years)	Plasma	HILIC-MS/MS	84	Betaine Methionine sulfoxide Serine	Association with incident T2D	[56]

Table S3. Metabolomics and lipidomics cohort studies focused on metabolic dysfunction-associated steatotic liver disease

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
Study cohort: n = 121,032	Participants with a mean 12.6-year follow-up	Plasma	NMR	170	3-Hydroxybutyric acid Acetic acid Acetoacetatic acid Acetone Alanine Citric acid Creatinine Glucose Glutamine Glycine Histidine Isoleucine Lactic acid Leucine Phenylalanine Pyruvic acid Tyrosine Valine Docosahexaenoic acid (FA 22:6) Linoleic acid (FA 18:2) Omega-3 FA Omega-6 FA	Positive and negative association with MASLD	[57]
Study cohort: n = 10,809	Participants with and without MASLD	Plasma	NMR	123	Tyrosine	Association with MASLD	[58]
Study cohort: n = 3,048	Participants have been followed up since birth, including questionnaires and clinical assessments starting from age 7 years	Plasma	NMR	154	3-Hydroxybutyric acid Acetic acid Acetoacetatic acid Alanine Creatinine Glutamine Histidine Isoleucine Leucine Phenylalanine Tyrosine Valine	Association with incident MASLD	[59]
Study cohort: n = 928 (67% female)	Participants with and without MASLD	Plasma	CE-MS	94	4-Methyl-2-oxopentanoic acid Alanine Glutamic acid Isoleucine Leucine Proline Tryptophan Tyrosine Valine Glycerophosphorylcholine	Association with both MASLD and cardio-ankle vascular index (CAVI)	[60]
Study cohort: n = 1,479 Study subcohort: n = 447 (known age)	Participants were not treated for cancer or infectious disease or had undergone surgery in the previous year, and they had no history of cancer or an infectious disease.	Serum	LC-MS/MS	N/A	Oleic acid-hydroxy oleic acid (OAHOA) Sphingosine Uric acid	Association with MASLD	[61]
Study cohort: n = 997 (53% female)	Participants free of prevalent myocardial infarction or congestive heart failure at the first examination cycle	Plasma	HILIC-MS/MS	179	Anandamide	Association with MASLD severity, the presence of nonalcoholic steatohepatitis, and fibrosis	[62]

Subjects (n)	Cohort	Matrix	Platforms	Reported metabolites (n)	Markers	Outcomes	Ref.
Study cohort: n = 559	Participants with and without MASLD	Plasma	LC-MS/MS	11	Dihydrothymine Serine Tryptophan LPC 18:1 LPE 20:0	• Screening tool for MASLD	[63]
Study cohort: n = 1,154 (50% female) Control cohort: n = 350	Participants with biopsy-proven MASLD and participants from the general population with similar gender and age to the cohort of patients with MASLD	Serum	LC-MS NMR	105	PC 32:0 PC 16:0_16:0 PC 32:2 PC 14:0_18:2 PC 34:2 PC 16:0_18:2 PC 36:1 PC 18:0_18:1 PC 36:3 PC 36:6 PC 18:3_18:3 PC 37:5 PC 38:2 PC 20:0_18:2 PC 38:3 PC 18:0_20:3 SM d32:1 SM d39:1 TG 48:3	• Identification of three MASLD subgroups, independent of histological disease severity	[64]
Study cohort: n = 627	Histologically characterized participants. Participants include the full spectrum of disease, from histologically normal liver tissue through NAFL to NASH-F4 (cirrhosis)	Serum	LC-MS/MS GC-MS/MS	211	<i>Markers of fibrosis 0–1 vs. 2–4:</i> 2-Hydroxybutyric acid 3-Hydroxybutyric acid LPC O-16:0 LPC P-16:0 LPC 18:2 LPC 20:4 Oleic acid PC 32:0 PC 16:0/16:0 PC 32:1 PC 37:4 PC O-34:2 PC O-34:3 PE 16:0/18:1 PE 34:2 PE 38:6 SM d42:1 SM d18:1/24:0 SM d36:0 SM d41:1 TG 56:4 TG 58:6	• Identification of a key metabolic ‘watershed’ in the progression of liver damage, separating severe disease from mild	[65]
Discovery cohort: n = 1,546 Internal validation cohort: n = 377 Prospective validation cohort: n = 749	Participants with and without MASLD (4 years follow-up)	Feces	LC-MS/MS	198	Taurocholic acid	• Positive association with both a higher microbiome risk score and MASLD risk	[66]

References

1. Akbaraly T, Würtz P, Singh-Manoux A, Shipley MJ, Haapakoski R, Lehto M, Desrumaux C, Kähönen M, Lehtimäki T, Mikkilä V, ET AL. Association of circulating metabolites with healthy diet and risk of cardiovascular disease: analysis of two cohort studies. *Sci Rep* 2018;8:8620. <https://doi.org/10.1038/s41598-018-26441-1>
2. Benson TW, Conrad KA, Li XS, Wang Z, Helsley RN, Schugar RC, Coughlin TM, Wadding-Lee C, Fleifel S, Russell HM, ET AL. Gut microbiota-derived trimethylamine N-oxide contributes to abdominal aortic aneurysm through inflammatory and apoptotic mechanisms. *Circulation* 2023;147:1079-1096. <https://doi.org/10.1161/CIRCULATIONAHA.122.060573>
3. Dehghanbanadaki H, Dodangeh S, Parhizkar Roudsari P, Hosseinkhani S, Khashayar P, Noorchenarboo M, Rezaei N, Dilmaghani-Marand A, Yoosefi M, Arjmand B, ET AL. Metabolomics profile and 10-year atherosclerotic cardiovascular disease (ASCVD) risk score. *Front Cardiovasc Med* 2023;10:<https://doi.org/10.3389/fcvm.2023.1161761>
4. Eichelmann F, Sellem L, Wittenbecher C, Jäger S, Kuxhaus O, Prada M, Cuadrat R, Jackson KG, Lovegrove JA, Schulze MB. Deep lipidomics in human plasma: Cardiometabolic disease risk and effect of dietary fat modulation. *Circulation* 2022;146:21-35. <https://doi.org/10.1161/circulationaha.121.056805>
5. Ferrell M, Wang Z, Anderson JT, Li XS, Witkowski M, DiDonato JA, Hilser JR, Hartiala JA, Haghikia A, Cajka T, ET AL. A terminal metabolite of niacin promotes vascular inflammation and contributes to cardiovascular disease risk. *Nat Med* 2024;30:424-434. <https://doi.org/10.1038/s41591-023-02793-8>
6. Fu Z, Liu Q, Liang J, Weng Z, Li W, Xu J, Zhang X, Xu C, Gu A. Association between NMR metabolomic signatures of healthy lifestyle and incident coronary artery disease. *Eur J Prev Cardiol* 2022;30:243-253. <https://doi.org/10.1093/eurjpc/zwac252>
7. Ganna A, Salihovic S, Sundström J, Broeckling CD, Hedman ÅK, Magnusson PKE, Pedersen NL, Larsson A, Siegbahn A, Zilmer M, ET AL. Large-scale metabolomic profiling identifies novel biomarkers for incident coronary heart disease. *PLOS Genet* 2014;10:e1004801. <https://doi.org/10.1371/journal.pgen.1004801>
8. Guasch-Ferré M, Hernández-Alonso P, Drouin-Chartier JP, Ruiz-Canela M, Razquin C, Toledo E, Li J, Dennis C, Wittenbecher C, Corella D, ET AL. Walnut consumption, plasma metabolomics, and risk of type 2 diabetes and cardiovascular disease. *J Nutr* 2021;151:303-311. <https://doi.org/10.1093/jn/nxaa374>
9. Harm T, Dittrich K, Brun A, Fu X, Frey M, Petersen Uribe A, Schwarz F-J, Rohlfing A-K, Castor T, Geisler T, ET AL. Large-scale lipidomics profiling reveals characteristic lipid signatures associated with an increased cardiovascular risk. *Clin Res Cardiol* 2023;112:1664-1678. <https://doi.org/10.1007/s00392-023-02260-x>
10. Huang Z, Klaric L, Krasauskaite J, Khalid W, Strachan MWJ, Wilson JF, Price JF. Combining serum metabolomic profiles with traditional risk factors improves 10-year cardiovascular risk prediction in people with type 2 diabetes. *Eur J Prev Cardiol* 2023;30:1255-1262. <https://doi.org/10.1093/eurjpc/zwad160>
11. Lauber C, Gerl MJ, Klose C, Ottosson F, Melander O, Simons K. Lipidomic risk scores are independent of polygenic risk scores and can predict incidence of diabetes and cardiovascular disease in a large population cohort. *PLOS Biol* 2022;20:e3001561. <https://doi.org/10.1371/journal.pbio.3001561>
12. Li XS, Wang Z, Cajka T, Buffa JA, Nemet I, Hurd AG, Gu X, Skye SM, Roberts AB, Wu Y, ET AL. Untargeted metabolomics identifies trimethyllysine, a TMAO-producing nutrient precursor, as a predictor of incident cardiovascular disease risk. *JCI Insight* 2018;3:<https://doi.org/10.1172/jci.insight.99096>
13. Lind L, Fall T, Ärnlöv J, Elmståhl S, Sundström J. Large-scale metabolomics and the incidence of cardiovascular disease. *J Am Heart Assoc* 2023;12:e026885. <https://doi.org/10.1161/JAHA.122.026885>
14. Lu Q, Chen J, Li R, Wang Y, Tu Z, Geng T, Liu L, Pan A, Liu G. Healthy lifestyle, plasma metabolites, and risk of cardiovascular disease among individuals with diabetes. *Atherosclerosis* 2023;367:48-55. <https://doi.org/10.1016/j.atherosclerosis.2022.12.008>
15. Margara-Escudero HJ, Paz-Graniel I, García-Gavilán J, Ruiz-Canela M, Sun Q, Clish CB, Toledo E, Corella D, Estruch R, Ros E, ET AL. Plasma metabolite profile of legume consumption and future risk of type 2 diabetes and cardiovascular disease. *Cardiovasc Diabetol* 2024;23:38. <https://doi.org/10.1186/s12933-023-02111-z>

16. Mundra PA, Barlow CK, Nestel PJ, Barnes EH, Kirby A, Thompson P, Sullivan DR, Alshehry ZH, Mellett NA, Huynh K, ET AL. Large-scale plasma lipidomic profiling identifies lipids that predict cardiovascular events in secondary prevention. *JCI Insight* 2018;3: <https://doi.org/10.1172/jci.insight.121326>
17. Nemet I, Saha PP, Gupta N, Zhu W, Romano KA, Skye SM, Cajka T, Mohan ML, Li L, Wu Y, ET AL. A cardiovascular disease-linked gut microbial metabolite acts via adrenergic receptors. *Cell* 2020;180:862-877.e22. <https://doi.org/10.1016/j.cell.2020.02.016>
18. Nemet I, Funabashi M, Li XS, Dwidar M, Sangwan N, Skye SM, Romano KA, Cajka T, Needham BD, Mazmanian SK, ET AL. Microbe-derived uremic solutes enhance thrombosis potential in the host. *mBio* 2023;14:e0133123. <https://doi.org/10.1128/mbio.01331-23>
19. Seah JYH, Chew WS, Torta F, Khoo CM, Wenk MR, Herr DR, Choi H, Tai ES, van Dam RM. Plasma sphingolipids and risk of cardiovascular diseases: a large-scale lipidomic analysis. *Metabolomics* 2020;16:89. <https://doi.org/10.1007/s11306-020-01709-8>
20. Tang WHW, Wang Z, Levison BS, Koeth RA, Britt EB, Fu X, Wu Y, Hazen SL. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. *N Engl J Med* 2013;368:1575-1584. <https://doi.org/10.1056/NEJMoa1109400>
21. Tzoulaki I, Castagné R, Boulangé CL, Karaman I, Chekmeneva E, Evangelou E, Ebbels TMD, Kaluarachchi MR, Chadeau-Hyam M, Mosen D, ET AL. Serum metabolic signatures of coronary and carotid atherosclerosis and subsequent cardiovascular disease. *Eur Heart J* 2019;40:2883-2896. <https://doi.org/10.1093/eurheartj/ehz235>
22. Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, DuGar B, Feldstein AE, Britt EB, Fu X, Chung Y-M, ET AL. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature* 2011;472:57-63. <https://doi.org/10.1038/nature09922>
23. Witkowski M, Nemet I, Alamri H, Wilcox J, Gupta N, Nimer N, Haghikia A, Li XS, Wu Y, Saha PP, ET AL. The artificial sweetener erythritol and cardiovascular event risk. *Nat Med* 2023;29:710-718. <https://doi.org/10.1038/s41591-023-02223-9>
24. Witkowski M, Nemet I, Li XS, Wilcox J, Ferrell M, Alamri H, Gupta N, Wang Z, Tang WHW, Hazen SL. Xylitol is prothrombotic and associated with cardiovascular risk. *Eur Heart J* 2024; <https://doi.org/10.1093/eurheartj/ehae244>
25. Würtz P, Havulinna AS, Soininen P, Tynkkynen T, Prieto-Merino D, Tillin T, Ghorbani A, Artati A, Wang Q, Tiainen M, ET AL. Metabolite profiling and cardiovascular event risk: A prospective study of 3 population-based cohorts. *Circulation* 2015;131:774-85. <https://doi.org/10.1161/circulationaha.114.013116>
26. Zhu W, Gregory JC, Org E, Buffa JA, Gupta N, Wang Z, Li L, Fu X, Wu Y, Mehrabian M, ET AL. Gut microbial metabolite TMAO enhances platelet hyperreactivity and thrombosis risk. *Cell* 2016;165:111-124. <https://doi.org/10.1016/j.cell.2016.02.011>
27. Balzano-Nogueira L, Ramirez R, Zamkovaya T, Dailey J, Ardisson AN, Chamala S, Serrano-Quílez J, Rubio T, Haller MJ, Concannon P, ET AL. Integrative analyses of TEDDY Omics data reveal lipid metabolism abnormalities, increased intracellular ROS and heightened inflammation prior to autoimmunity for type 1 diabetes. *Genome Biol* 2021;22:39. <https://doi.org/10.1186/s13059-021-02262-w>
28. La Torre D, Seppänen-Laakso T, Larsson HE, Hyötyläinen T, Ivarsson SA, Lernmark Å, Orešič M, Group atDS. Decreased cord-blood phospholipids in young age-at-onset type 1 diabetes. *Diabetes* 2013;62:3951-3956. <https://doi.org/10.2337/db13-0215>
29. Lamichhane S, Ahonen L, Sparholt Dyrland T, Dickens AM, Siljander H, Hyöty H, Ilonen J, Toppari J, Veijola R, Hyötyläinen T, ET AL. Cord-blood lipidome in progression to islet autoimmunity and type 1 diabetes. *Biomolecules* 2019;9:33. <https://doi.org/10.3390/biom9010033>
30. Lamichhane S, Ahonen L, Dyrland TS, Kemppainen E, Siljander H, Hyöty H, Ilonen J, Toppari J, Veijola R, Hyötyläinen T, ET AL. Dynamics of plasma lipidome in progression to islet autoimmunity and type 1 diabetes – Type 1 diabetes prediction and prevention study (DIPP). *Sci Rep* 2018;8:10635. <https://doi.org/10.1038/s41598-018-28907-8>

31. Lamichhane S, Kempainen E, Trošt K, Siljander H, Hyöty H, Ilonen J, Toppari J, Veijola R, Hyötyläinen T, Knip M, ET AL. Circulating metabolites in progression to islet autoimmunity and type 1 diabetes. *Diabetologia* 2019;62:2287-2297. <https://doi.org/10.1007/s00125-019-04980-0>
32. Li Q, Parikh H, Butterworth MD, Lernmark Å, Hagopian W, Rewers M, She J-X, Toppari J, Ziegler A-G, Akolkar B, ET AL. Longitudinal metabolome-wide signals prior to the appearance of a first islet autoantibody in children participating in the TEDDY study. *Diabetes* 2020;69:465-476. <https://doi.org/10.2337/db19-0756>
33. Ma L, Liu J, Deng M, Zhou L, Zhang Q, Xiao X. Metabolomics analysis of serum and urine in type 1 diabetes patients with different time in range derived from continuous glucose monitoring. *Diabetol Metab Syndr* 2024;16:21. <https://doi.org/10.1186/s13098-024-01257-4>
34. Oresic M, Gopalacharyulu P, Mykkänen J, Lietzen N, Mäkinen M, Nygren H, Simell S, Simell V, Hyöty H, Veijola R, ET AL. Cord serum lipidome in prediction of islet autoimmunity and type 1 diabetes. *Diabetes* 2013;62:3268-74. <https://doi.org/10.2337%2Fdb13-0159>
35. Tapia G, Suvaivaal T, Ahonen L, Lund-Blix NA, Njølstad PR, Joner G, Skriverhaug T, Legido-Quigley C, Størdal K, Stene LC. Prediction of type 1 diabetes at birth: Cord blood metabolites vs genetic risk score in the Norwegian Mother, Father, and Child cohort. *J Clin Endocrinol Metab* 2021;106:e4062-e4071. <https://doi.org/10.1210/clinem/dgab400>
36. Webb-Robertson B-JM, Nakayasu ES, Frohnert BI, Bramer LM, Akers SM, Norris JM, Vehik K, Ziegler A-G, Metz TO, Rich SS, ET AL. Integration of infant metabolite, genetic, and islet autoimmunity signatures to predict type 1 diabetes by age 6 years. *J Clin Endocrinol Metab* 2022;107:2329-2338. [10.1210/clinem/dgac225](https://doi.org/10.1210/clinem/dgac225)
37. Zhang J, Wu W, Huang K, Dong G, Chen X, Xu C, Ni Y, Fu J. Untargeted metabolomics reveals gender- and age- independent metabolic changes of type 1 diabetes in Chinese children. *Front Endocrinol* 2022;13:1-11. <https://doi.org/10.3389/fendo.2022.1037289>
38. Ahola-Olli AV, Mustelin L, Kalimeri M, Kettunen J, Jokelainen J, Auvinen J, Puukka K, Havulinna AS, Lehtimäki T, Kähönen M, ET AL. Circulating metabolites and the risk of type 2 diabetes: a prospective study of 11,896 young adults from four Finnish cohorts. *Diabetologia* 2019;62:2298-2309. <https://doi.org/10.1007/s00125-019-05001-w>
39. Delgado-Velandia M, Gonzalez-Marrachelli V, Domingo-Relloso A, Galvez-Fernandez M, Grau-Perez M, Olmedo P, Galan I, Rodriguez-Artalejo F, Amigo N, Briongos-Figuero L, ET AL. Healthy lifestyle, metabolomics and incident type 2 diabetes in a population-based cohort from Spain. *Int J Behav Nutr Phys Act* 2022;19:8. <https://doi.org/10.1186/s12966-021-01219-3>
40. Fall T, Salihovic S, Brandmaier S, Nowak C, Ganna A, Gustafsson S, Broeckling CD, Prenni JE, Kastenmüller G, Peters A, ET AL. Non-targeted metabolomics combined with genetic analyses identifies bile acid synthesis and phospholipid metabolism as being associated with incident type 2 diabetes. *Diabetologia* 2016;59:2114-2124. <https://doi.org/10.1007/s00125-016-4041-1>
41. Ferrannini E, Natali A, Camastra S, Nannipieri M, Mari A, Adam KP, Milburn MV, Kastenmüller G, Adamski J, Tuomi T, ET AL. Early metabolic markers of the development of dysglycemia and type 2 diabetes and their physiological significance. *Diabetes* 2013;62:1730-7. <https://doi.org/10.2337/db12-0707>
42. Floegel A, Stefan N, Yu Z, Mühlenbruch K, Drogan D, Joost HG, Fritsche A, Häring HU, Hrabě de Angelis M, Peters A, ET AL. Identification of serum metabolites associated with risk of type 2 diabetes using a targeted metabolomic approach. *Diabetes* 2013;62:639-48. <https://doi.org/10.2337/db12-0495>
43. Lee KS, Lee YH, Lee SG. Alanine to glycine ratio is a novel predictive biomarker for type 2 diabetes mellitus. *Diabetes Obes Metab* 2024;26:980-988. <https://doi.org/10.1111/dom.15395>
44. Lee DH, Jin Q, Shi N, Wang F, Bever AM, Liang L, Hu FB, Song M, Zeleznik OA, Zhang X, ET AL. The metabolic potential of inflammatory and insulinaemic dietary patterns and risk of type 2 diabetes. *Diabetologia* 2024;67:88-101. <https://doi.org/10.1007/s00125-023-06021-3>
45. Lee H-S, Xu T, Lee Y, Kim N-H, Kim Y-J, Kim J-M, Cho SY, Kim K-Y, Nam M, Adamski J, ET AL. Identification of putative biomarkers for type 2 diabetes using metabolomics in the Korea Association REsource (KARE) cohort. *Metabolomics* 2016;12:178. <https://doi.org/10.1007/s11306-016-1103-9>

46. Lemaitre RN, Jensen PN, Wang Z, Fretts AM, McKnight B, Nemet I, Biggs ML, Sotoodehnia N, de Oliveira Otto MC, Psaty BM, ET AL. Association of Trimethylamine N-Oxide and Related Metabolites in Plasma and Incident Type 2 Diabetes: The Cardiovascular Health Study. *JAMA Netw Open* 2021;4:e2122844-e2122844. <https://doi.org/10.1001/jamanetworkopen.2021.22844>
47. Liu J, Semiz S, van der Lee SJ, van der Spek A, Verhoeven A, van Klinken JB, Sijbrands E, Harms AC, Hankemeier T, van Dijk KW, ET AL. Metabolomics based markers predict type 2 diabetes in a 14-year follow-up study. *Metabolomics* 2017;13:104. <https://doi.org/10.1007/s11306-017-1239-2>
48. Lu J, Lam SM, Wan Q, Shi L, Huo Y, Chen L, Tang X, Li B, Wu X, Peng K, ET AL. High-coverage targeted lipidomics reveals novel serum lipid predictors and lipid pathway dysregulation antecedent to type 2 diabetes onset in normoglycemic chinese adults. *Diabetes Care* 2019;42:2117-2126. <https://doi.org/10.2337/dc19-0100>
49. Menni C, Fauman E, Erte I, Perry JR, Kastenmüller G, Shin SY, Petersen AK, Hyde C, Psatha M, Ward KJ, ET AL. Biomarkers for type 2 diabetes and impaired fasting glucose using a nontargeted metabolomics approach. *Diabetes* 2013;62:4270-6. <https://doi.org/10.2337/db13-0570>
50. Merino J, Leong A, Liu C-T, Porneala B, Walford GA, von Grotthuss M, Wang TJ, Flannick J, Dupuis J, Levy D, ET AL. Metabolomics insights into early type 2 diabetes pathogenesis and detection in individuals with normal fasting glucose. *Diabetologia* 2018;61:1315-1324. <https://doi.org/10.1007/s00125-018-4599-x>
51. Peddinti G, Cobb J, Yengo L, Froguel P, Kravić J, Balkau B, Tuomi T, Aittokallio T, Groop L. Early metabolic markers identify potential targets for the prevention of type 2 diabetes. *Diabetologia* 2017;60:1740-1750. <https://doi.org/10.1007/s00125-017-4325-0>
52. Prada M, Wittenbecher C, Eichelmann F, Wernitz A, Drouin-Chartier J-P, Schulze MB. Association of the odd-chain fatty acid content in lipid groups with type 2 diabetes risk: A targeted analysis of lipidomics data in the EPIC-Potsdam cohort. *Clin Nutr* 2021;40:4988-4999. <https://doi.org/10.1016/j.clnu.2021.06.006>
53. Qiu G, Wang H, Yan Q, Ma H, Niu R, Lei Y, Xiao Y, Zhou L, Yang H, Xu C, ET AL. A lipid signature with perturbed triacylglycerol co-regulation, identified from targeted lipidomics, predicts risk for type 2 diabetes and mediates the risk from adiposity in two prospective cohorts of chinese adults. *Clin Chem* 2022;68:1094-1107. <https://doi.org/10.1093/clinchem/hvac090>
54. Rebholz CM, Yu B, Zheng Z, Chang P, Tin A, Köttgen A, Wagenknecht LE, Coresh J, Boerwinkle E, Selvin E. Serum metabolomic profile of incident diabetes. *Diabetologia* 2018;61:1046-1054. <https://doi.org/10.1007/s00125-018-4573-7>
55. Sun L, Liang L, Gao X, Zhang H, Yao P, Hu Y, Ma Y, Wang F, Jin Q, Li H, ET AL. Early prediction of developing type 2 diabetes by plasma acylcarnitines: A population-based study. *Diabetes Care* 2016;39:1563-1570. <https://doi.org/10.2337/dc16-0232>
56. Walford GA, Ma Y, Clish C, Florez JC, Wang TJ, Gerszten RE. Metabolite profiles of diabetes incidence and intervention response in the diabetes prevention program. *Diabetes* 2016;65:1424-33. <https://doi.org/10.2337/db15-1063>
57. Gagnon E, Manikpurage HD, Mitchell PL, Girard A, Gobeil É, Bourgault J, Bégin F, Marette A, Thériault S, Arsenault BJ. Large-scale metabolomic profiling and incident non-alcoholic fatty liver disease. *iScience* 2023;26:107127. <https://doi.org/10.1016/j.isci.2023.107127>
58. Gobeil É, Maltais-Payette I, Taba N, Brière F, Ghodsian N, Abner E, Bourgault J, Gagnon E, Manikpurage HD, Couture C, ET AL. Mendelian randomization analysis identifies blood tyrosine levels as a biomarker of non-alcoholic fatty liver disease. *Metabolites* 2022;12:440. <https://doi.org/10.3390/metabo12050440>
59. Hartley A, Santos Ferreira DL, Anderson EL, Lawlor DA. Metabolic profiling of adolescent non-alcoholic fatty liver disease [version 2; peer review: 2 approved]. *Wellcome Open Res* 2019;3:166. <https://doi.org/10.12688/wellcomeopenres.14974.2>
60. Hirata A, Harada S, Iida M, Kurihara A, Fukai K, Kuwabara K, Kato S, Matsumoto M, Sata M, Miyagawa N, ET AL. Association of Nonalcoholic Fatty Liver Disease with Arterial Stiffness and its Metabolomic Profiling in Japanese Community-Dwellers. *J Atheroscler Thromb* 2024;31:1-17. <http://doi.org/10.5551/jat.64616>

61. Hu X-Y, Li Y, Li L-Q, Zheng Y, Lv J-H, Huang S-C, Zhang W, Liu L, Zhao L, Liu Z, ET AL. Risk factors and biomarkers of non-alcoholic fatty liver disease: an observational cross-sectional population survey. *BMJ Open* 2018;8:e019974. <http://dx.doi.org/10.1136/bmjopen-2017-019974>
62. Kimberly WT, O'Sullivan JF, Nath AK, Keyes M, Shi X, Larson MG, Yang Q, Long MT, Vasan R, Peterson RT, ET AL. Metabolite profiling identifies anandamide as a biomarker of nonalcoholic steatohepatitis. *JCI Insight* 2017;2:<https://doi.org/10.1172/jci.insight.92989>
63. Khusial R, Cioffi C, Caltharp S, Krasinskas A, Alazraki A, Knight-Scott J, Cleeton R, Castillo-Leon E, Jones D, Pierpont B, ET AL. Development of a plasma screening panel for pediatric nonalcoholic fatty liver disease using metabolomics. *Hepatol Commun* 2019;3:<https://doi.org/10.1002/hep4.1417>
64. Martínez-Arranz I, Bruzzzone C, Nouredin M, Gil-Redondo R, Mincholé I, Bizkarguenaga M, Arretxe E, Iruarizaga-Lejarreta M, Fernández-Ramos D, Lopitz-Otsoa F, ET AL. Metabolic subtypes of patients with NAFLD exhibit distinctive cardiovascular risk profiles. *Hepatology* 2022;76:1121-1134. <https://doi.org/10.1002/hep.32427>
65. McGlinchey AJ, Govaere O, Geng D, Ratzu V, Allison M, Bousier J, Petta S, de Oliveira C, Bugianesi E, Schattenberg JM, ET AL. Metabolic signatures across the full spectrum of non-alcoholic fatty liver disease. *JHEP Rep* 2022;4:100477. <https://doi.org/10.1016/j.jhepr.2022.100477>
66. Zeng F, Su X, Liang X, Liao M, Zhong H, Xu J, Gou W, Zhang X, Shen L, Zheng J-S, ET AL. Gut microbiome features and metabolites in non-alcoholic fatty liver disease among community-dwelling middle-aged and older adults. *BMC Med* 2024;22:104. <https://doi.org/10.1186/s12916-024-03317-y>