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Liver glucocorticoid receptor and heat shock protein 70 levels in rats exposed to different stress models

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Summary

The aim of the present study was to define the stress induced pattern of cytosolic glucocorticoid receptor (GR) and Hsp70 protein in the liver of male Wistar rats exposed to different stress models: acute (2h/day) immobilization or cold (4°C); chronic (21days) isolation, crowding, swimming or isolation plus swimming and combined (chronic plus acute stress). Changes in plasma levels of corticosterone were studied by radioimmunoassay (RIA). The results obtained by Western immunoblotting showed that both acute stressors led to a significant decrease in cytosolic GR and Hsp70 levels. Compared to acute stress effects, chronic stress models led to a weakly decrease in the levels of GR and Hsp70. Chronically stressed rats, which were subsequently exposed to novel acute stressors (immobilization or cold), showed a lower extent of GR down-regulation when compared to acute stress. The exception was swimming, which partially restores this down-regulation. The observed changes in the levels of these, major stress related cellular proteins in liver cytosol lead to the conclusion that, chronic stressors compromise intracellular GR down-regulation in liver.

Key words: Rat, Liver, Stress, Glucocorticoid receptor, Heat shock protein 70

Introduction

Glucocorticoids (GCs) are involved in the regulation of various physiological processes that are essential for the maintenance of vital functions. These hormones are necessary for normal growth and development, liver and immune functions as well as in mediating stress responses (Sapolsky *et al.* 2000). The biological actions of GCs are dependent upon tissue-specific levels of the glucocorticoid receptor (GR). GR exists in the cytoplasm of target cells in the form of multiprotein complexes containing several heat shock proteins (Hsps) and additional components of the molecular chaperon family (Pratt and Toft 1997). Upon steroid binding, the GR is translocated to the cell nucleus where it acts as a ligand induced transcription factor (Caamano *et al.* 2001). The steroid receptors are associated predominantly with Hsp70 (Hightower and Hendershot 1997), which is the chaperoning role in protein folding (Hendrick and Hartl 1993) and nuclear import (Shi and Thomas 1992). Also, the expression of Hsp70 is related to the preservation and recovery of cellular proteins under stress (Benjamin and McMillan 1998).

Tissue sensitivity to hormone signals is directly related to the levels of circulating cortisol and the number of GR found in cells (Oakley and Cidlowski 1993, Dong *et al.* 1988). In addition, it has been shown that animals chronically exposed to a particular stressor, display an exaggerated response of the hypothalamo-pituitary adrenal (HPA) axis to various novel stressors, judged by plasma levels of the adenocorticotrophic hormone and corticosterone (CORT) (McEwen 1998, Garcia *et al.* 2000, Sapolsky *et al.* 1984). The exaggerated effect of novel stressors is an interesting phenomenon that has not been studied at the levels of GR and Hsp70 in liver. Recently, we have shown that significant down-regulation of GR levels in acute stress was almost completely compromised by chronic stressors alone, or in combination with novel acute stressors, in higher centres of the HPA axis *i.e.* brain cortex and hippocampus (Filipović *et al.* 2005). Since the liver is a well studied target for GCs, we have chosen this organ to examine the

effects of different stress models (Filipović *et al.* 2005) on the levels of GR in the liver cytosol and possibly reveal if these stress models influence the intracellular cytosolic GR regulation. Considering that GCs stimulate the synthesis of heat shock proteins under basal conditions (Kasambalides and Lanks 1983), it was of interest to investigate if these hormones might influence the levels of Hsp70 during the various stress models applied in this investigation. Therefore, the present study was conducted to define the stress induced pattern of GR and Hsp70 protein in liver cytosol of adult Wistar male rats exposed to acute (immobilization or cold), chronic (isolation, crowding, swimming or isolation plus swimming) or combined (chronic plus acute) stress models. Plasma CORT levels were also monitored. It was expected that patterns of cytosolic GR and Hsp70 in the liver of stressed rats could be one of the cytosolic parameters that may serve to help make a distinction between stress duration (acute, chronic), stress intensity (by protein expression level) or type (psychosocial, physical).

Materials and methods

Animals and stress models

Adult Wistar rat males, weighing 330-400g were housed in groups of four individuals *per* cage and offered water and food (commercial rat pellets) *ad libitum*. The light was kept on between 7.00 a.m. and 7.00 p.m., and the room temperature was 20±2°C. The animal experiments were approved by the Ethical Committee for the Use of Laboratory Animals of the “Vinča” Institute which works according to the guide lines of the registered “Serbian Society for the Use of Animals in Research and Education”. For experimental purposes, the animals were randomly divided into ten groups. Each group consisted of 6-8 animals. The Group I consisted of the unstressed animals (control group); Group II consisted of of animals exposed to acute stress by 2h of immobilization (IIa) or cold (IIb) (4°C); Group III was exposed to chronic isolation *i.e.* the

rats were individually housed for 21 days; Group IV was exposed to chronic crowding for 21 days, with eight animals *per* cage; Group V comprising four animals *per* cage was exposed to chronic forced 15 minutes/daily swimming for 21 days, Group VI was exposed to chronic isolation plus 15 minutes/daily swimming for 21 days. Animals exposed to either of the four chronic stressors (groups III-VI) were subsequently exposed to immobilization or cold (4°C) for 2h (isolation plus immobilization or cold VIIa/VIIb, crowding plus immobilization or cold VIIIa/VIIIb, swimming plus immobilization or cold IXa/IXb, isolation/swimming plus immobilization or cold Xa/Xb). Immobilization stress was induced as described by Kvetnansky and Mikulaj (1970). The animals exposed to cold were initially kept at ambient temperature (20±2°C) and then carefully transferred into a cold chamber at 4°C with lights on.

Preparation of rat blood plasma and liver cell extracts

The blood sampling for determining plasma corticosterone (CORT) content was described previously (Dronjak *et al*, 2004). Plasma CORT was measured using the radioimmunoassay (RIA) commercial kit (ICN, Biochemical, Costa Mesa, CA, USA) and the values expressed as ng/ml. All samples were measured in duplicate in one assay. The variation between duplicate samples was less than 7%. The lowest detectable limit of hormone detection in this assay system is 25ng/ml.

Unstressed controls or stressed animals were sacrificed 2hrs following the end of the stress procedure by decapitation with a guillotine (Harvard Apparatus, South Natick, MA, USA). The livers were rapidly perfused *in situ* with ice-cold 50mM Tris-HCl buffer pH 7.55, containing 0.25M sucrose, 25mM KCl, 10mM MgCl₂ and protease-inhibitor cocktail (1.0mM DTT, 0.1mM PMSF, 5µg/ml aprotinin), excised, placed on ice, rinsed with ice-cold buffer, weighed and homogenized in 2 vol (w/v) of the same buffer by 10 strokes in the Potter-Elvehjem teflon-glass homogenizer. The homogenates were centrifuged (10 min, 10,000 rpm, 4°C) and the resulting supernatants centrifuged again (60 min, 45,000 rpm, 4°C; Beckman L8-M Ultracentrifuge) to

obtain purified cell cytosols. Protein content was determined by the method of Lowry *et al.* (1951) using bovine serum albumin (BSA, Sigma Aldrich) as a reference. The samples were mixed with denaturing buffer according to Laemmli (1970) and kept at -70 °C before the analysis.

Separation of proteins by SDS-polyacrylamide gel electrophoresis and quantification by immunoblotting

Aliquots of liver cytosols were boiled (100°C, 2 min), cooled and analyzed on 10% SDS-polyacrylamide gels using a Mini-Protean II Electrophoresis Cell (Bio-Rad Laboratories, Hercules CA, USA) according to Laemmli (1970). The gel calibration and Western transfer of proteins to nitrocellulose membranes was performed as described previously (Wrange *et al.* 1984). The membrane was incubated overnight (4°C) with either monoclonal anti-rat glucocorticoid receptor antibodies Ab-2 (GR32L, Clone BuGR2; 5µg/mL TBST; Oncogene Research Product, San Diego, CA, USA) or monoclonal anti-human Hsp70 antibodies conjugated to alkaline phosphatase (1:500 = antibody-TBST buffer; Calbiochem, La Jolla, CA, USA). The anti-human Hsp70 antibody recognized both the constitutive and inducible Hsp70 form (Čvoro *et al.* 1998). In the case of BuGR2, the secondary antibody conjugated to alkaline phosphatase (Immuno-Reporter™ Reagent DC05L, Oncogene Research Products, San Diego, CA, USA) was used for detection. The β-actin was used as an appropriate loading control. The blots were stripped with 0.1M glycine-HCl (pH 2.8) twice for 30 min at 60°C and SDS 2% for 10 min, followed by overnight saturation in 4% BSA at 4°C. They were then incubated with primary polyclonal goat anti-actin antibody (SC-1616; Santa Cruz Biotechnology) followed with alkaline phosphatase conjugated secondary anti-goat IgG antibody. The quantification of specific antigen bands was achieved by GelDoc scanning and PC processing. For figure presentation, the levels of GR and Hsp70 in stressed animals were expressed as % change in relation to those in

unstressed animals taken as 100% (controls). The data were presented as means \pm S.E.M. of 5-6 animals per group.

Statistical analysis

The obtained data were analyzed by two-way ANOVA (the factors were acute or chronic stress, and the levels for the acute stress were none, immobilization, cold, and for the chronic stress were none, isolation, crowding, swimming, isolation plus swimming). The Tukey post-hoc test was used to evaluate the differences between the groups. Statistical significance was accepted at $p < 0.05$.

Results

Levels of GR and Hsp70 in liver of animals subjected to acute, chronic and combined stress models.

In order to examine the influences of acute, chronic and combined stressors on the change in the levels of the GR and the Hsp70, we performed SDS-PAGE electrophoresis and Western blot of cytosol rat liver obtained from stressed and compared with unstressed control rats. The results of Western immunoblotting of both proteins in liver cytosol are presented in Fig. 1. The lanes of Western blot images of GR and Hsp70 are presented in Fig. 2.

The Two-way ANOVA analysis showed that animal exposure to either of the acute stressors, immobilization (Immob) or cold, resulted in a significant down-regulation of the GR levels ($F_{2,60}=44.60$, $p < 0.001$) and Hsp70 levels ($F_{2,60}=42.56$, $p < 0.001$), see Fig. 1, left panel. Both acute stressors induced a similar down-regulation of GR levels ($p < 0.001$), compared to the unstressed controls, in spite of the differences in the type and the intensity of the acute stressor. The changes in levels of Hsp70 under these models closely followed the changes in levels of cytosol GR.

Compared to acute stress effects, chronic stress models led to a weakly decrease in levels of GR ($F_{4,60}=17.60$, $p<0.001$) and Hsp70 ($F_{4,60}=8.19$, $p<0.001$). Within chronic stressors, swimming or isolation plus swimming were the stressors that led to a significant decrease in the cytosolic GR levels ($p<0.001$), (Fig. 1, middle panel) while the other two chronic stressors such as isolation or crowding did not change the cytosolic GR levels ($p>0.05$), when compared to the unstressed controls (Fig. 1, middle panel). In addition to that, the levels of GR and Hsp70 after applying all chronic stressors were significantly different from their levels after acute stress ($p<0.05$). The only exception was the chronic isolation plus 15 minutes/daily swimming in relation to the acute stress immobilization ($p>0.05$).

Rats that had been subjected to chronic stressors and then exposed to acute stressors, exhibited a significant down-regulation of hepatic GR ($F_{8,60}=19.20$, $p<0.001$) and Hsp70 levels ($F_{8,60}=3.26$, $p<0.01$), (Fig. 1, right panel) the intensity of this down-regulation was to a lower extent when compared to the acute stressors. The levels of GR and Hsp70 were between the values obtained after acute or chronic stresses alone. The only exception was the GR level after chronic crowding followed by immobilization ($p>0.05$). The levels of GR after combined stressors were significantly different from their levels after acute stressors ($p<0.01$). The only exception was in the case of chronic swimming or isolation plus swimming followed by immobilization ($p>0.05$). In the case of Hsp70 levels, the combined stress of chronic isolation, crowding or swimming followed by cold were most effective in down-regulating of this protein level ($p<0.001$) (Fig. 1, right panel). The levels of Hsp70 after all combined stressors were closer to those of acute stress and a significant difference between combined and acute stress was observed only after chronic isolation plus swimming followed by cold ($p<0.05$).

Plasma levels of corticosterone in stressed animals

The plasma levels of corticosterone (CORT) were also determined in all stressed animal groups, and the results of these measurements are presented in Fig. 3. In the acutely stressed

animals, the levels of CORT in blood plasma were significantly increased ($F_{2,75}=675.90$, $p<0.001$). Acute exposure to immobilization resulted in a significant 6-fold increase of plasma CORT levels, in relation to the unstressed controls. Also, cold as acute stress led to the 3-fold increase of CORT level, in relation to the unstressed controls.

Evaluation of chronic stress data indicated a significant increase of plasma CORT level ($F_{4,75}=94.80$, $p<0.001$). Within chronic stressors, isolation induced the most significant increase of plasma CORT levels ($p<0.001$), while other types of chronic stressors showed a relatively weak activation of the CORT secretion system which was not statistically significant ($p>0.05$), when compared to unstressed controls.

Exposure of chronically stressed rats to novel stressors (immobilization or cold) induces an exaggerated response in plasma CORT levels ($F_{8,75}=32.70$, $p<0.001$). Statistical analysis showed a 5-8 fold increase of CORT content, when compared to the unstressed controls (Fig. 3, right panel). When the results of the combined stress models were compared to those of acute stress models, it was observed that the additional acute cold stress, which resulted in a significant increase of CORT content, induced a statistically highly significant difference ($p<0.001$), while the differences between combined and acute immobilization stress were statistically slightly less significant ($p<0.01$).

Discussion

Glucocorticoids (GCs) and heat shock proteins (Hsp) are essential for normal growth, differentiation and development as well as for adaptation and survival of cells and organisms under stressors of various origins. Considering that the various stress models lead to the increase in the levels of GCs, the understanding of GCs actions *via* GR during different stress models is important to achieve an effective insight into the pathophysiology of stress-related diseases. Since the GCs are an important factor for the regulation of Hsp70 levels during the maintenance

of homeostasis, it is of special interest to know if various stress models may potentially modulate the heat shock response.

In our study, the most intensive decrease in the level of cytosolic GR and Hsp70 was observed in acutely stressed animals. As seen from the Fig. 1, the pattern of both cytosolic proteins was similar for both acute stressors, which might be explained by the existence of a functional relationship between heat shock response and GR action. At the same time, cold or immobilization stress induced marked 3- to 6-fold elevations of plasma corticosterone (CORT) levels. The results of CORT content could be connected to the data of other authors, (Alexandrova 1994, Meaney *et al.* 1991) who found that the acute immobilization stress on Sprague-Dawley rats resulted in a 3- to 4-fold elevation in plasma CORT levels. The decrease in the levels of cytosolic GR and Hsp70 in rat liver after acute elevation of GCs, is due to the translocation of cytosolic GR to the nucleus (Caamano *et al.* 2001) or its transcriptional down-regulation (Dong *et al.* 1988, Schaaf and Cidlowski 2002). The results of molecular mechanisms of GCs action showed that, at least three mechanisms could be involved in down-regulation of GR. Down-regulation may be partially controlled at the transcriptional level (Rosewicz *et al.* 1988) so that GCs administration reduced GR mRNA levels in adult rats (Kalinyak *et al.* 1987). Besides, repression of GR gene expression may be independent of the GR promoter and sequences outside the GR promoter region were involved in down-regulation of GR (Okret *et al.* 1986). Finally, a rapid decrease in receptor levels is due to a decrease in GR protein half-life (Silva *et al.* 1994) probably due to hormone-induced degradation of GR by the proteasome complex (Wallace and Cidlowski 2001, Boone and Vijayan, 2002). Concerning the cytosolic levels of Hsp70, its significant decrease in acute stress conditions may include either its decreased synthesis as GCs inhibits Hsp70 transcription via HSF1 (Wadekar *et al.* 2001) and/or its increased degradation (Boone and Vijayan, 2002). Also, it is interesting to note that plasma CORT levels in acutely immobilization stressed animals were 3-fold above the level of acutely

cold stressed animals and did not influence the intensity of GR and Hsp70 levels. This discrepancy was explained by the floor effect in which minimal levels of constitutive GR production exists (Sapolsky *et al.* 1984, Wang *et al.* 1998).

Opposite to the acute stressors, chronic stress conditions led to a weakly decrease in the levels of GR and Hsp70. At first glance, it seems that the animals were accomodating to the mild chronic stress models, especially in the group of animals exposed to isolation or crowding (psychosocial stress), judging by the GR and Hsp70 levels. The most significant decrease of GR levels was in chronic swimming or isolation plus swimming stress. These findings could be related to the data of Alexandrova and Farkas (1992) who found that the 15 days of repeated swimming stress resulted in down-regulation of GR in the rat liver. It seems that the swimming or isolation plus swimming stressor as simultaneous psychosocial and physical stress were of lower intensity than isolation or crowding as psychosocial stressors, as judged by the cytosolic levels of GR. In addition, swimming in combination with isolation may diminish detrimental effects of chronic isolation stress, judging by the nearly preserved down-regulation of cytosolic GR levels, although this helpful effect was not observed in the case of Hsp70 levels (Filipović *et al.* 2007). In the case of Hsp70, chronic stress showed a slight decrease of this protein level and had a tendency to increase under chronic isolation plus swimming stress. It may be speculated that the regulation of Hsp70 may be separate from GR regulation in simultaneous psychosocial and physical stress. At the same time, chronically stressed rats subjected to crowding, did not show changes in plasma CORT levels, swimming or isolation plus swimming showed a relatively weak activation of the CORT secretion system, while chronically isolated animals showed 5-fold increased CORT levels, when compared to the unstressed controls. At the intracellular level, CORT induced down-regulation of GR and Hsp 70. Since the intensity of down-regulation of GR in chronic isolation was less pronounced than its down-regulation under acute stress, it may be suggested that GR may be structurally affected yielding the non-functional

receptor unable to interrupt chronic stress CORT signal by its own down-regulation. Also, the existence of oxidative stress in chronic isolation (Filipović and Radojčić 2005) may lead to oxidation of GR sulfhydryl groups and its functional alteration (Elez *et al.*, 2000).

Chronically stressed rats, which were subsequently exposed to novel acute stressors (immobilization or cold), showed significantly elevated plasma CORT levels, when compared to chronic stressors. This is in agreement with the findings of Hashiguchi *et al.* (1997) who suggested that CORT is especially sensitive to novel treatment. The plasma CORT levels between combined and acute cold stress showed a high statistical significance, while the differences between combined and acute stress immobilization were less significant. At the same time, combined stress models showed a lower extent of GR and Hsp70 down-regulation as compared to acute stress models. The intensity of these changes for GR protein was between values obtained for acute or chronic stress alone. There were highly significant statistical differences of GR protein levels between combined and acute stress, while the differences between combined and chronic stress were less significant. These still high levels of cytosolic GR and Hsp70 after subsequent both acute stressors could be the result of impaired nucleocytoplasmic shuttling of GR under chronic stress leading to the incomplete down-regulation of GR and Hsp70. Our results suggest that impaired translocation of cytosolic GR to the nucleus after all chronic stressors, compromise signaling transduction of consecutive acute stressors. It is interesting to speculate that chronic stress-induced effects might reflect the alternations in the GR and Hsp70 phosphorylation status which is casually linked to its nucleocytoplasmic shuttling (DeFranco *et al.* 1991, Rouse *et al.* 1994). In addition, the phospho-deficient mutant of mouse GR does not undergo ligand-dependent down-regulation (Webster *et al.* 1997). Although the precise mechanism of regulation of Hsp70 expression is not well understood, it seems, at least taking into consideration the stress models in this investigation, that both GR and Hsp70 were mutually inter-regulated (Čvoro *et al.* 1998) or co-regulated by co-

activators (Jonat *et al.* 1990, McKay and Cidlowski 1998, Doucas *et al.* 2000) concerning both their pattern levels, as well as their stress-dependent pattern.

Finally, changes in the levels of GR in chronic or combined stress, lead to the conclusion that, in addition to the hypothalamo-pituitary adrenal (HPA) axis (Filipović *et al.* 2005), chronic stressors also compromise intracellular GR down-regulation in the liver. In addition, GR and Hsp70 levels, the major stress related cellular proteins in liver, could serve as cytosolic parameters for the distinction between acute and chronic stress, rather than stress intensity or type. Since the GCs stimulate the process of hepatic gluconeogenesis, resulting in elevated plasma glucose (Rizza *et al.* 1982) and promote the deposition of liver glycogen by the expression of genes of the phosphoenolpyruvate carboxykinase (Imai *et al.* 1993), incomplete down-regulation of GR after chronic stress as well as high levels of GCs may result in significant metabolic and physiological perturbation in this organ as well as in the stressed organism as a whole. In view that CuZnSOD expression is regulated by GR (Kim *et al.* 1994) as well as the level of antioxidative enzyme CuZnSOD was moderately increased under these conditions in the brain cortex and hippocampus of Wistar rats (Filipović and Radojčić 2005), it may be speculated that a similar antioxidative status could also be expected in the liver cells. Further studies are needed to clarify this notation.

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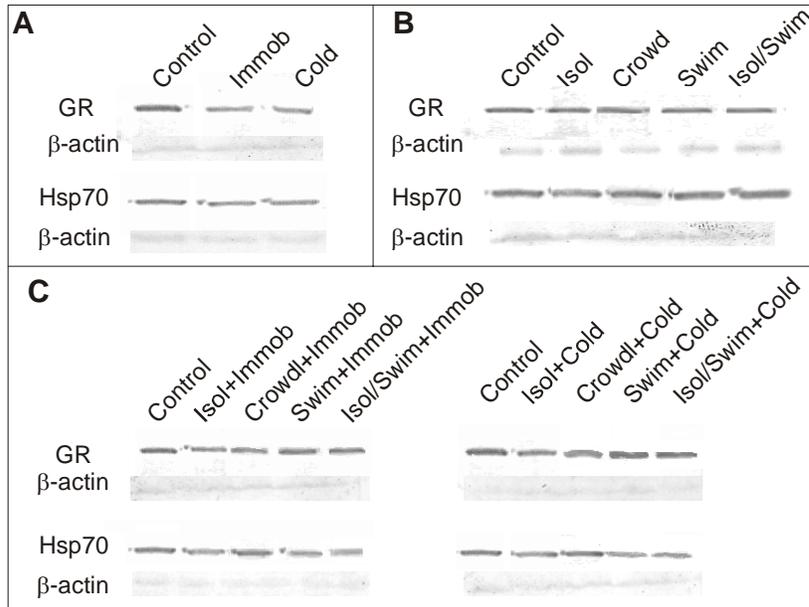
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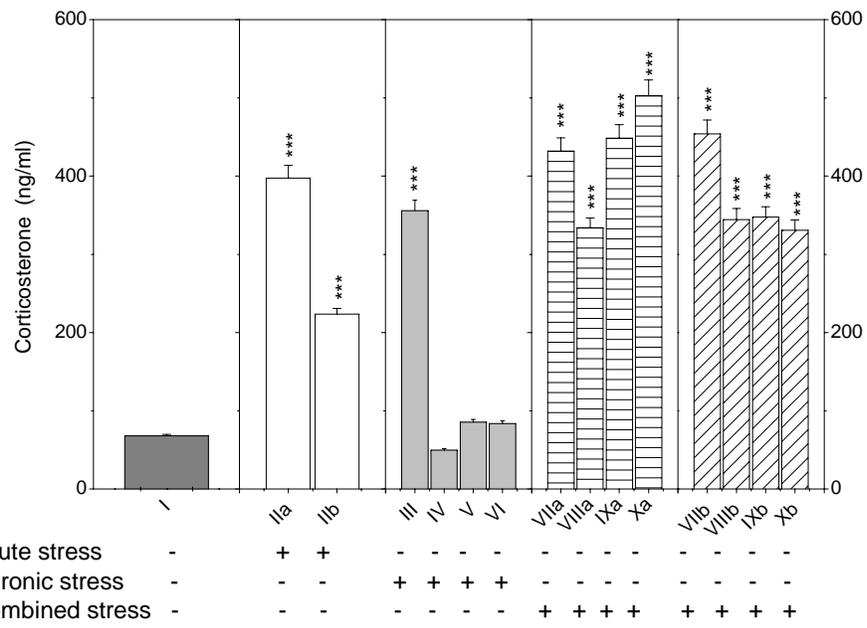
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Legends

Fig. 1

Relative changes, expressed as % of unstressed controls, in the levels of glucocorticoid receptor (GR) and heat shock protein 70 (Hsp70) in the liver of animals exposed to acute stressors: immobilization (IIa) or cold (IIb); chronic stressors: isolation (III), crowding (IV), swimming (V), isolation plus swimming (VI) or combined stressors such as: isolation plus immobilization or cold VIIa/VIIb, crowding plus immobilization or cold VIIIa/VIIIb, swimming plus immobilization or cold IXa/IXb, isolation/swimming plus immobilization or cold Xa/Xb, as indicated. The results are shown as the mean \pm S.E.M. Asterisks indicate a significant difference between the respective stress groups and unstressed controls; * $p < 0.05$ and *** $p < 0.001$ by Tukey post-hoc test.

Fig. 2 Western blot images of specific immunoreactivity of antibodies to glucocorticoid receptor (GR) and heat shock protein 70 (Hsp70) in the liver cytosol of control animals and animals exposed to: A-acute stressors immobilization (Immob) or cold; B-chronic stressors isolation (Isol), crowding (Crowd), swimming (Swim), isolation plus swimming (Isol/Swim) and C-combined stressors (data are presented as described in the legend of Fig.1), as indicated.

Fig. 3 Plasma levels of corticosterone (CORT) (ng/ml) in the control animals and animals exposed to acute stressors: immobilization (IIa) or cold (IIb); chronic stressors: isolation (III), crowding (IV), swimming (V), isolation plus swimming (VI) or combined stressors such as: isolation plus immobilization or cold VIIa/VIIb, crowding plus immobilization or cold VIIIa/VIIIb, swimming plus immobilization or cold IXa/IXb, isolation/swimming plus immobilization or cold Xa/Xb, as indicated. The results are shown as the mean \pm S.E.M.

Asterisks indicate a significant difference between the respective stress groups and unstressed controls; *** $p < 0.001$ by Tukey post-hoc test.