

**SKIN BLOOD FLOWMOTION AND MICROVASCULAR REACTIVITY
INVESTIGATION IN HYPERCHOLESTEROLEMIC PATIENTS WITHOUT
CLINICALLY MANIFEST ARTERIAL DISEASES**

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Short title: Skin blood flowmotion in hypercholesterolemic patients

Summary

Fourier spectral analysis of forearm skin laser Doppler flowmetry (LDF) signal was performed in fifteen hypercholesterolemic patients (HP), without clinically manifest arterial diseases, and in fifteen age matched healthy control subjects (CS), in order to investigate skin blood flowmotion (SBF). The LDF frequency intervals studied were: 0.01-1.6 Hz total spectrum, as well as 0.01-0.02 Hz (endothelial), 0.02-0.06 Hz (sympathetic), 0.06-0.2 Hz (myogenic), 0.2-0.6 Hz (respiratory) and 0.6-1.6 Hz (cardiac) sub-intervals. Skin microvascular reactivity (MVR) to acetylcholine (ACh) and to sodium nitroprusside (SNP) iontophoresis was also investigated. HP showed a lower post-ACh increase in power spectral density (PSD) of the 0.01-0.02 Hz SBF sub-interval compared to CS ($1.80 \pm 1.73 \text{ PU}^2/\text{Hz}$ vs $3.59 \pm 1.78 \text{ PU}^2/\text{Hz}$, respectively; $p < 0.005$), while they did not differ in MVR from CS. In eleven HP the 0.01-0.02 Hz SBF sub-interval showed a higher post-ACh PSD increase near to the statistical significance after ten weeks of rosuvastatin therapy (10 mg/day) compared to pre-treatment test ($3.04 \pm 2.95 \text{ PU}^2/\text{Hz}$ vs $1.91 \pm 1.94 \text{ PU}^2/\text{Hz}$, respectively; $p = 0.07$). The blunted post-ACh increase in PSD of the 0.01-0.02 Hz SBF sub-interval in HP suggests a skin endothelial dysfunction in these patients. This SBF abnormality showed a tendency to improve after rosuvastatin therapy in eleven treated patients.

Keys words: hypercholesterolemia, endothelial function, laser Doppler flowmetry, flowmotion, spectral Fourier analysis, acetylcholine, sodium nitroprusside.

Introduction

A large number of studies demonstrated endothelial dysfunction of large arteries in hypercholesterolemic patients and suggested that this abnormality is the principal mechanism responsible for the atherosclerotic damage and cardiovascular risk in these patients (Manninen *et al.* 1992, Vogel *et al.* 1999, Stein *et al.* 2001, Davignon *et al.* 2004). More recently, studies based on laser Doppler flowmetry (LDF) investigated skin microvascular reactivity to the endothelial-dependent vasodilator acetylcholine (ACh) (Khan *et al.* 1999, Binggeli *et al.* 2003), as well as to ischemic or thermal stimuli (Štulc *et al.* 2003), in hypercholesterolemic patients. The interest for the study of microvascular reactivity in hypercholesterolemic patients arises from the hypothesis, recently suggested, that skin microvascular function can mirror the state of microcirculation in other microvascular beds, including cardiac muscle (Jung *et al.* 2001, Shamin-Uizzaman *et al.* 2002). Using the above mentioned method, a blunted skin vasodilator response to ACh, consistently with endothelial dysfunction, has been found has been demonstrated in hypercholesterolemic patients with manifest coronary artery disease (Khan *et al.* 1999), while a preserved skin vasoreactivity to ACh has been observed in hypercholesterolemic patients without manifest vasculopathic state (Khan *et al.* 1999), consistently with a preserved endothelial function in these patients. Moreover, a preserved skin microvascular reactivity to ischemic and thermal stimuli was found in hypercholesterolemic patients without coronary artery disease (Štulc *et al.* 2003), However, no studies in hypercholesterolemic patients explored the skin blood flow oscillation (the so called flowmotion), which has been demonstrated to be in part related to the rhythmic contraction and dilation of skin arterioles, the so called vasomotion (Colantuoni *et al.* 1984, Colantuoni *et al.* 1994).

Vasomotion, and the consequent skin blood flowmotion (SBF), has been demonstrated to be due to different mechanisms such as the endothelial activity and the spontaneous myogenic activity of the microvascular wall (Kvernmo *et al.* 1999, Kvandal *et al.* 2003). The efficiency of these two specific vasomotion mechanisms can be indirectly evaluated by investigating SBF by the spectral analysis of the skin LDF signal (Stefanovska *et al.* 1997, Stefanovska *et al.* 1999, Kvernmo *et al.* 1999, Rossi *et al.* 2004, Rossi *et al.* 2006). Five different frequency sub-intervals have been described within the 0.01-1.6 Hz SBF total spectrum in these studies (Stefanovska *et al.* 1997, Stefanovska *et al.* 1999, Kvernmo *et al.* 1999, Rossi *et al.* 2004, Rossi *et al.* 2006). Two of them, the SBF frequency sub-interval of 0.01-0.02 Hz and the SBF frequency sub-interval of 0.06-0.2 Hz, have been showed to be related to the endothelial-dependent or to the myogenic-dependent vasomotion, respectively (Stefanovska *et al.* 1999, Kvernmo *et al.* 1999). The SBF frequency sub-interval of 0.02-0.06 Hz has been attributed to the local sympathetic activity (Staass *et al.* 1998), and finally the SBF frequency sub-intervals of 0.2-0.6 Hz and of 0.6-1.6 Hz have been considered due to the transmission to the skin microcirculation of the central haemodynamic modifications synchronous with respiratory (Bollinger *et al.* 1993) and heart activity (Stefanovska *et al.* 1997), respectively.

The aims of our study were: 1) to study the SBF in hypercholesterolemic patients without clinically manifest arterial diseases in order to investigate skin microvascular endothelial function in these patients; 2) to evaluate possible changes of SBF in the same patients following rosuvastatin therapy.

Subjects and Methods

Subjects

Fifteen patients affected by hypercholesterolemia were selected from patients who underwent cardiovascular check during the last year in our outpatients clinic. Eligible were patients over 18

years old, with LDL-cholesterol serum levels > 140 mg/dl, never treated with lipid-lowering medication. Patients suffering from diabetes mellitus, arterial hypertension, chronic renal or heart failure, and with clinical evidence of lower limb vascular disease, coronary or cerebral artery disease were excluded, as well as smokers. Fifteen normocholesterolemic (with LDL-cholesterol serum levels <100 mg/dl) normotensive, non-smoker healthy subjects, age matched with the hypercholesterolemic patients, were recruited among companions of outpatients referring to our clinic, to form control group. Table 1 shows the clinical and laboratory characteristics of the fifteen hypercholesterolemic patients and of the fifteen control subjects recruited for the study. Hypercholesterolemic patients showed a statistically significant higher mean value of total cholesterol ($p<0.005$), of LDL-cholesterol serum level ($p<0.005$), as well as of diastolic blood pressure value ($p<0.05$), than control subjects. No significant difference was observed in systolic blood pressure mean value as well as in respiratory and heart rate mean value between the studied groups. All recruited subjects gave their consent to take part on the study and the local Department Committee approved the study.

Skin blood flux measurement

Skin blood flux was measured in conventional perfusion units (PU: 1 PU= 10 mV) by means of a LDF apparatus (Periflux PF4001, Perimed, Järfälla, Sweden), equipped with a not heated probe (PF408). The laser characteristics were: 780 nm wavelength, 10 Hz-19 KHz bandwidth, 0.1 s time constant, 32 Hz sampling frequency. Calibration was performed before each test session, by a specific device (Perimed, Järfälla, Sweden) containing colloidal latex particles whose Brownian motion provides the standard values. The LDF signal has been recorded continuously by an interfaced computer (Acer, Travelmate 202 T) equipped with Perisoft dedicated software.

Acetylcholine and sodium nitroprusside iontophoresis

ACh and sodium nitroprusside (SNP) iontophoresis was performed as previously described (Morris *et al.* 1995). A battery-powered iontophoresis controller (PeriIont 328, Perimed, Järfälla, Sweden), equipped with a drug delivery electrode (PF 383, Perimed, Järfälla, Sweden) was used for delivering ACh or SNP to the skin. The drug delivery electrode was connected to the LDF apparatus in order to register the blood flux before and during iontophoresis. An indifferent electrode (PF 384, Perimed, Järfälla, Sweden), was used to provide the current needed for ACh and SNP delivery.

Each subject avoided food, drugs, tobacco, alcohol, coffee or tea 10 h prior to the test and had 20 min of acclimatisation in supine position before the test. Room temperature was between 22 - 24°C. Before the examination, a drug delivery electrode was filled with 0.05 ml of 1 % ACh solution and then attached to the dorsal aspect of the third right finger by double-sided adhesive disc. The indifferent electrode was attached on the dorsal aspect of the right hand. ACh was then delivered to the skin by means of an anodal current: nine pulses (0.1 mA for 20 seconds each) with a 60 second interval between one to the other. After ACh iontophoresis, a drug delivery electrode, previously filled with 0.05 ml of 1 % SNP solution, was attached to the dorsal aspect of the third left finger by double-sided adhesive disc. The indifferent electrode was attached on the dorsal aspect of the left hand. SNP was then delivered to the skin by means of a cathodal current: seven pulses (0.2 mA for 20 seconds each) with a 180 seconds interval between one to the other. Maximal skin blood flux reached during ACh or SNP iontophoresis was measured in PU. The maximal hyperaemic response during ACh or to SNP iontophoresis was also expressed as maximal per cent change from baseline.

Skin blood flowmotion investigation

Skin blood flowmotion was investigated by means of spectral analysis of the skin LDF signal.

For the spectral analysis, as in previous studies (Serné *et al.* 2002, Rossi *et al.* 2004, Rossi *et al.* 2006) we used a Perisoft dedicated software (Perimed, Järfälla, Sweden). This software measures in PU^2/Hz the power spectral density (PSD) of LDF signal, using the basic Fast Fourier Transform algorithm. In this algorithm the beginning and the end of the signal are attenuated by means of a windowing “Parzen” function to avoid the well known “leakage phenomenon” (frequency components in the spectra “leaking” into other frequencies). In the windowing “Parzen” function, a short-time Fourier transform, with a different window length for each frequency interval, was used. The frequency interval from 0.01 to 1.6 Hz was divided into five sub-intervals: 0.01–0.02 Hz (endothelial activity), 0.02–0.06 Hz (sympathetic activity), 0.06–0.2 Hz (vascular myogenic activity), 0.2–0.6 Hz (respiratory activity) and 0.6–1.6 Hz (heart activity). The PSD in a definite frequency sub-interval has been calculated as the highest PSD in that sub-interval. PSD of the 0.01–1.6 Hz SBF total spectrum was obtained by the sum of the PSD of the five SBF sub-interval considered.

Using the above described method, the spectral analysis was performed on the skin LDF tracing registered during 10 minutes before ACh iontophoresis and on the skin LDF tracing registered during the whole duration of ACh iontophoresis procedure. The post-ACh iontophoresis change in PSD for each SBF sub-interval considered was expressed as difference between the PSD value reached during ACh iontophoresis and the baseline PSD value of that sub-interval. Similarly the post-ACh iontophoresis change in PSD of SBF total spectrum was expressed.

Lipid-lowering intervention

Hypercholesterolemic patients were treated with rosuvastatin, 10 mg daily for 10-weeks, starting the treatment in the day of LDF-iontophoresis test. The day after the last rosuvastatin assumption hypercholesterolemic patients underwent a second LDF-iontophoresis test.

Laboratory measurements

The standard biochemical tests were performed in the day before the beginning of rosuvastatin therapy and in the last day of rosuvastatin therapy by automated analyzer methods.

Statistical analysis

Results were expressed as mean \pm SD or as mean \pm SEM when specified.

Student's "t" test for paired data was used to compare basal LDF blood flux values and maximal LDF blood flux values following Ach or SNP iontophoresis obtained in each group, as well as LDF blood flux values and laboratory parameter values obtained before and after rosuvastatin therapy in hypercholesterolemic patients. Student's "t" test for non paired data was used to compare LDF blood flux values obtained in hypercholesterolemic patients and in control subjects.

Wilcoxon test for non paired data was used to compare PSD values obtained in the two studied groups. Wilcoxon test for paired data was used to compare PSD values obtained in baseline and during ACh iontophoresis in each group of subjects, as well as PSD values obtained before and after rosuvastatin therapy in hypercholesterolemic patients. When Wilcoxon test was used, the p value was corrected by means of Bonferroni method.

A p value <0.05 was considered as statistically significant.

Results

Hypercholesterolemic patients versus control subjects

The skin blood flux values obtained in control subjects and in hypercholesterolemic patients are reported in the upper part of table 2. There was no significant difference in skin blood flux under basal conditions between hypercholesterolemic patients and control subjects (25.6 ± 11.6 PU and 19.9 ± 12.3 PU, respectively). Following ACh or SNP iontophoresis there was a significant increase

in skin blood flux both in control subjects and in hypercholesterolemic patients. Hypercholesterolemic patients and control subjects did not differ in per cent maximal skin hyperemic response to Ach iontophoresis (803 ± 496 % and 791 ± 456 %, respectively) or to SNP iontophoresis (549 ± 222 % and 671 ± 367 %, respectively) .

As concerned the results of SBF investigation, hypercholesterolemic patients and control subjects did not statistically differ in baseline PSD of SBF total spectrum (3.44 ± 2.59 PU²/Hz vs 4.54 ± 2.45 PU²/Hz, respectively) as well as in baseline PSD of endothelial-dependent 0.01-0.02 Hz sub-interval (0.83 ± 0.59 PU²/Hz vs 1.12 ± 0.55 PU²/Hz, respectively), sympathetic-related 0.02-0.06 Hz sub-interval (0.91 ± 0.69 PU²/Hz vs 1.02 ± 0.47 PU²/Hz, respectively), myogenic-related 0.06-0.2 Hz sub-interval (0.60 ± 0.45 PU²/Hz vs 1.11 ± 1.06 PU²/Hz, respectively), respiration-related 0.2-0.6 Hz sub-interval (0.34 ± 0.30 PU²/Hz vs 0.26 ± 0.10 PU²/Hz, respectively) and heart activity-related 0.6-1.6 Hz sub-interval (0.78 ± 0.82 PU²/Hz vs 1.03 ± 0.89 PU²/Hz, respectively).

Following ACh iontophoresis, a significant increase in PSD of 0.01-1.6 Hz total spectrum, as well as in PSD of each of sub-interval considered, was observed both in control subjects (from $p < 0.01$ to $p < 0.0001$) and in hypercholesterolemic patients (from $p < 0.05$ to $p < 0.005$). However, compared to control subjects, hypercholesterolemic patients showed a statistically significant lower post-ACh iontophoresis increase in PSD of 0.01-1.6 total spectrum (7.90 ± 7.70 PU²/Hz vs 11.44 ± 4.79 PU²/Hz, $p < 0.05$), as well as of the endothelial-dependent 0.01-0.02 Hz sub-interval (1.80 ± 1.73 PU²/Hz vs 3.59 ± 1.78 PU²/Hz, $p < 0.005$) and of the respiration related 0.2-0.6 Hz sub-interval (0.30 ± 0.42 PU²/Hz vs 0.53 ± 0.30 PU²/Hz, $p < 0.005$) (figure 1). No significant difference in post-ACh iontophoresis PSD increase of the other SBF sub-intervals considered was observed between hypercholesterolemic patients and control subjects (figure 1).

Changes associated to rosuvastatin therapy in hypercholesterolemic patients

Eleven hypercholesterolemic patients completed the 10 week period of rosuvastatin therapy and underwent post-therapy laboratory and LDF tests.

Basal skin blood flux mean values and the maximal absolute or per cent hyperaemic response to ACh or to SNP iontophoresis did not significantly change at the end of rosuvastatin therapy compared to before therapy in the eleven hypercholesterolemic patients (lower part of table 2).

Table 3 shows the laboratory data observed before and at the end of rosuvastatin therapy in the same patients. A statistically lower total cholesterol ($p < 0.0001$) and LDL-cholesterol ($p < 0.01$) serum levels mean value was observed at the end of rosuvastatin therapy compared to before therapy. No significant change in HDL-cholesterol, triglycerides or glucose serum levels, as well as in diastolic or systolic blood pressure values was observed in the same patients at the end of rosuvastatin therapy compared to before therapy.

As concerned the results of SBF investigation, no statistically significant difference in baseline PSD of 0.01-1.6 Hz total spectrum values was observed between before and after rosuvastatin therapy in the eleven hypercholesterolemic patients (3.08 ± 2.17 vs 4.86 ± 3.59), as well as in baseline PSD of endothelial-dependent 0.01-0.02 Hz sub-interval ($0.71 \pm 0.53 \text{ PU}^2/\text{Hz}$ vs $1.19 \pm 1.02 \text{ PU}^2/\text{Hz}$, respectively), sympathetic-related 0.02-0.06 Hz sub-interval ($0.84 \pm 0.67 \text{ PU}^2/\text{Hz}$ vs $1.16 \pm 0.98 \text{ PU}^2/\text{Hz}$, respectively), myogenic-related 0.06-0.2 Hz sub-interval ($0.53 \pm 0.37 \text{ PU}^2/\text{Hz}$ vs $0.93 \pm 0.74 \text{ PU}^2/\text{Hz}$, respectively), respiration-related 0.2-0.6 Hz sub-interval ($0.26 \pm 0.14 \text{ PU}^2/\text{Hz}$ vs $0.38 \pm 0.29 \text{ PU}^2/\text{Hz}$, respectively) or heart activity-related 0.6-1.6 Hz sub-interval ($0.74 \pm 0.76 \text{ PU}^2/\text{Hz}$ vs $1.20 \pm 0.74 \text{ PU}^2/\text{Hz}$, respectively). No statistically significant change in post-ACh PSD increase from baseline (mean \pm SEM difference from baseline) was observed from before to after rosuvastatin therapy for each of the SBF sub-intervals investigated, as well as for SBF total

spectrum (figure 2). However, the change from before to after rosuvastatin therapy in post-ACh PSD increase from baseline of the endothelial-dependent 0.01-0.02 Hz SBF sub-interval was very near to the statistical significance (from $1.91 \pm 1.94 \text{ PU}^2/\text{Hz}$ to $3.04 \pm 2.85 \text{ PU}^2/\text{Hz}$; $p=0.07$) (figure 2).

Discussion

To our knowledge, this is the first study which explored in hypercholesterolemic patients the LDF skin blood flowmotion, whose investigation has been proposed for evaluating the skin microvascular function in clinical setting (Kvernmo *et al.* 1999, Kvandal *et al.* 2003). We observed a blunted ACh-induced increase in the spectral amplitude of 0.01-0.02 Hz SBF component, known to be related to endothelial activity (Stefanovska *et al.* 1999, Kvernmo *et al.* 1999), in our hypercholesterolemic patients without clinically manifest arterial disease. The same patients also showed a blunted ACh-induced increase in spectral amplitude of the 0.01-1.6 Hz SBF total spectrum, as well as of the 0.2-0.6 Hz SBF component (known to be related to respiratory activity), while they did not differ from control subjects in the response to ACh of the other SBF components. Moreover, similarly to a previous study (Khan *et al.* 1999), our hypercholesterolemic patients without clinically manifest arterial disease exhibited a preserved skin vasodilator response to ACh and SNP iontophoresis.

The spectral amplitude increase of the 0.01-0.02 SBF Hz component in response to the endothelium-dependent vasodilator ACh has been shown to reflect the efficiency of skin microvascular endothelial function (Kvernmo *et al.* 1999, Kvandal *et al.* 2003). Therefore, the finding of a blunted ACh-induced spectral amplitude increase of the 0.01-0.02 Hz SBF component, we observed in our hypercholesterolemic patients, could be an early sign of skin microvascular endothelial dysfunction in these patients. Considering the exclusion criteria, we used for selecting hypercholesterolemic patients, such as arterial hypertension, smoke or diabetes, potentially able to induce skin microvascular endothelial dysfunction, this finding may be ascribed to the hypercholesterolemic condition.

Previous studies, based on LDF measurement of the skin vasodilator response to ACh and SNP iontophoresis, failed to clearly demonstrate endothelial dysfunction in the skin microvascular bed of hypercholesterolemic patients without manifest vascular disease (Khan *et al.* 1999). However, our finding of blunted ACh-induced increase of the endothelial-dependent SBF component in hypercholesterolemic patients, in spite of a preserved skin vasodilator response to ACh in the same patients, suggests that SBF investigation is more sensitive in the study of skin endothelial function than the LDF measurement of skin blood flux response to ACh. On the other hand, the study of the endothelial function in the skin microvascular bed is clinically important in hypercholesterolemic patients, taken into account that endothelial cell dysfunction is foremost a microvascular disease (Stewart *et al.* 2004) and that the investigation of the cutaneous microcirculation using LDF may adequately reflect the vascular involvement of other vascular beds, including cardiac muscle (Jung *et al.* 2001, Shamin-Uizzaman *et al.* 2002).

The blunted ACh-induced spectral amplitude increase of the 0.2-0.6 Hz SBF component, related to respiratory activity, occurred in hypercholesterolemic patients without change in respiratory rate, suggesting that this finding was due to a reduced transmission to the skin microcirculation of the central haemodynamic modifications induced by respiration. Moreover, the blunted ACh-induced spectral increase of the SBF 0.01-1.6 Hz total spectrum could be only a secondary finding due to the blunted ACh-induced increase of the endothelial and respiration-related SBF components.

A second aim of our study was to evaluate possible changes in microvascular endothelial function associated to rosuvastatin therapy in our hypercholesterolemic patients.

Previous studies (Khan *et al.* 1999, Haak *et al.* 2001, Štulc *et al.* 2003), based on LDF measurement of skin vasodilator response to different stimuli, failed to clearly demonstrate a positive effect of statin therapy on skin microvascular endothelial function in hypercholesterolemic patients.

Fluvastatin therapy induced a significant increase in skin vasodilator response to SNP, but no to ACh, in hypercholesterolemic patients with peripheral arterial obstructive disease (Khan *et al.* 1999), consistently only with an improvement of the skin endothelium-independent microvascular reactivity. A marginal improvement in the skin vasodilator response to thermal stimulus was observed in hypercholesterolemic patients after fluvastatin therapy (Haak *et al.* 2001). Moreover, no change in skin microvascular reactivity to ischemic or thermal stimuli was observed in hypercholesterolemic patients after atorvastatin therapy (Štulc *et al.* 2003) .

Similarly to the study of Khan *et al.*, we did not observe any significant increase in skin vasodilator response to ACh iontophoresis in our hypercholesterolemic patients after statin therapy. However, at the end of rosuvastatin therapy there was an ACh-induced spectral amplitude increase of the endothelial-related SBF component very near to the statistical significance compared to the pre-treatment investigation, which could reflect the tendency to an improvement in skin endothelial function associated to rosuvastatin therapy. Further studies based on a controlled design and with a bigger number of patients are needed to clearly demonstrate a positive effect of rosuvastatin therapy on endothelial function in hypercholesterolemic patients.

A possible limitation of our study was the relatively small number of the studied subjects which could make low the power of the statistical analysis of our findings on SBF investigation. We did not performed any study on the SBF investigation reproducibility. However, in a previous study (Bernardi *et al.* 1997) the inter- and intra-day variation of the low-frequency or high-frequency SBF components was not higher than 15% or 13% respectively. These findings may give sufficient reliability to our statistical analysis. A possible source of misinterpretation of our findings on the study of SBF during ACh iontophoresis could arise from the instability of LDF signal during this procedure. However, the ACh iontophoresis protocol, we used in the present study, elicited a rapid and progressive increase of the laser Doppler signal amplitude beginning from the first iontophoretic pulses, followed by a stable laser Doppler signal. The absence of wide and rapid LDF

tracing fluctuations following each ACh iontophoresis stimulation makes sufficiently reliable the results of the SBF spectral analysis investigation.

A possible caveat of this study was the prevalence of women among control subjects, while men were prevalent among hypercholesterolemic patients. However the post-menopausal age of the recruited women in the control group may minimize this possible caveat. Another possible limitation of this study is the statistically higher diastolic blood pressure in hypercholesterolemic patients compared to control subjects. However, no hypercholesterolemic patient exceeded the normal values of arterial blood pressure.

In conclusion, our study in hypercholesterolemic patients without clinically manifest arterial diseases showed a blunted ACh-induced increase in spectral amplitude of 0.01-0.02 Hz endothelial related SBF component, which could be an early sign of skin endothelial dysfunction in these patients. A preserved skin vasodilator response to ACh or to SNP was also observed in the same patients. After rosuvastatin therapy there was an ACh-induced spectral amplitude increase of the endothelial-related SBF component very near to the statistical significance, which could reflect the tendency to an improvement in skin endothelial function associated to rosuvastatin therapy. The assessment of SBF by spectral analysis of skin LDF signal seems to be a promising method for implementing the efficacy of conventional LDF tests in investigating skin microvascular endothelial function in clinical setting.

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Abbreviations

LDF = laser Doppler flowmetry
ACh = acetylcholine
SBF = skin blood flowmotion
SNP = sodium nitroprusside
PSD = power spectral density
LDL = low density lipoprotein
SD = standard deviation
SEM = standard error medium

Legendes

Table 1

Clinical and laboratory features in the fifteen hypercholesterolemic patients and in fifteen control subjects are reported (values are expressed as mean \pm SD).

* $p < 0.05$

** $p < 0.005$

Table 2

In the upper part of the table, basal skin blood flux values (mean \pm SD) and maximal skin blood flux values (mean \pm SD) following acetylcholine or sodium nitroprusside iontophoresis, observed in fifteen hypercholesterolemic patients and in fifteen control subjects, are reported. Maximal skin blood flux per cent changes from baseline observed in the same two groups are also reported.

In the lower part of the table, basal skin blood flux values and maximal skin blood flux values following acetylcholine or sodium nitroprusside iontophoresis, observed in eleven hypercholesterolemic patients before and after rosuvastatin therapy, are reported. Maximal per cent changes from baseline, observed before and after rosuvastatin therapy in the same patients, are also reported.

The “p” values in the table are referred to the statistic difference between baseline and following acetylcholine or sodium nitroprusside iontophoresis.

PU = perfusion unit

ACh = acetylcholine

SNP = sodium nitroprusside

Table 3

Laboratory parameters values (mean \pm SD) observed before and after rosuvastatin therapy in eleven hypercholesterolemic patients.

* $p < 0.0001$

** $p < 0.01$

Figure 1

Differences in power spectral density (PU^2/Hz) between acetylcholine iontophoresis and baseline (mean \pm SEM) for each skin blood flowmotion sub-interval considered, as well as for 0.01-1.6 Hz skin blood flowmotion total spectrum, observed in fifteen hypercholesterolemic patients (black histograms) and in fifteen control subjects (grey histograms).

PSD = power spectral density; ACh = acetylcholine

* = $p < 0.005$ between hypercholesterolemic patients and control subjects.

* * = $p < 0.05$ between hypercholesterolemic patients and control subjects.

Figure 2

Differences in power spectral density (PU^2/Hz) between acetylcholine iontophoresis and baseline (mean \pm SEM) for each skin blood flowmotion sub-interval considered, as well as for 0.01-1.6 Hz skin blood flowmotion total spectrum, observed before (grey histograms) and after rosuvastatin (black histograms) therapy in eleven hypercholesterolemic patients.

PSD = power spectral density; ACh = acetylcholine

Table 1

Clinical or laboratory feature	Control subjects (n° 15)	Hypercholesterolemic patients (n° 15)
Age (years)	57 ± 11	59 ± 10
Sex (M/F)	5 / 10	12 / 3
Body-mass index	24.8 ± 3.4	26.2 ± 3.3
Systolic blood pressure(mmHg)	126 ± 13	133 ± 15
Diastolic blood pressure (mmHg)	77 ± 6	82 ± 7 *
Total cholesterol (mmol/l)	4.99 ± 0.55	6.90 ± 1.24 * *
LDL cholesterol (mmol/l)	2.72 ± 0.65	4.51 ± 0.77 * *
LDH cholesterol (mmol/l)	1.65 ± 0.72	1.44 ± 0.24
Triglycerides (mmol/l)	0.84 ± 0.28	2.43 ± 2.51
Glucose (mmol/l)	4.87 ± 0.41	5.19 ± 0.85

Table 2

	Basal blood flux (PU)	Post-ACh maximal blood flux (PU)	Post-ACh change from baseline (%)	Basal blood flux (PU)	Post-SNP maximal blood flux (PU)	Post-SNP change from baseline (%)
Control subjects (n= 15)	25.6± 11.6	173.7±68.7 p<0.000001	803±496	27.4±12.9	139.3±67.0 p<0.000005	549±222
Hyperchol. pat. (n= 15)	19.9±12.3	131.2±86.7 p< 0.0005	791±456	22.0±12.1	129.7±84.1 p< 0.0005	671±367

	Basal blood flux (PU)	Post-ACh maximal blood flux (PU)	Post-ACh change from baseline (%)	Basal blood flux (PU)	Post-SNP maximal blood flux (PU)	Post-SNP change from baseline (%)
Before rosuvastatin (n= 11)	20.4±12.7	116.1±81.1 p< 0.005	710±435	23.5±12.6	150.3±88.9 p< 0.001	732±383
After rosuvastatin (n= 11)	36.0±26.2	187.6±104.5 p<0.0005	602±193	27.5±19.6	173.9±95.4 p< 0.0005	856±475

Table 3

Laboratory parameter	Before rosuvastatin	After rosuvastatin
Total cholesterol (mmol/l)	7.07 ± 1.34	4.96 ± 1.08 *
LDL cholesterol (mmol/l)	4.53 ± 0.82	2.94 ± 0.89 **
HDL cholesterol (mmol/l)	1.47 ± 0.24	1.35 ± 0.17
Triglycerides (mmol/l)	2.75 ± 2.86	1.64 ± 0.93
Glucose (mmol/l)	5.31 ± 0.87	5.46 ± 0.77

Figure 1

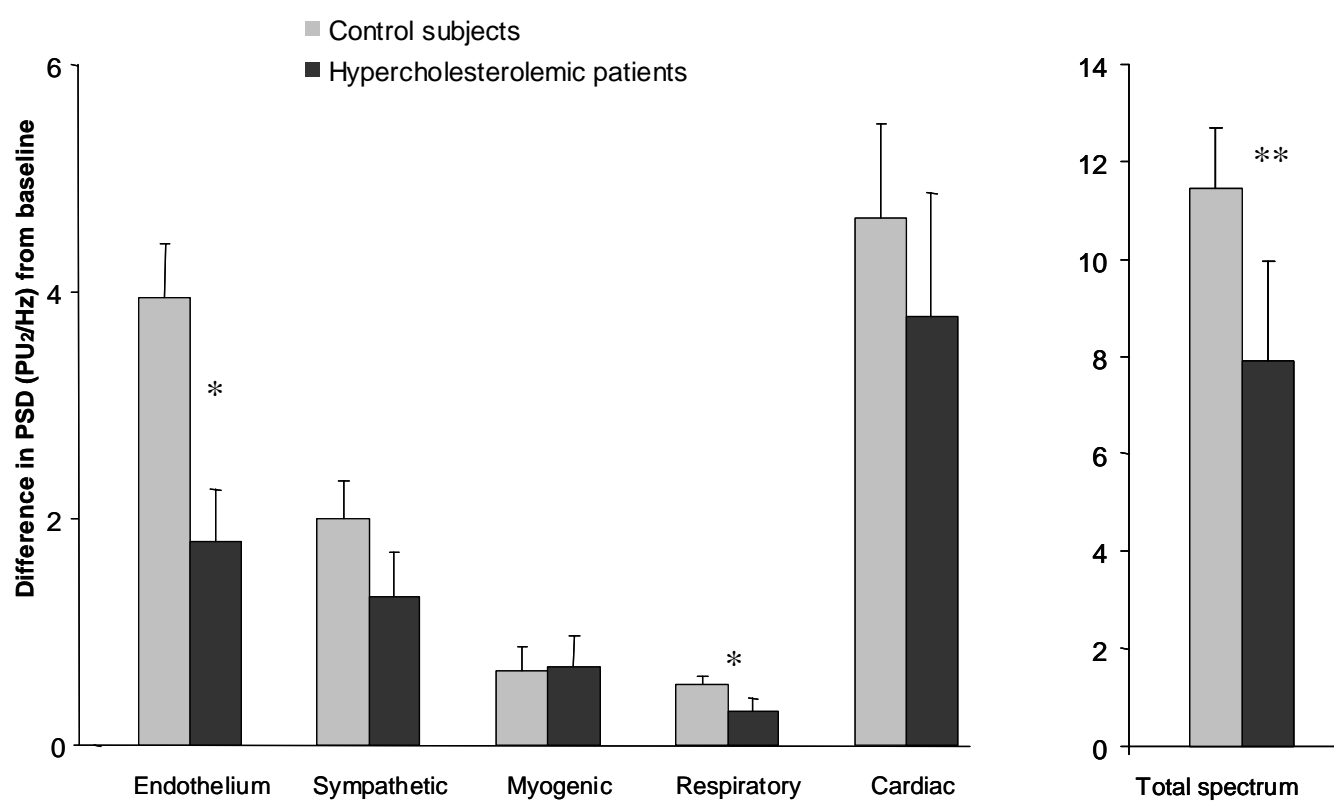


Figure 2

