Allergic sensitization increases contractile responses to 5-HT in guinea pig aorta

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Summary

Epidemiological and clinical studies suggest that asthma is associated with adverse cardiovascular outcomes, but its mechanism is uncertain. 5-Hydroxytryptamine (5-HT) is a mediator involved in asthma and in cardiovascular functioning. Thus, in the present study, we explored whether allergic sensitization in guinea pigs modifies 5-HTinduced contractile responses and 5-HT_{2A} receptor expression in thoracic aorta rings. We found that sensitization produced a significant increase of 100 µM 5-HT-induced contractions of aorta rings (~27% greater contraction than in non-sensitized animals, p<0.05). Preincubation with 10 nM ketanserin (a 5-HT_{2A} receptor antagonist) reduced by $\sim 30\%$ (p=0.003) and $\sim 36\%$ (p=0.005) the area under the curve of 5-HT-induced contractions in aortas from non-sensitized and sensitized animals, respectively. There were no differences between sensitized and non-sensitized animals with respect to mRNA (qPCR) and protein (Western blot) expression of 5-HT_{2A} receptor in thoracic aortas. We concluded that in this guinea pig model of asthma, allergic sensitization is not confined to airways, but also affects arterial contractile responses to 5-HT; changes in the expression of the 5-HT_{2A} receptor appear not to be involved in this phenomenon.

Key words: 5-HT; serotonin; 5-HT_{2A} receptor; asthma model; vascular responsiveness; thoracic aorta; vascular smooth muscle.

Introduction

Asthma is characterized by exaggerated airway contractile responses to a variety of physical or chemical stimuli. A number of inflammatory mediators have been proposed as being involved in the pathophysiology of asthma, but to date the role of 5-hydroxytryptamine (5-HT, serotonin) has been relatively poorly investigated. In animals, 5-HT induces airway smooth muscle contraction (Long *et al.* 2009), blockade of the 5-HT_{2A} receptor by ketanserin decreases allergen-induced airway hyperreactivity (De Bie *et al.* 1998), and plasma 5-HT levels are increased in sensitized animals compared with nonsensitized controls (Arreola-Ramirez *et al.* 2013). Although inhaled 5-HT proved to have weak or negligible bronchoconstrictor activity in healthy or asthmatic subjects (Tønnesen 1985), more recent studies suggest that 5-HT participates in human asthma. For example, Lechin et al. found that free plasma levels of 5-HT were increased in asthmatic patients and correlated with the severity of the disease (Lechin *et al.* 1996), and that tianeptine, a drug that potentiates 5-HT uptake, improved pulmonary function in asthmatic children (Lechin *et al.* 1998).

Some reports indicate that patients with asthma are at an increased risk for developing cardiovascular diseases (CVD) as compared with nonasthmatic subjects (Iribarren *et al.* 2004, Tattersall *et al.* 2015). In this context, the involvement of 5-HT in different physiological processes of the cardiovascular system is widely recognized, including the generation of bradycardia or tachycardia, hypotension or hypertension, and the modulation of vascular resistance with vasoconstriction or vasodilation (Saxena and Villalón 1990). These apparently opposing effects are due to the large variety of 5-HT receptors, including 14 subtypes grouped in seven families (5-HT₁₋₇) (Pytliak *et al.* 2011). The increased levels

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of 5-HT found in patients with asthma (Lechin *et al.* 1996) could partially explain the higher cardiovascular risk documented in these patients. Moreover, allergic sensitization might induce additional mechanisms that lead to the potentiation of 5-HT responses. For example, mRNA expression of 5-HT_{2A} and 5-HT_{3A} receptors in peripheral blood mononuclear cells appear to be higher in patients with asthma, as compared with control subjects (Ahangari *et al.* 2015). In contrast, mRNA expression of the 5-HT_{2A} receptor in CD4+ T-cells is reduced in patients with asthma as compared with healthy subjects (Chen *et al.* 2016). In animal models, the protein expression of 5-HT_{2A} and 5-HT_{2A} and 5-HT₄ receptors in guinea pig lung was found to be increased after systemic sensitization with ovalbumin (OVA) (Cordoba-Rodriguez *et al.* 2016).

The present work aimed to investigate modifications of the aorta contractile responses to 5-HT in a guinea pig model of OVA-induced allergic asthma, and the potential role of the 5-HT_{2A} receptor.

Male Hartley guinea pigs (500-600 g body weight) bred under conventional conditions in our institutional animal facilities were used (National Research Council (US) 2011). The protocol was approved by the Scientific and Bioethics Committees of the Instituto Nacional de Enfermedades Respiratorias (approval No. B10-13) and the Instituto Mexicano del Seguro Social (approval No. R-2013-3601-7). Experiments were performed following international guidelines for the use and care of laboratory animals.

Animals were deeply anesthetized with pentobarbital ($35 \text{ mg} \cdot \text{kg}^{-1}$, intraperitoneally) and exsanguinated. The thoracic aorta was dissected and four rings were obtained (each ring was submitted to different experimental conditions). Each aorta ring was hung in a 5-mL organ bath with Krebs solution and attached to an isometric transducer (model

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HDW100A, Biopac Systems Inc, Santa Barbara, CA, USA). Isometric transducers were connected to a preamplifier (model MP150, Biopac), signals were digitized and monitored through the Acqknowledge 3.9.1 software.

Tissues were placed under a resting tension of 1.5 g during 20 min, followed by three successive conditioning stimulations with 80 mM KCl. All responses to 5-HT were expressed as the percentage of the third KCl response. Temporal evolution of single concentrations of 5-HT (0.1, 1, 3.2, 10, 32, and 100 μ M) was evaluated in separate aorta rings. Based on these responses, subsequent experiments were carried out utilizing 100 μ M 5-HT. Some tissues were preincubated during 15 min with 10 nM ketanserin (an antagonist of the 5-HT_{2A} receptor) before adding 5-HT. Because 5-HT is a light-sensitive chemical compound, all experiments were performed under darkness conditions.

The guinea pigs were sensitized at day 0 with 60 μ g OVA and 1 mg Al(OH)₃ in 0.5 mL saline (0.9% NaCl), equally applied by the intraperitoneal and subcutaneous routes. At days 8 and 15, animals were nebulized with 3 mg·mL⁻¹ OVA in saline during 5 min, and with 1 mg·mL⁻¹ OVA in saline during 1 min, respectively. Guinea pigs were studied on days 21-25.

Total RNA was extracted from aorta tissue using TRIzol reagent (cat. 10296028, Life Technologies, Grand Island, NY, USA). Quality of RNA obtained was evaluated by resolution on denatured 1% agarose gels and measurement of 260/280 nm absorbance ratios. Synthesis of complementary DNA (cDNA) was performed by using 1 µg of total RNA, 1 µg of DNasa and 1 µL Buffer DNasa (cat. EN0523, Fermentas, Waltham, MA, USA), and reverse-transcribed (High-Capacity cDNA Reverse Transcription Kit with RNase Inhibitor, cat. 4374966, Applied Biosystems, Carlsbad, CA, USA).

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amplification was accomplished in a i-Cycler iQ Detection System (Bio-Rad Laboratories Inc., Hercules, CA, USA). Primers for the 5-HT_{2A} receptor were designed in Probefinder (http://qpcr.probefinder.com/organism.jsp) with the primer sense and antisense sequences 5'CTGCAGGATGATTCCAAGGT'3 and 3'AAATGCCACGAAAGAACCAA'5, respectively. 18S rRNA was used as an internal control gene (cat. 4352930E, Applied Biosystem). All qPCR determinations were performed in triplicate. Individual values were log-transformed to obtain a normal distribution, and the relative expression of the 5-HT_{2A} receptor, as compared with the expression of the internal control gene (18S rRNA) in the same tissue, was calculated by the $2^{-\Delta CT}$ formula. In addition, the relative expressions of the 5-HT_{2A} receptor in tissues from sensitized guinea pigs were compared with those observed in non-sensitized animals by employing the $2^{-\Delta ACT}$ method.

Western blotting analysis was performed to detect 5HT_{2A} receptor expression in aorta tissues. Proteins were extracted using lysis buffer and their concentrations were measured using the DCTM Protein Assay Kit (Bio-Rad). Proteins in a 60 µg sample were separated by polyacrylamide gel electrophoresis (SDS-PAGE), electro-transferred onto a polyvinylidene fluoride (PVDF) membrane (Bio-Rad) and blocked with 3% skim milk in TBS-T at 4°C overnight. Membranes were incubated during 120 min at room temperature with goat polyclonal antibody against the 5-HT_{2A} receptor (1:1000; sc-15073; Santa Cruz Biotechnology, Inc., Dallas, TX, USA) or with mouse monoclonal antibody against GAPDH (1:1500; sc-47724, Santa Cruz Biotechnology), the latter utilized as an internal control. The secondary antibody (1:15,000) was peroxidase horse anti-goat (PI-9500) and anti-mouse IgG (PI-200) (Vector Laboratories, Burlingame, CA, USA). Protein bands were identified by means of chemiluminescence (LuminataTM Classico Western HRP Substrate,

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Millipore Corporation, Billerica, MA, USA) and measured by densitometry employing the ImageJ software.

Contractile responses to 5-HT were evaluated in two ways: at each 2-min timeinterval, and as the whole area under the curve. After corroborating the normal distribution of data (Kolmogorov-Smirnov test), the non-paired Student's *t*-test was used to evaluate differences in responses to 5-HT between sensitized and non-sensitized tissues and, in separate experiments, to evaluate the effect of ketanserin. Statistical significance was set at one-tailed p<0.05, and the data were reported as the mean \pm standard error of the mean.

Increasing concentrations of 5-HT produced time- and concentration-dependent sustained contractions in guinea pig aortas (n=5-6 per group, data not shown). From these curves, we decided to use a 100 μ M 5-HT concentration in subsequent experiments because at this concentration we obtained the highest contractile response.

Compared with non-sensitized animals (n=7), contractile responses to 100 μ M 5-HT were increased in sensitized animals (n=5) (Fig. 1A), reaching statistically significant differences from 10 min onward. According to the area under the curve, responses in sensitized tissues represented ~27% greater contraction than in control animals (p=0.04).

In separate experiments, the effect of ketanserin was evaluated in sensitized and nonsensitized tissues. In aortas from non-sensitized guinea pigs, preincubation with 10 nM ketanserin caused a notable diminution of the 5-HT-induced contraction, which reached statistical significance from 12 min onward (n=6 per group) (Fig. 1B). Globally, this change represented ~30% diminution of the area under the curve (p=0.003). This same trend was also observed in aortas from sensitized animals, in which ketanserin caused a noticeable decrease of 5-HT-responses, especially from 14 min onward (n=6 per group, p<0.01) (Fig. 1C), and the area under the curve diminished ~36% (p=0.005).

According to the qPCR analysis by means of the $2^{-\Delta\Delta CT}$ method, the mRNA relative expression of the 5-HT_{2A} receptor in aortas from sensitized guinea pigs (n=5) corresponded to 0.296 of the relative expression in non-sensitized animals (reference value=1, n=4). However, this apparent diminution lacked statistically significant differences (p=0.74) (Fig. 2A).

There was no significant difference in the relative expression (target protein/control protein) of the 5-HT_{2A} receptor from non-sensitized guinea pigs (0.83 ± 0.07 , n=6) with respect to the sensitized group (0.95 ± 0.05 , n=7), with a p=0.17 (Fig. 2B).

In the present work, we found that allergic sensitization increases the aorta contractile responses to 5-HT. This suggests that, at least in this guinea pig model of asthma, the effect of the allergic process is not solely confined to airways or lung parenchyma, but goes beyond the latter to affect the cardiovascular system as well.

Clinical experience and epidemiological studies have shown that asthma and CVD often coincide in the same patients. Several reports revealed that patients with asthma had an increased prevalence of CVD (Bellocchia *et al.* 2013), with a 32% to 60% higher risk for CVD occurrence when compared with persons without asthma (Iribarren *et al.* 2012, Tattersall *et al.* 2015, Tattersall *et al.* 2016, Wang *et al.* 2017). Although it has been argued that the association of asthma and CVD may be simply explained because both conditions share the same risk factors, it is also possible that a cause-effect relationship exists in this association. A potential mediator involved in this link is 5-HT, inasmuch as this mediator appears to be involved in the pathogenetic processes of both asthma and CVD. Thus,

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clinical studies have demonstrated that plasma levels of 5-HT were elevated in a variety of CVD-related pathologies, including hypertension, coronary artery disease, valvular heart disease, atherothrombosis, aterosclerosis, and myocardial infarction (Kaumann and Levy 2006). Likewise, increased 5-HT blood levels were found in patients with asthma (Lechin *et al.* 1996), as well as in animal models of asthma (Segura *et al.* 2010, Arreola-Ramirez *et al.* 2013). Moreover, platelets constitute the major source of 5-HT in blood, and it is known that, in humans, these elements are involved in disorders of the cardiovascular system (Papapanagiotou *et al.* 2016), as well as in asthma (Idzko *et al.* 2015).]

A number of studies support the concept that 5-HT_{2A} receptors mediate 5-HTinduced vascular smooth muscle contraction in arterial blood vessels (McKune and Watts 2001). In this context, in the present work, our first approach comprised assessing whether allergic sensitization causes an increase in 5-HT_{2A} receptor expression. We did not find statistically significant modifications in the amount of mRNA or protein expressions. Some alternative mechanisms for explaining these increased responses might be speculated upon. sensitization might enhance intracellular signaling mechanisms, Allergic for example, through the generation of reactive oxygen species (ROS), leading to enhanced responsiveness to 5-HT (Rubanyi and Vanhoutte 1986, Lounsbury et al. 2000, Zuo et al. 2013, Qu et al. 2017). On the other hand, although the main mechanism by which 5-HT produces vascular smooth muscle contraction is stimulation of the 5- HT_{2A} receptor and activation of Gq proteins (Misarkova *et al.* 2016), this response might be modified by the simultaneous effect of 5-HT on other 5-HT receptors, such as the pre-junctional 5-HT₃, 5-HT₄, and 5-HT₇ receptors (Segura *et al.* 2010). Thus, sensitization could be changing

expression or functionality of these latter receptors, instead of the expression of the 5-HT_{2A} receptor.

We found that ketanserin only inhibited a relatively moderate portion of the response, mainly in its second half. This result clearly indicates that factors other than 5- HT_{2A} receptor activation were responsible for the major part of the serotoninergic contractile response. Notably, the effect of ketanserin was relatively the same in sensitized and in non-sensitized animals, indicating that the $5HT_{2A}$ receptor-independent contractile response to 5-HT was not affected by allergic sensitization.

In conclusion, our results revealed that in this asthma model, allergic sensitization was not confined to airways, but it also affected arterial responses to 5-HT, given rise to increasing contractile responses. Modifications of 5-HT_{2A} receptor expression appear not to be involved in this phenomenon.

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Conflict of interest statement

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Legends for figures

Fig. 1. Modifications of the contractile responses to 5-HT by allergic sensitization and ketanserin in guinea pig thoracic aorta rings. Allergic sensitization to ovalbumin caused increased contractile responses to 100 μ M 5-HT (**A**). Preincubation with the 5-HT_{2A} receptor antagonist ketanserin (KT, 10 nM) diminished the contractile response to 100 μ M 5-HT in either non-sensitized (**B**) and sensitized (**C**) tissues. Symbols correspond to the mean \pm standard error of n=5-7 experiments per group. *p < 0.05 and †p < 0.01 (Student's *t*-test).

Fig. 2. Effect of allergic sensitization to ovalbumin on the mRNA (**A**) and protein (**B**) expression of the 5-HT_{2A} receptor in guinea pig thoracic aortas. Bars and vertical lines correspond to the geometric mean \pm standard error, respectively, of n=4-6 (non-sensitized) or n=5-7 (sensitized) experiments. There was no statistically significant difference between non-sensitized and sensitized animals.



